

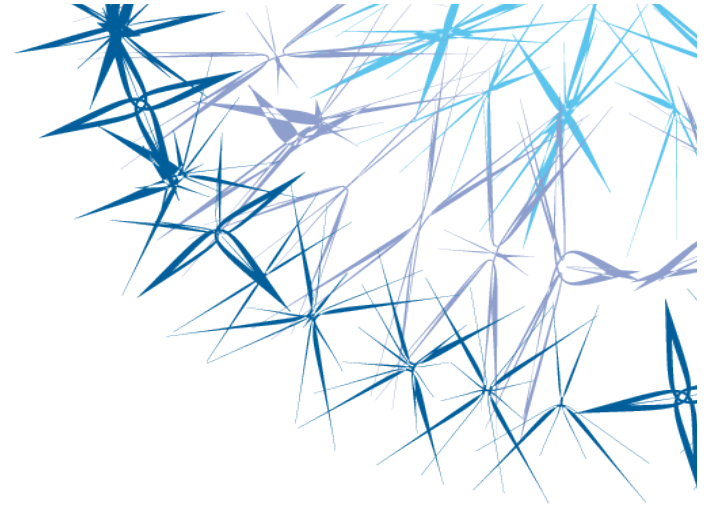


COSBI

Programming-based systems biology

Corrado Priami





outline



Biology needs a theory able to highlight causality and abstract data into knowledge to elucidate the architecture of biological complexity.

Sidney Brenner

The grand challenge for all scientific and engineering disciplines in the 21st century is complexity.

Lee Hood

IMPACT CORE COMPUTER SCIENCE AND SYSTEMS BIOLOGY

Quantitative operational descriptions
of the mechanistic behavior of
biological systems

Computational thinking

Not ambiguous specifications for
simulation and analysis tools

Coping with combinatorial explosion
of systems description

**MOVING BEYOND MATH MODELING:
ADDRESS CONCURRENCY AND
COMPLEXITY**



BIOINFORMATICS IS DIFFERENT

Comparison of strings

Storage of experiment results

Visualization of complex data

Search and analysis of data sets...

Mainly Structural/Static descriptions

MATHEMATICAL AND COMPUTATIONAL BIOLOGY ARE DIFFERENT

Static abstract relationships

Computer assisted solutions

Global pictures of dynamics

OUR DRIVING CHALLENGES

Interaction

Emergence

Partial knowledge

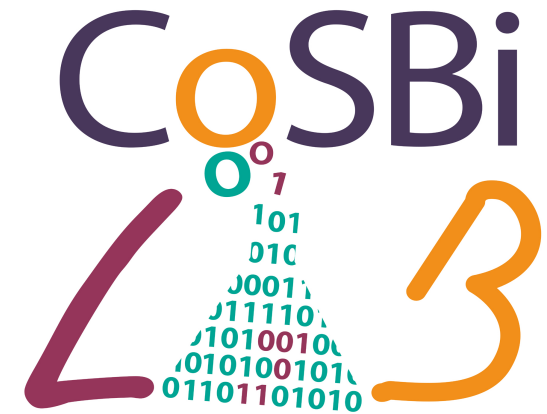
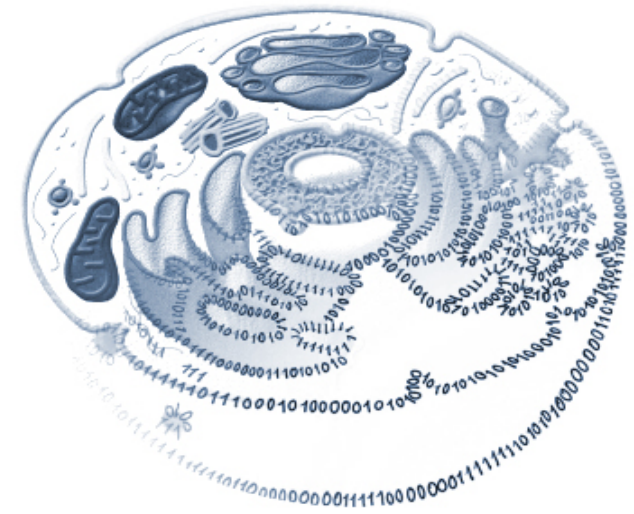
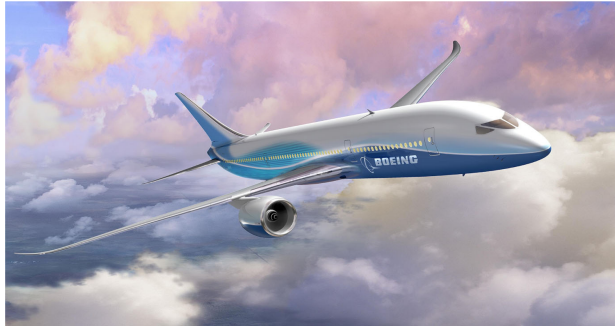
Ambiguous observations

Multi-level, multi-scale in space and time

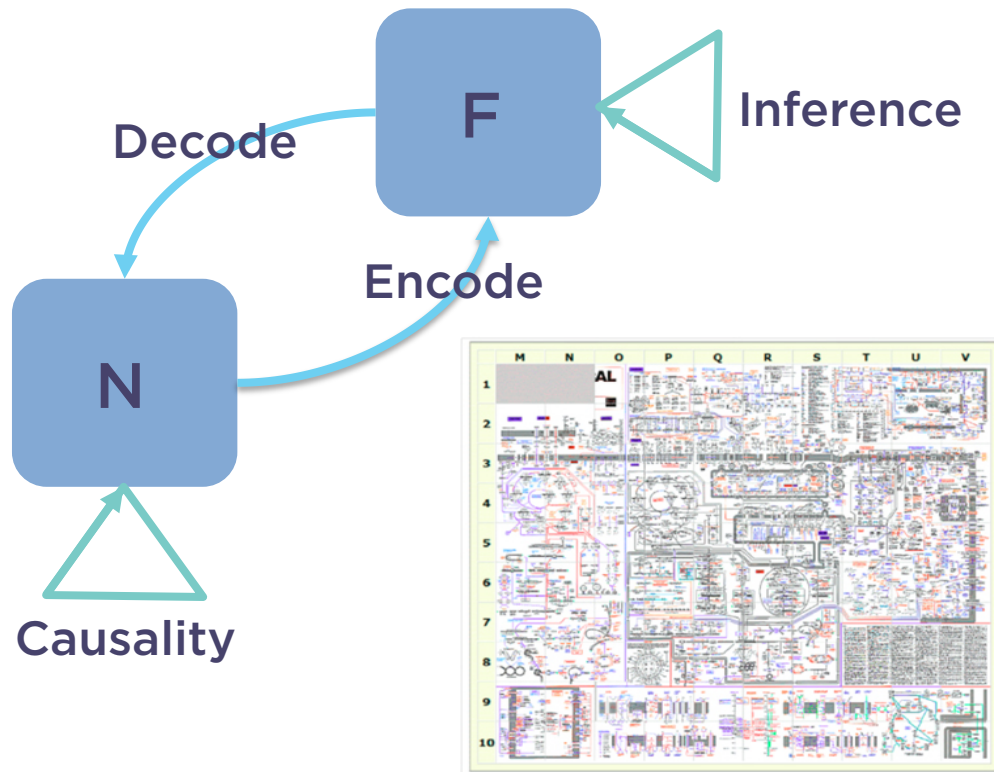
Causal relationships and context-awareness

LOW-LEVEL LOCAL MECHANISMS AFFECT HIGH-LEVEL GLOBAL BEHAVIOR

SIMULATION-BASED, DATA-DRIVEN SCIENCE



A MODELING FORMALISM



Encode info manipulation by bio-systems

A formal framework to reason about bio-systems

Predict biological behavior and identify new hypotheses

Unambiguous description to share knowledge

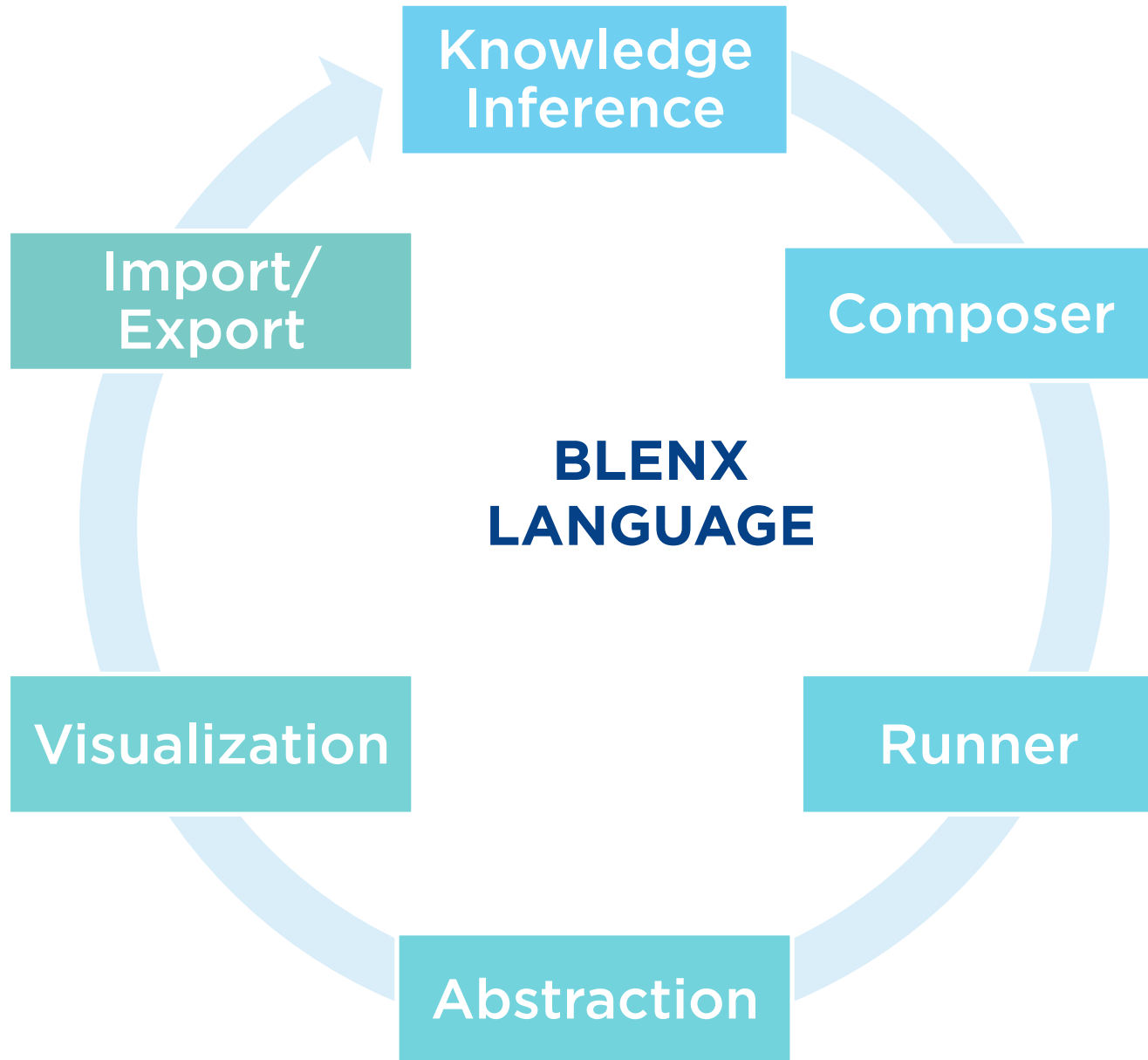
the development of the appropriate languages to describe information processing in biological systems and the generation of more effective methods to translate biochemical descriptions into the functioning of the logic circuits that underpin biological phenomena

Paul Nurse

THE METAPHOR

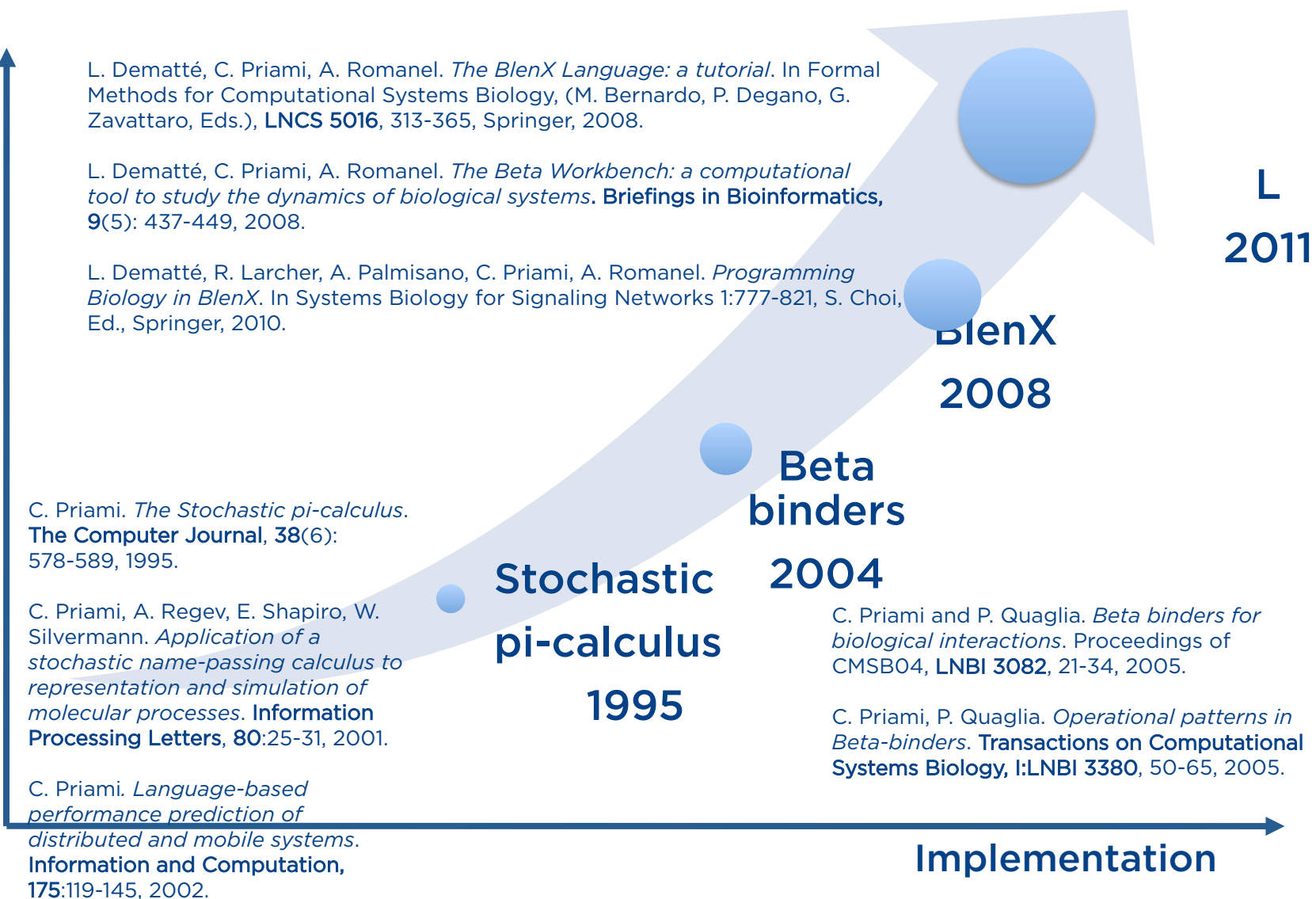
biological entities	processes
interaction capabilities	interfaces
complex/decomplex	binding/unbinding
dynamics	state change

THE ENVIRONMENT



BLENX GENESIS

Case
Studies



PI-CALCULUS

Processes

- Domains, Molecules/Proteins, Systems

Primitives

- Join/Split, Conditional Events
- Restriction for Compartments, Membranes, Complexes

Complex/
Decomplex

- Not explicitly modeled
- Interplay between private names and scope size

Low-level
programming

- Reversibility of interactions

Modeling

- Emergent behavior must be programmed

Interaction

- Key-lock mechanism

Implementation

- General structural congruence

BETA-BINDERS

Processes

- Domains, Molecules/Proteins, Systems

Primitives

- Join/Split, Conditional Events
- Restriction for Compartments, Membranes, Complexes

Complex/ Decomplex

- Not explicitly modeled
- Interplay between private names and scope size

Low-level programming

- Reversibility of interactions

Modeling

- Emergent behavior must be programmed

Interaction

- Key-lock mechanism

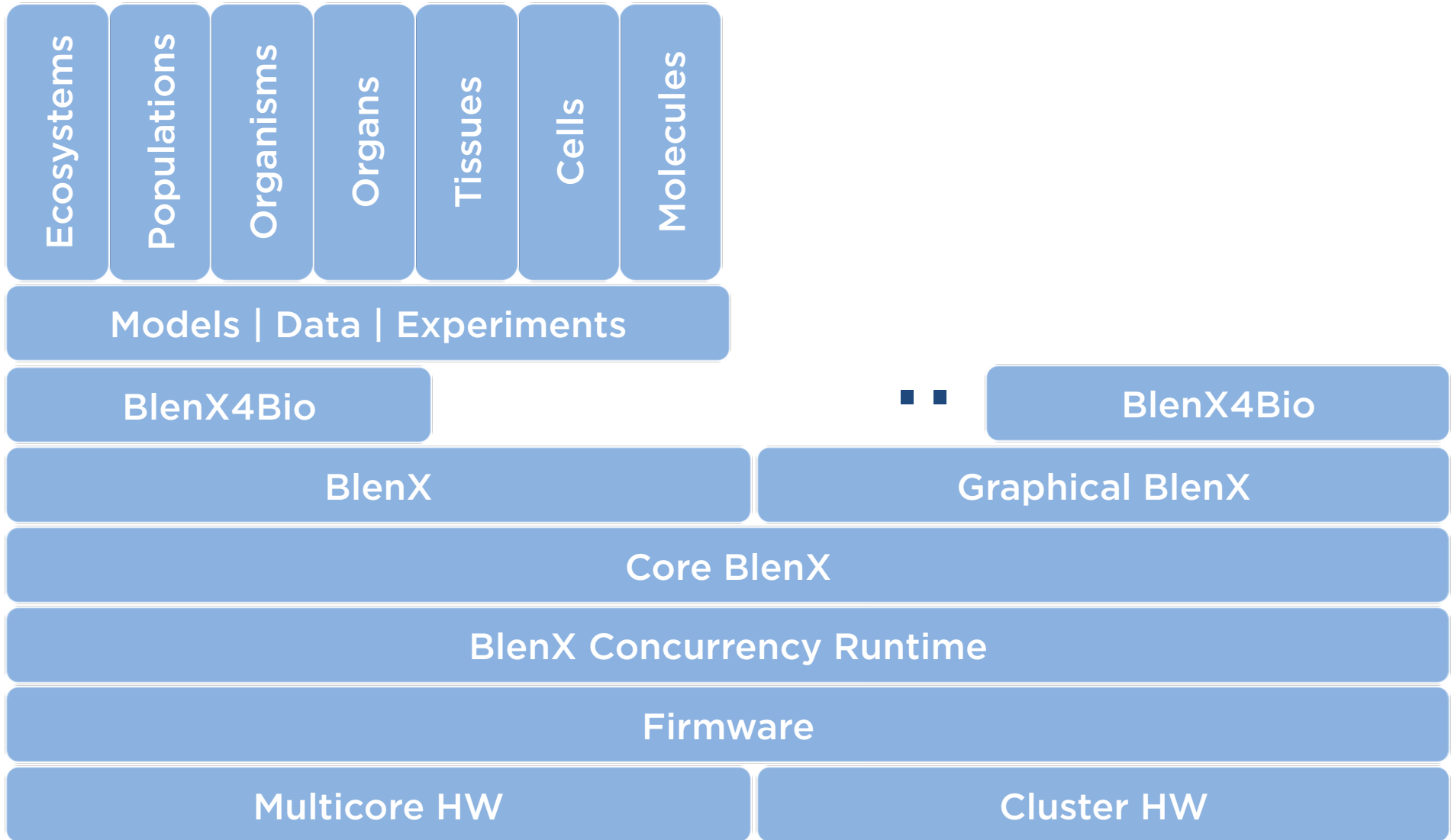
Implementation

- General structural congruence

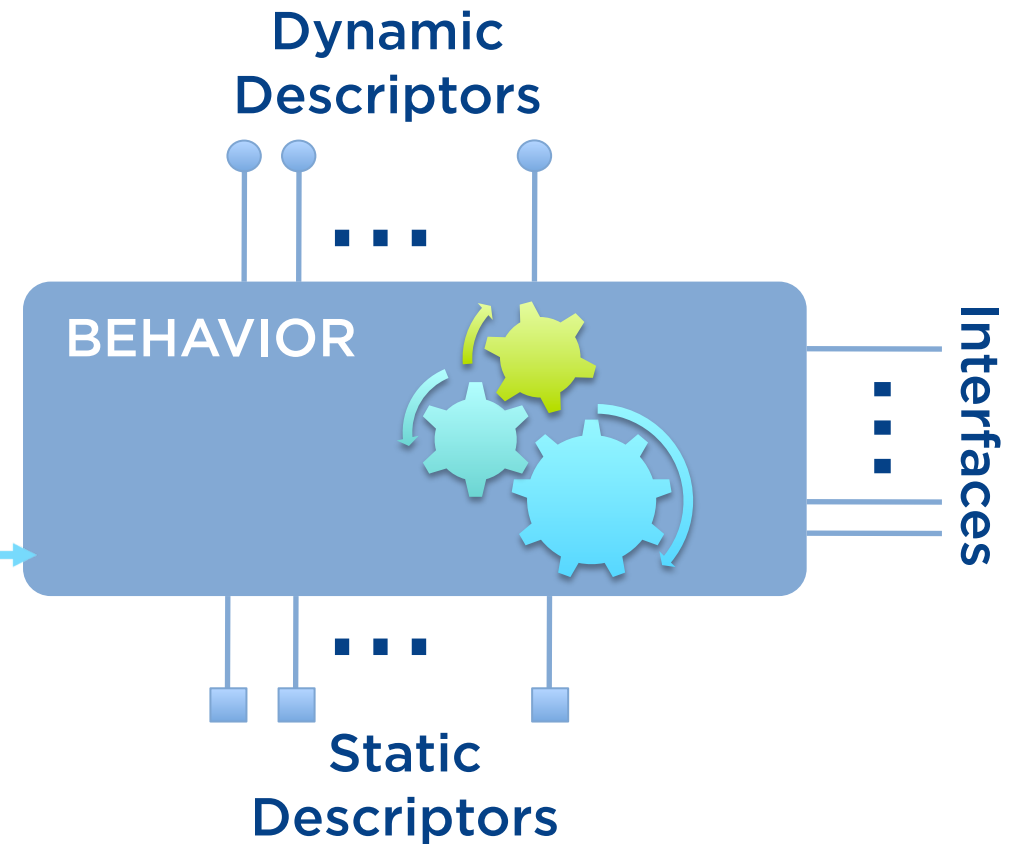
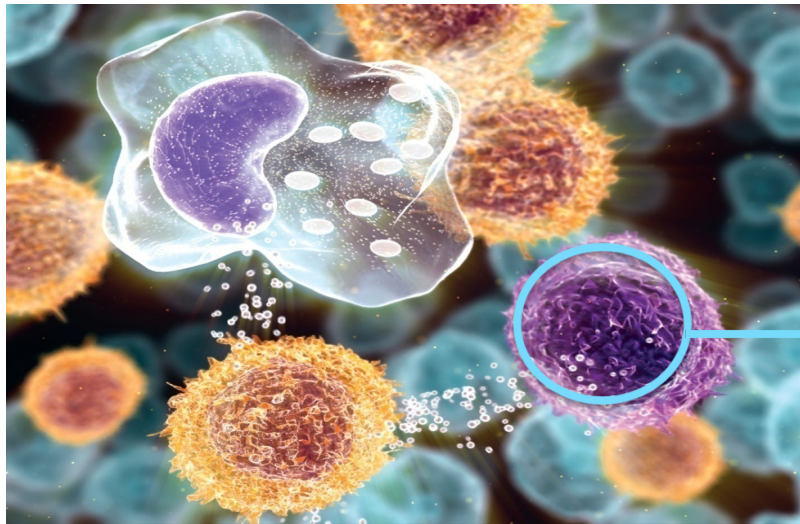
BLENX PRINCIPLES

- Set of interacting boxes with internal behavior
- Non key-lock interaction mechanism
- Complex description and their Dynamic generation
- Typed and dynamic varying interfaces
- 1-1 correspondence with Biological elements
- Events

BLENX TOWER



BLENX CORE ELEMENTS



C. Priami, P. Quaglia and A. Romanel. *BlenX - static and dynamic semantics*. Proceedings of CONCUR09, LNCS 5710, 37-52, Springer, 2009.

A. Romanel. Dynamic Biological Modelling: a language-based approach. PhD Thesis. COSBI TR PhD-1-2010.

BLENX PROGRAMMING LANGUAGE

```
// CDP
let CDP_ : bproc = #(y,CDP) [ rep start?().Prog_CDP | Prog_CDP ];
let dCDP_ : bproc = #(y,dCDP) [ rep start?().Prog_CDP | Prog_CDP ];
let dCTP_ : bproc = #(y,dCTP) [ rep start?().Prog_CDP | Prog_CDP ];

// RR
let RR_ : bproc = #(r,RR) [ rep start?().Prog_RR | Prog_RR ];

// CELL
// f1
let Cell : bproc = #(cell,Alive) [ nil ];
when ( Cell :: cell_death_function ) delete(1);
run 100 CDP_ || 100 dFdC_ || 1000 CDA_ || 1000 RR_ || 100 Cell || 100 DNA_healthy
```

LANGUAGE FLAVOR: BLENX

```
// Tree.prog

[ steps = 10, delta = 10 ]

<< BASERATE:inf, HIDE:inf, UNHIDE:inf, CHANGE:inf >>

// Initiator Definition
let Initiator : bproc = #(out,I) [ out?().out!(root).nil ];

// Node Definition
let p1 : pproc = !out1?().out2?().out1!(node).out2!(node).nil ;
let p2 : pproc = !out1?().out2?().inp!().inp?(m).out1!(m).out!(m).nil ;
let nodeP : pproc =
  inp!().inp?(t).( t!() | (
    node?().unhide(out1).unhide(out2).p2 +
    root?().unhide(out1).unhide(out2).p1
  ) );

let Node : bproc = #(inp,IN),#h(out1,ONE),#h(out2,TWO)
  [ nodeP ];

// Init
run 2 Initiator || 10 Node
```

```
// Tree.types

{ I, IN, ONE, TWO }
%%
{
  (I,IN,100,0,inf),
  (ONE,IN,1,0,inf),
  (TWO,IN,1,0,inf)
}
```

a static list of the entities of
the initial configuration

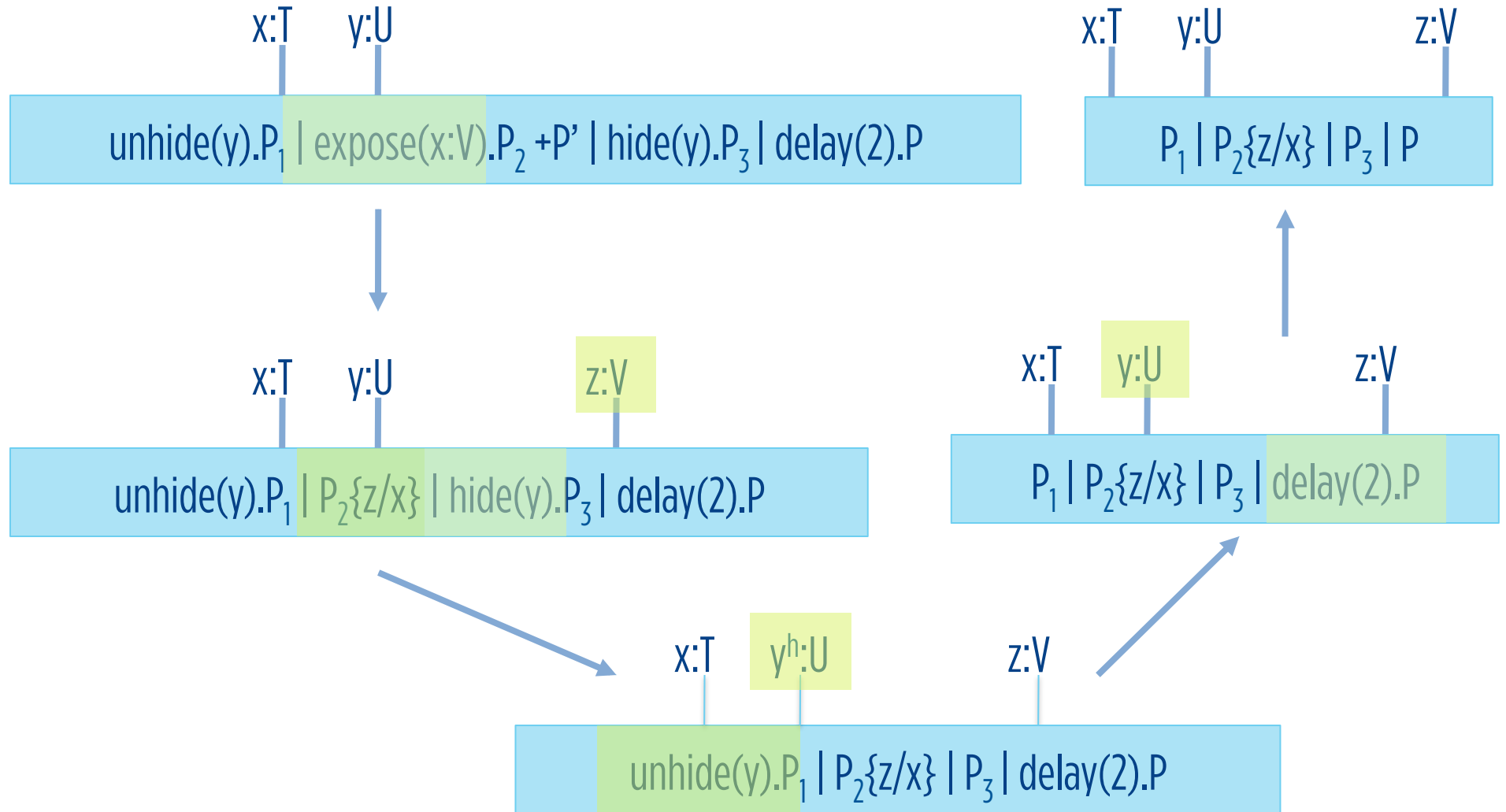
+

their amount and specificity
for interaction.

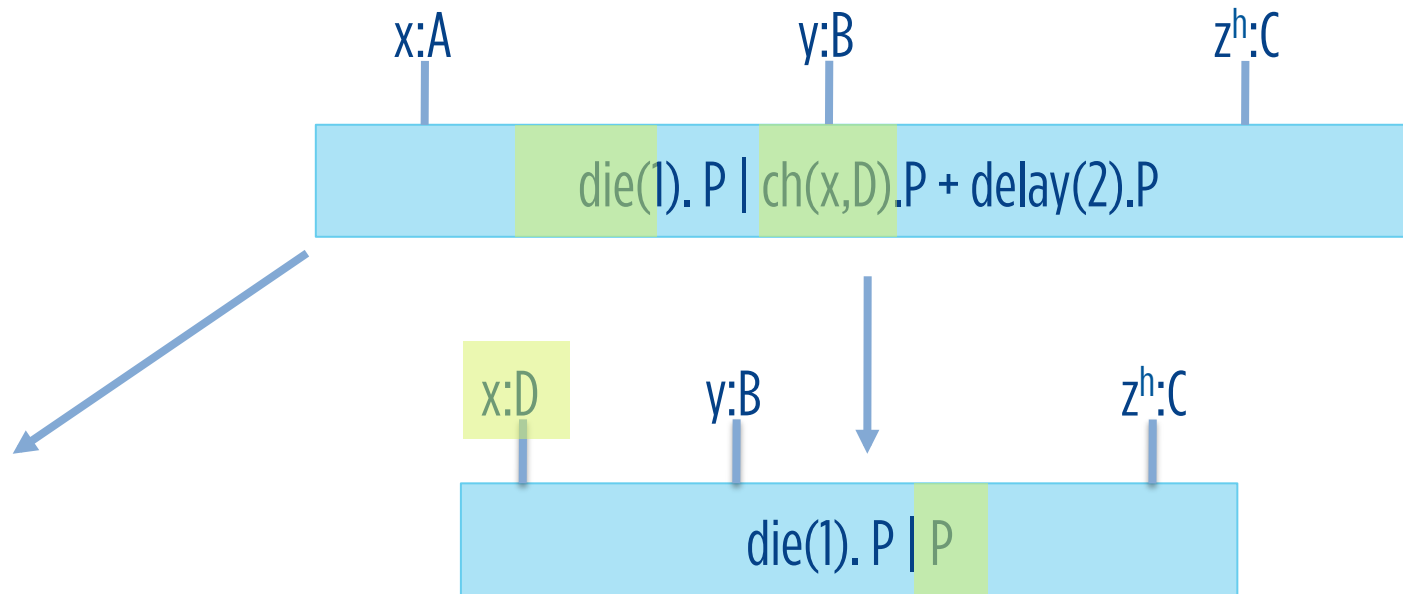
ENABLING LIBRARY-BASED MODELING

Further examples and downloads: <http://www.cosbi.eu/index.php/research/prototypes/beta-wb>

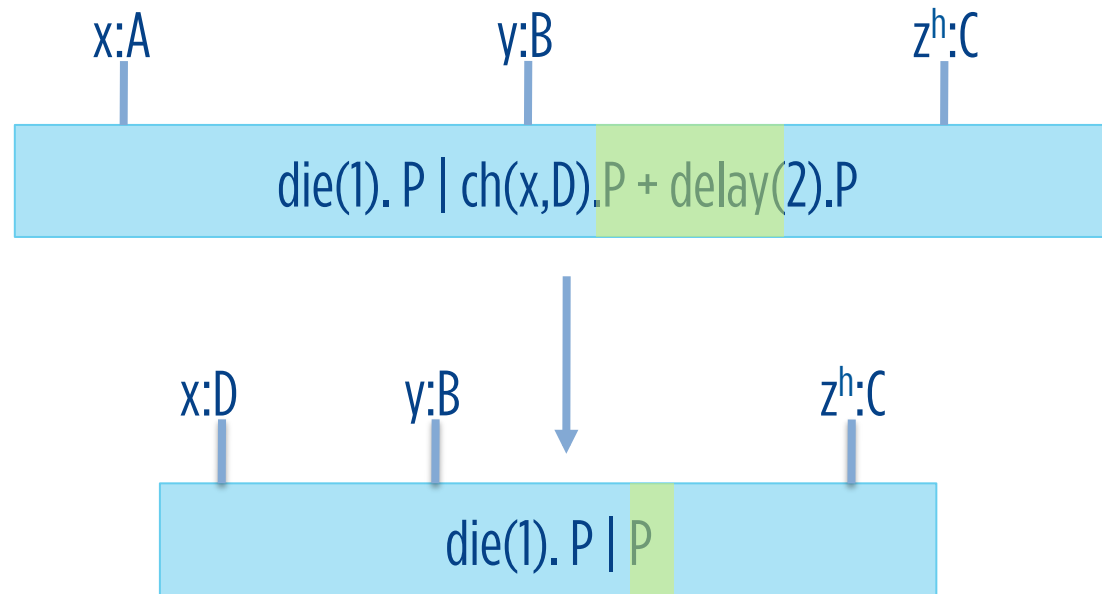
BLENX: MONOMOLECULAR ACTIONS



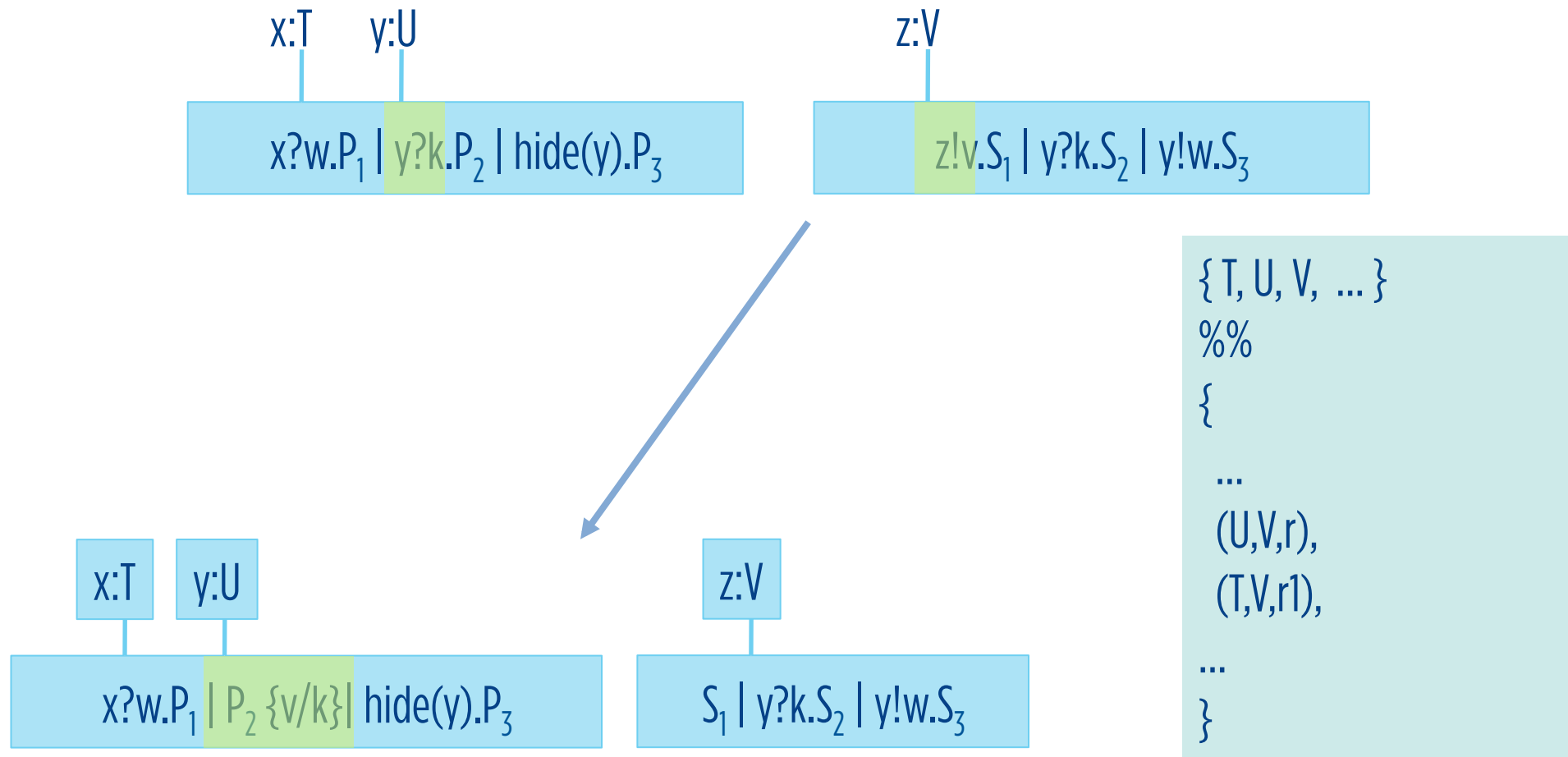
BLENX: MONOMOLECULAR ACTIONS



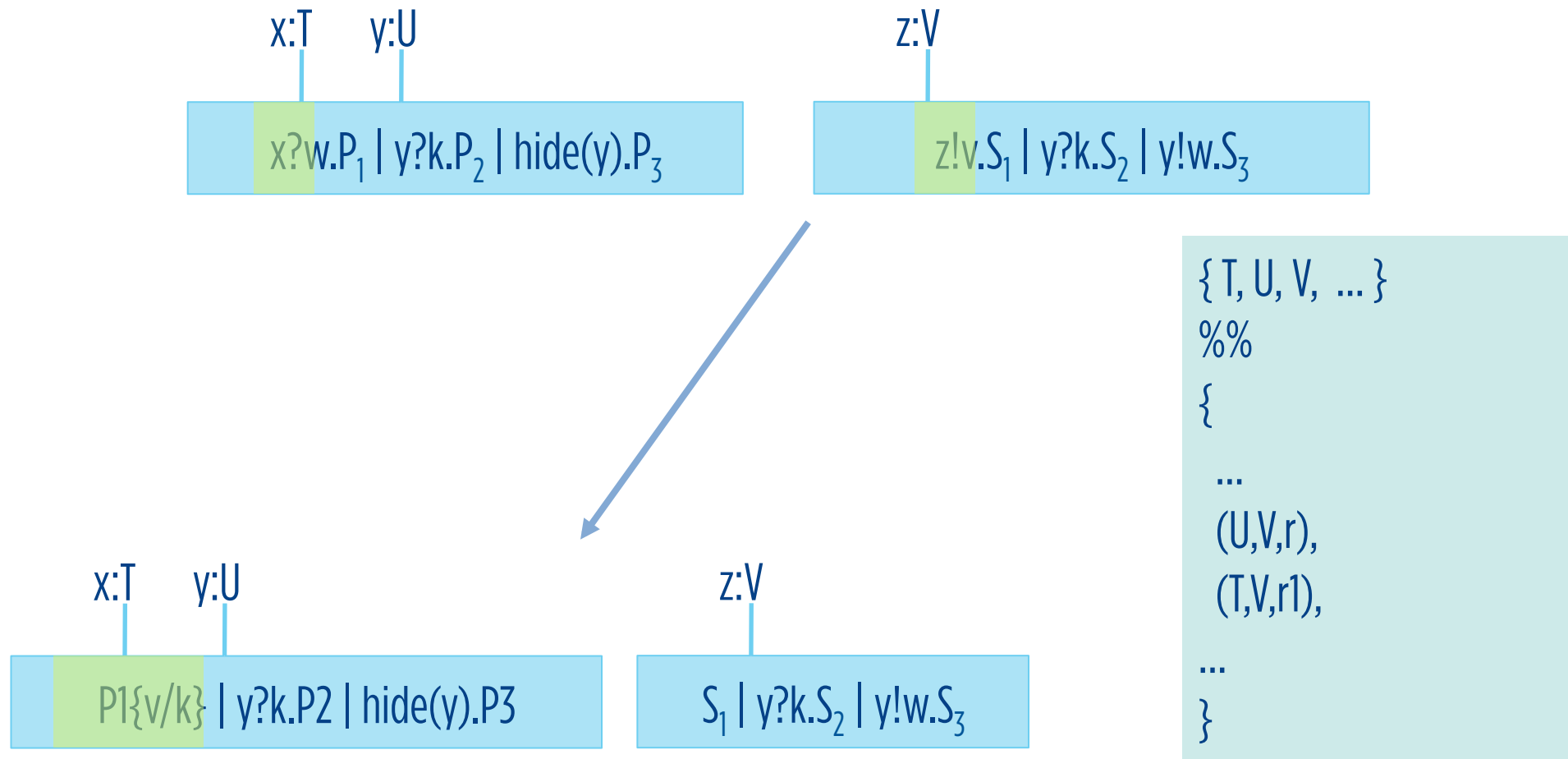
BLENX: MONOMOLECULAR ACTIONS



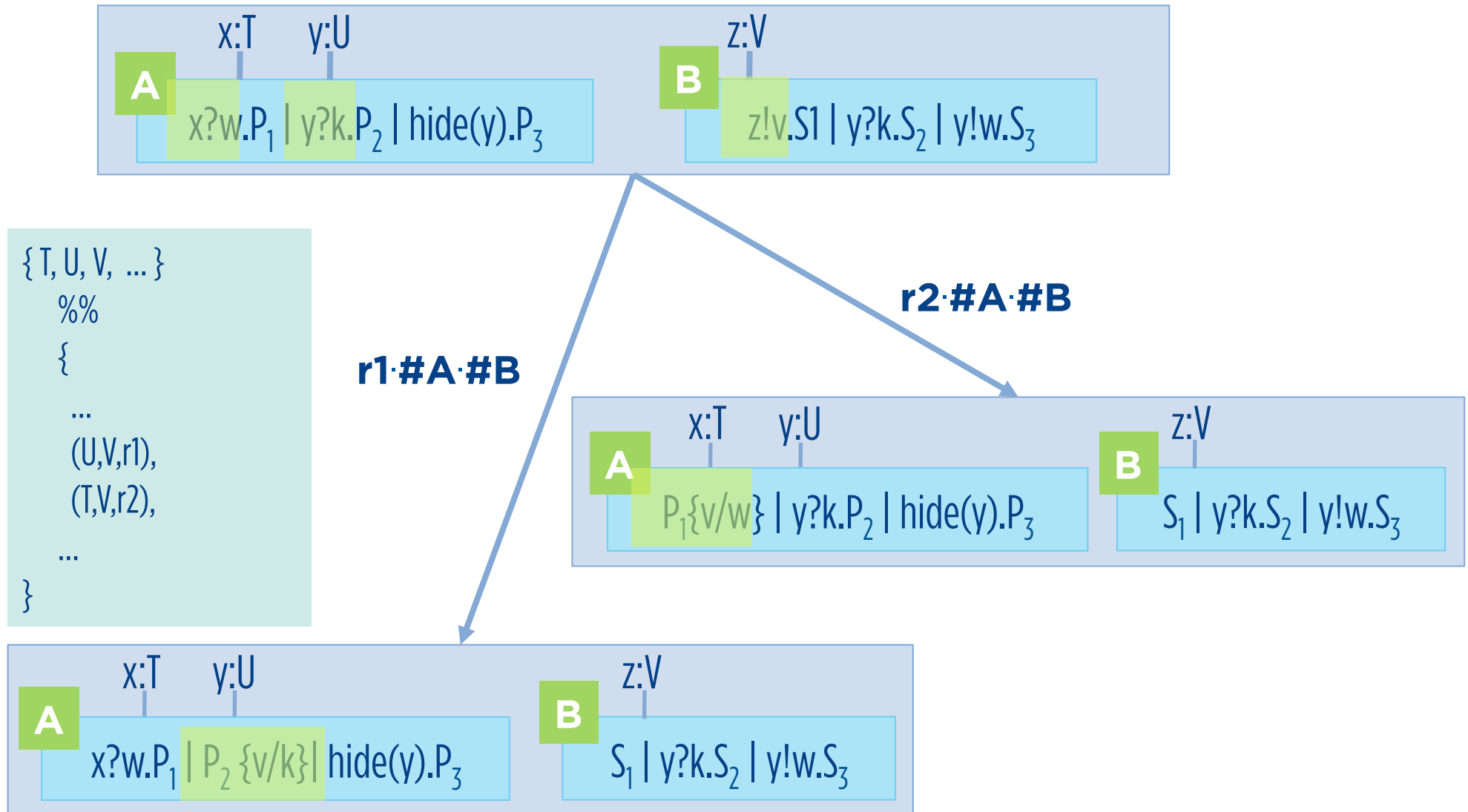
BLENX: BIMOLECULAR ACTIONS



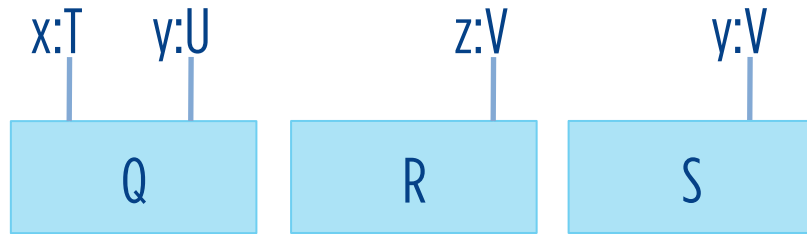
BLENX: BIMOLECULAR ACTIONS



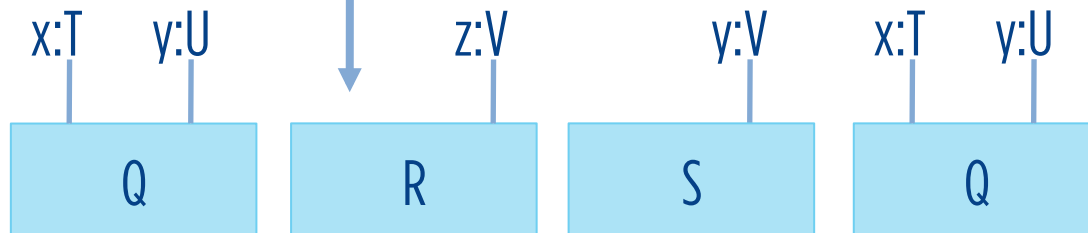
BLENX: BIMOLECULAR ACTIONS



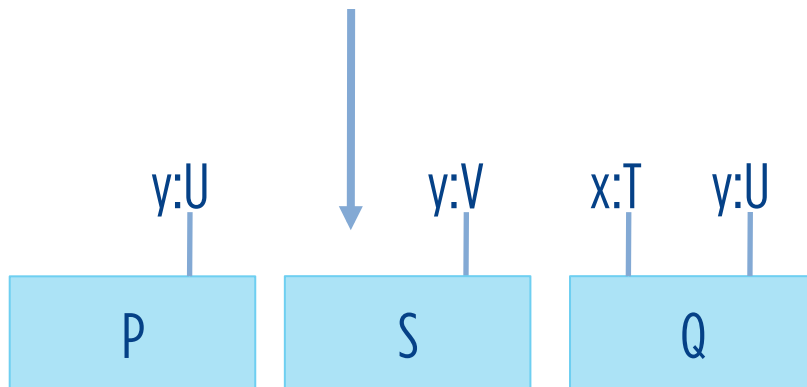
BLENX: EVENTS



when (Q :: 10) new (1);

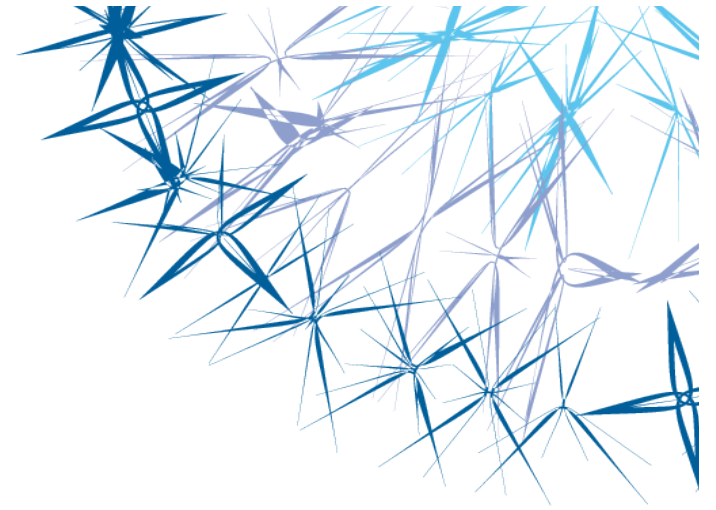


when (R , Q :: f) join (P);
let f = |S|*sqrt(|Q|)/k

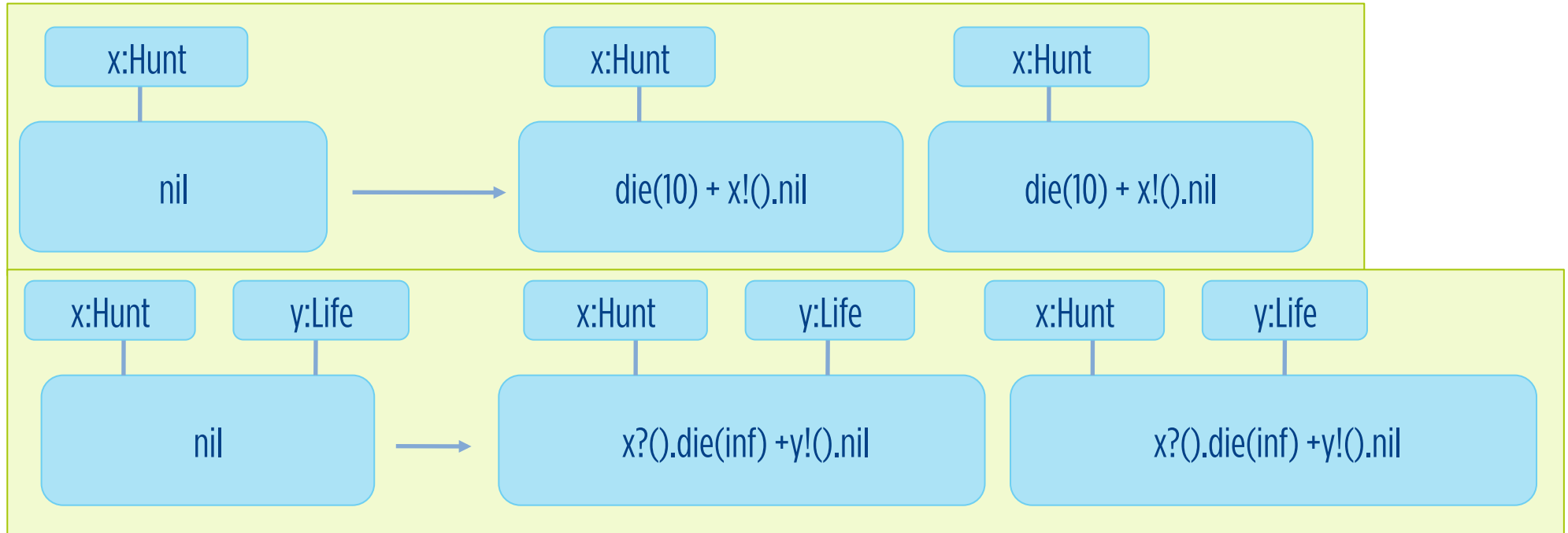


Example

Predator - Prey



PREDATOR - PREY

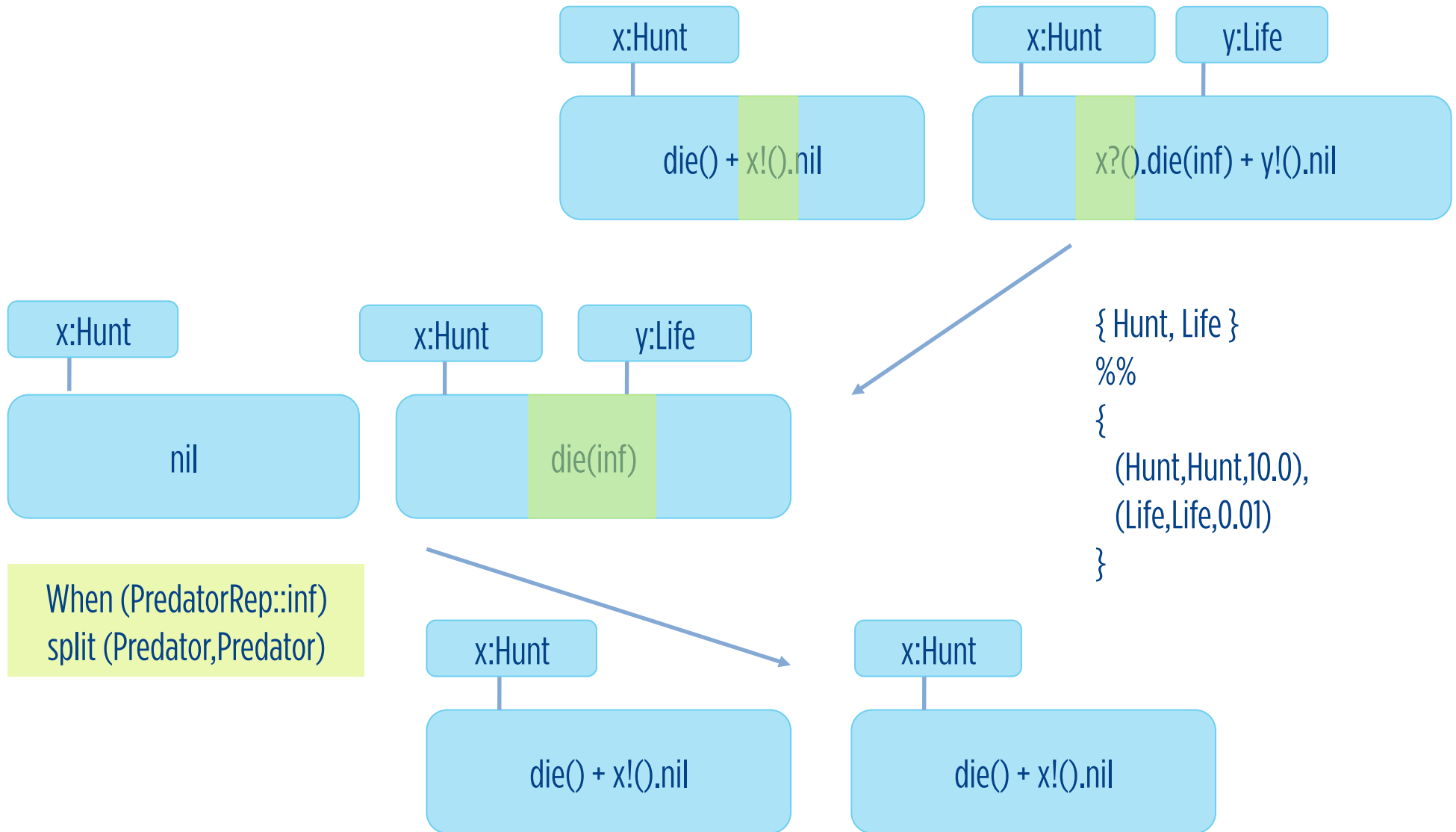


```
let PredatorRep : bproc = #(x,Hunt) [nil ];
when (PredatorRep :: inf) split (Predator,Predator);
```

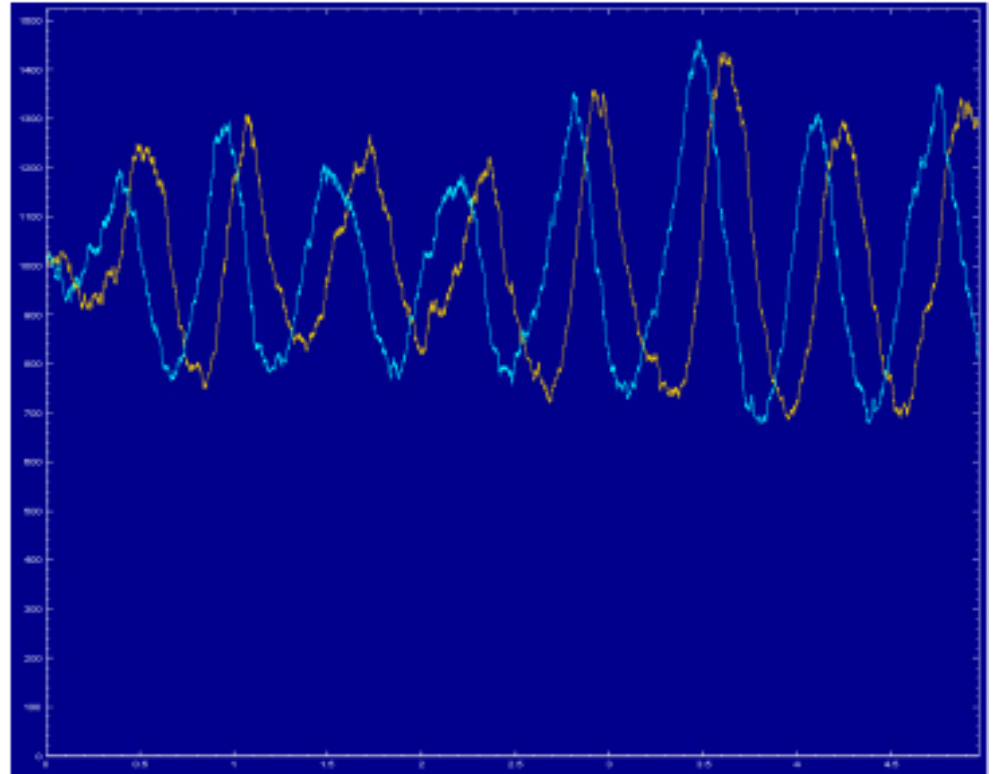
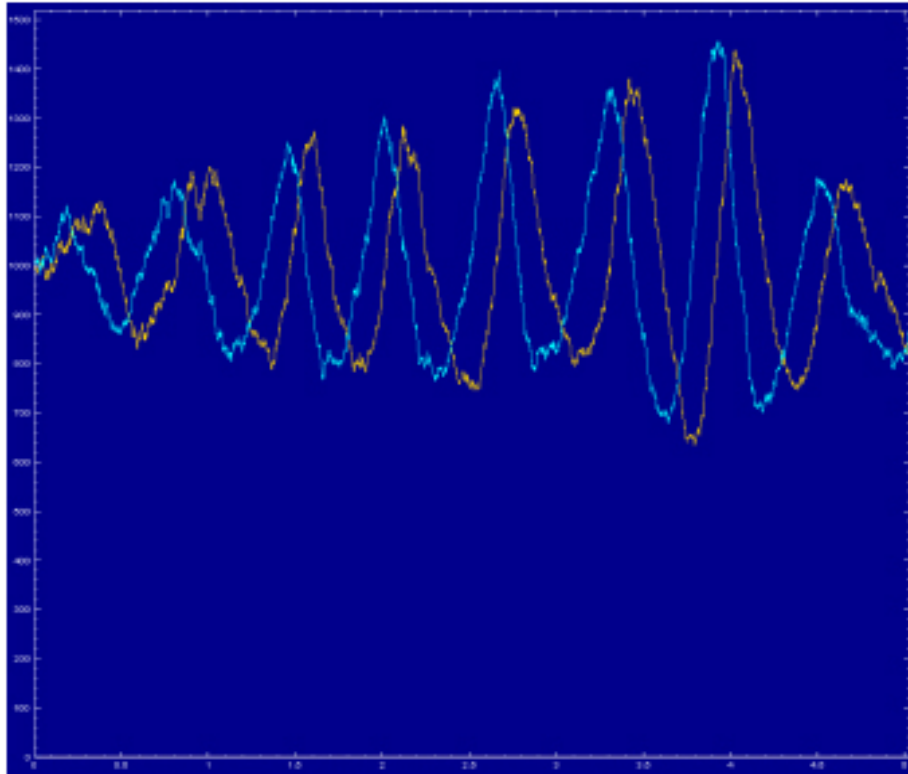
```
let PreyRep : bproc = #(x,Hunt),#(y,Life) [nil ];
when (PreyRep :: inf) split (Prey,Prey) ;
```

run 1000 Predator || 1000 Prey || 1 Nature

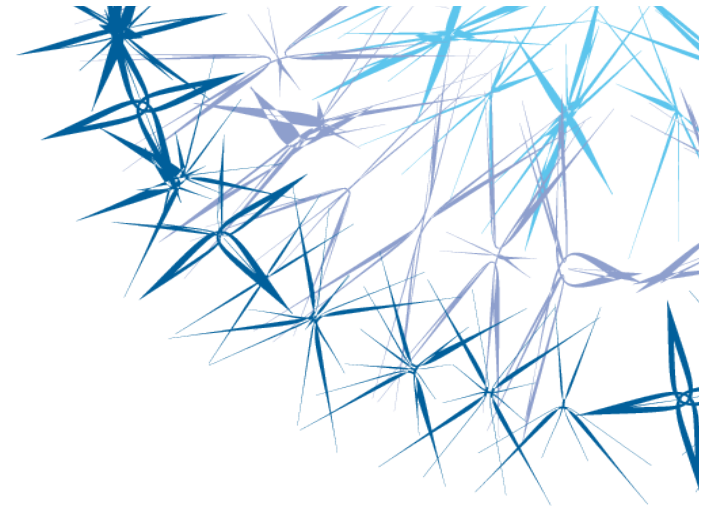
PREDATOR - PREY



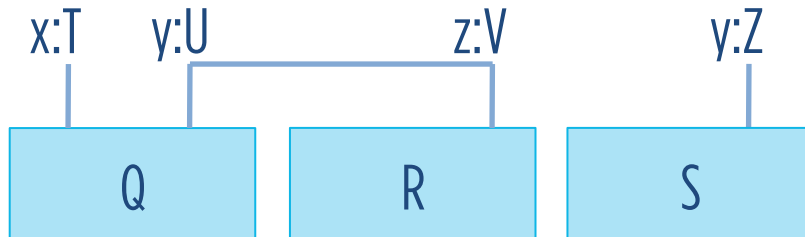
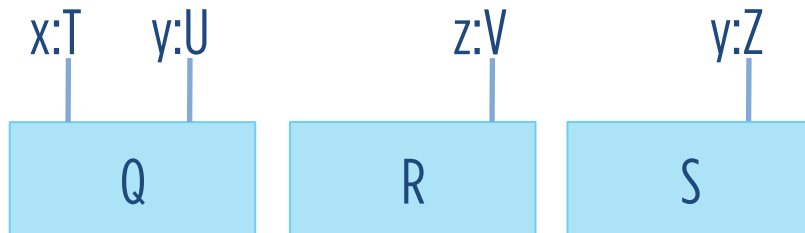
PREDATOR - PREY



END Example
Predator - Prey



BLENX: COMPLEXES



{T, U, V, Z, ... }

%%

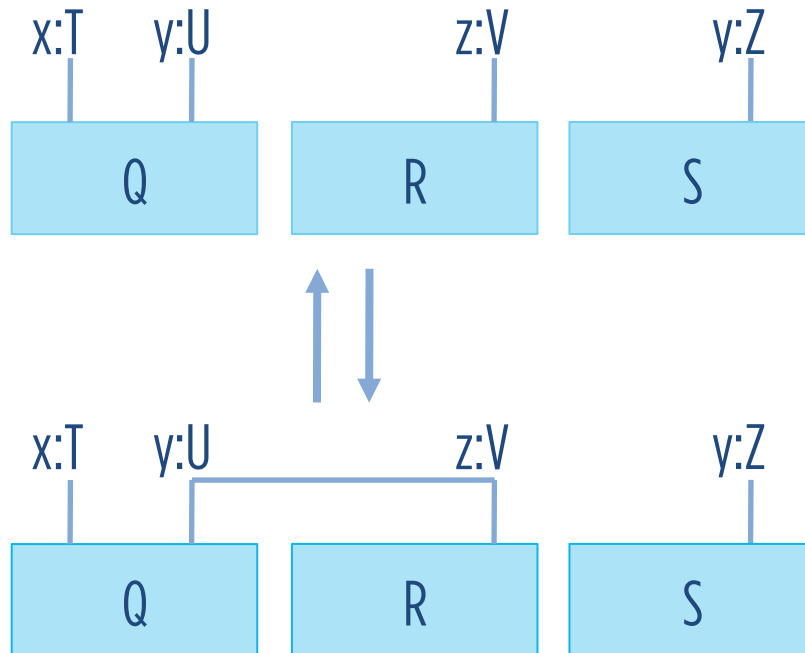
{

(U, Z, 3.5),

(U, V, 1.5)

}

BLENX: COMPLEXES



{T, U, V, Z, ... }

%%

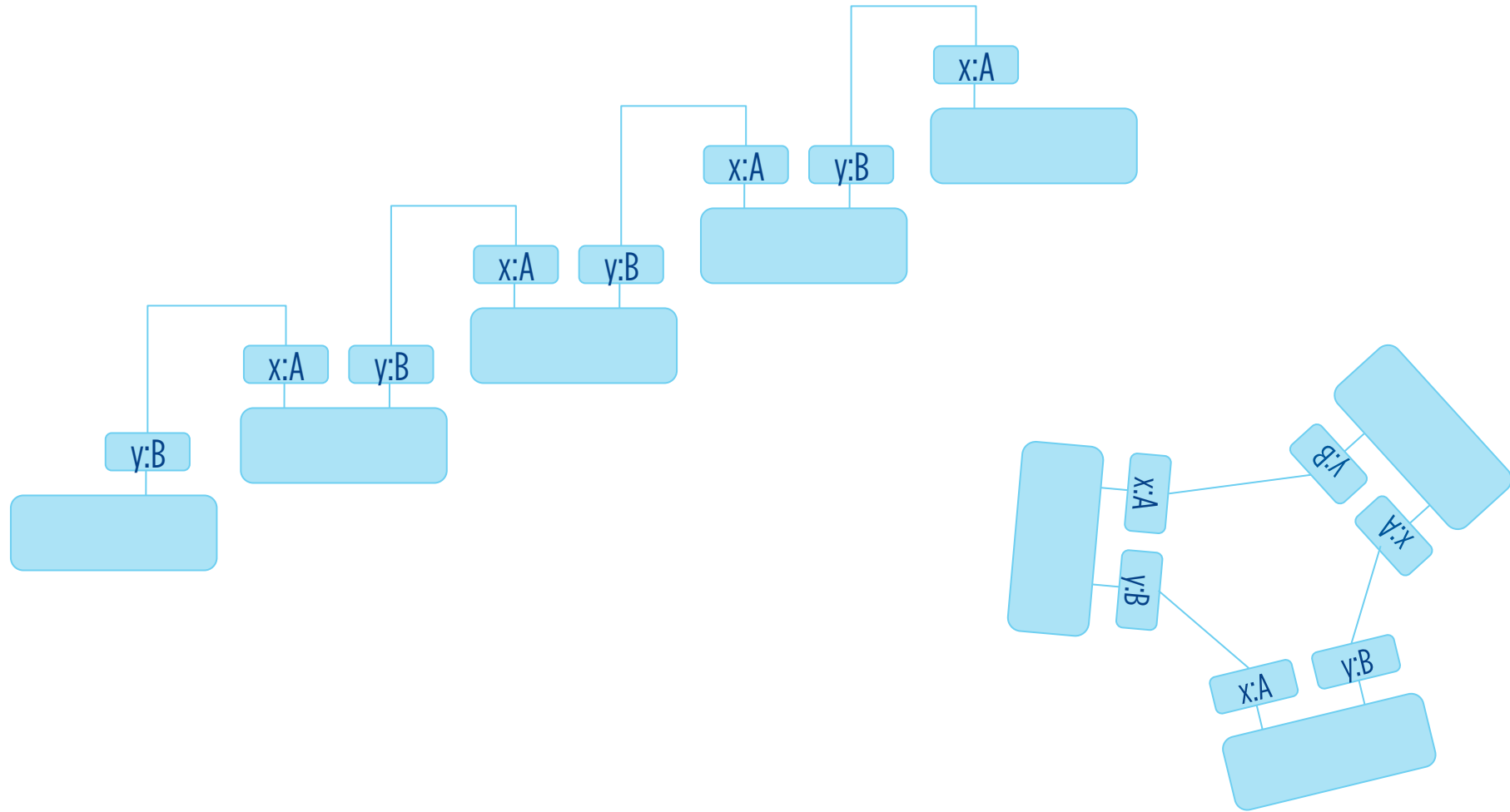
{

(U, Z, 3.5),

(U, V, 1.5, **2.5, 10**)

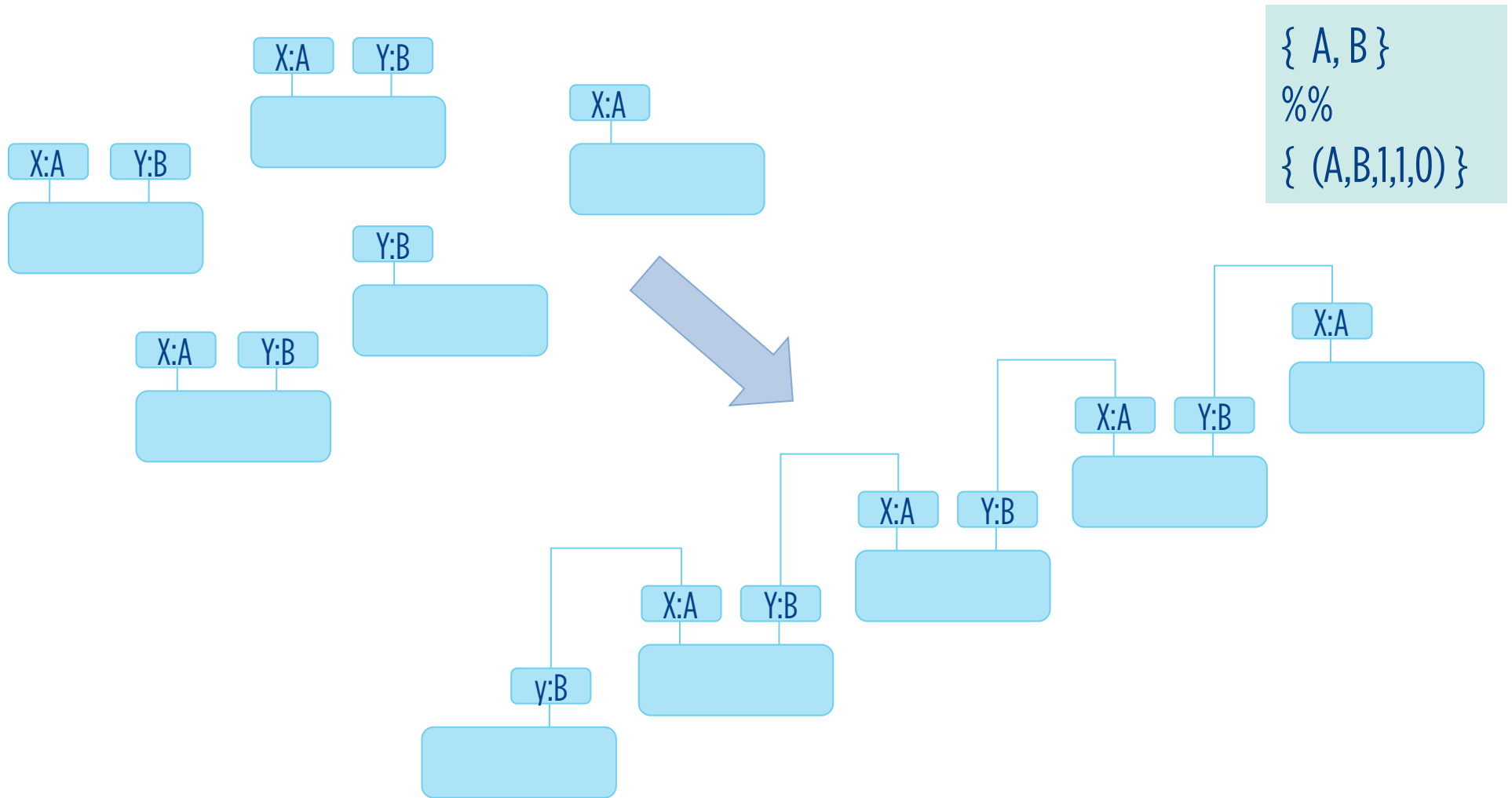
}

BLENX: STRUCTURES

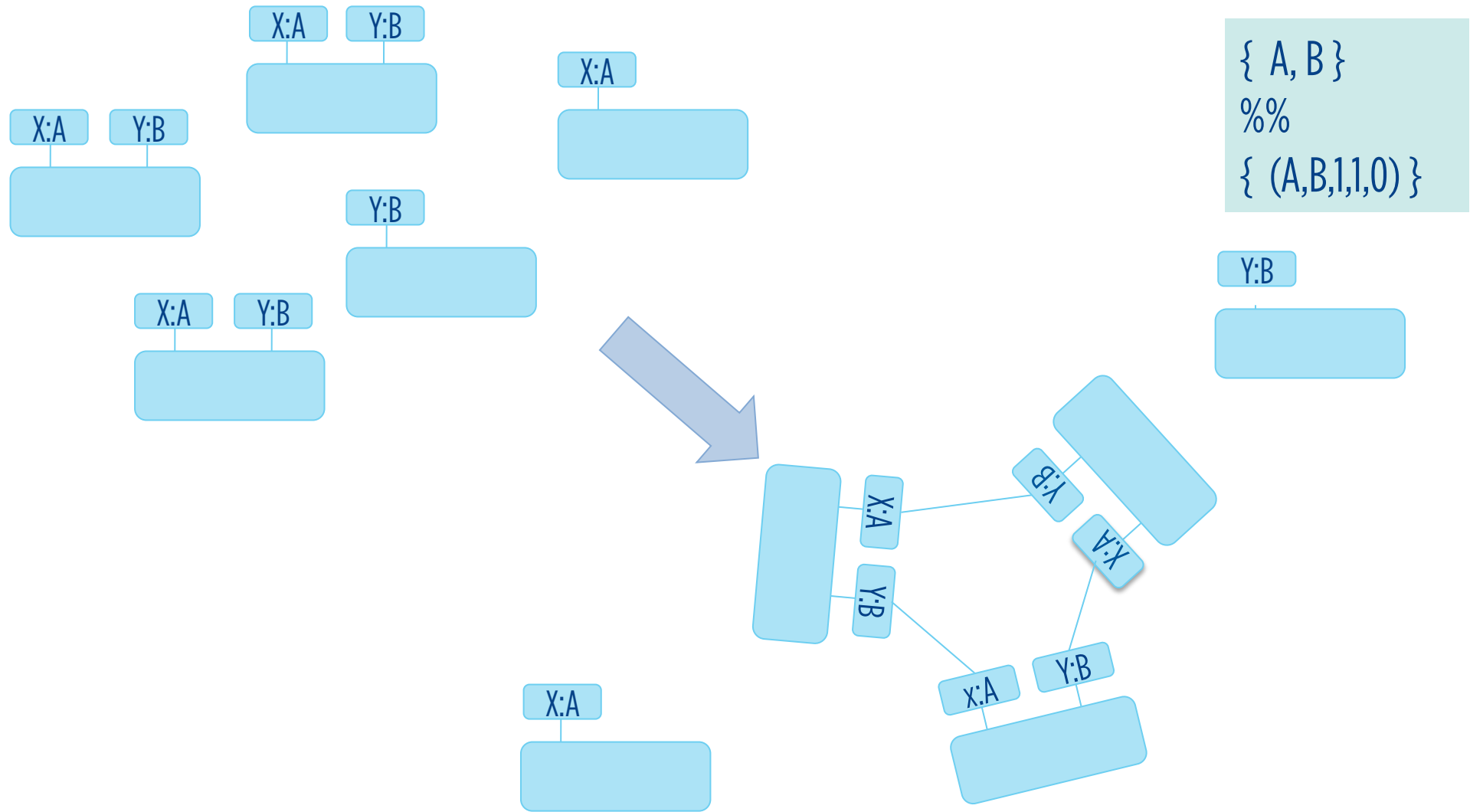


R. Larcher, C. Priami and A. Romanel. *Modelling self-assembly in BlenX*.
Transactions on Computational Systems Biology, XII:LNBI 5945, 163-198, 2010.

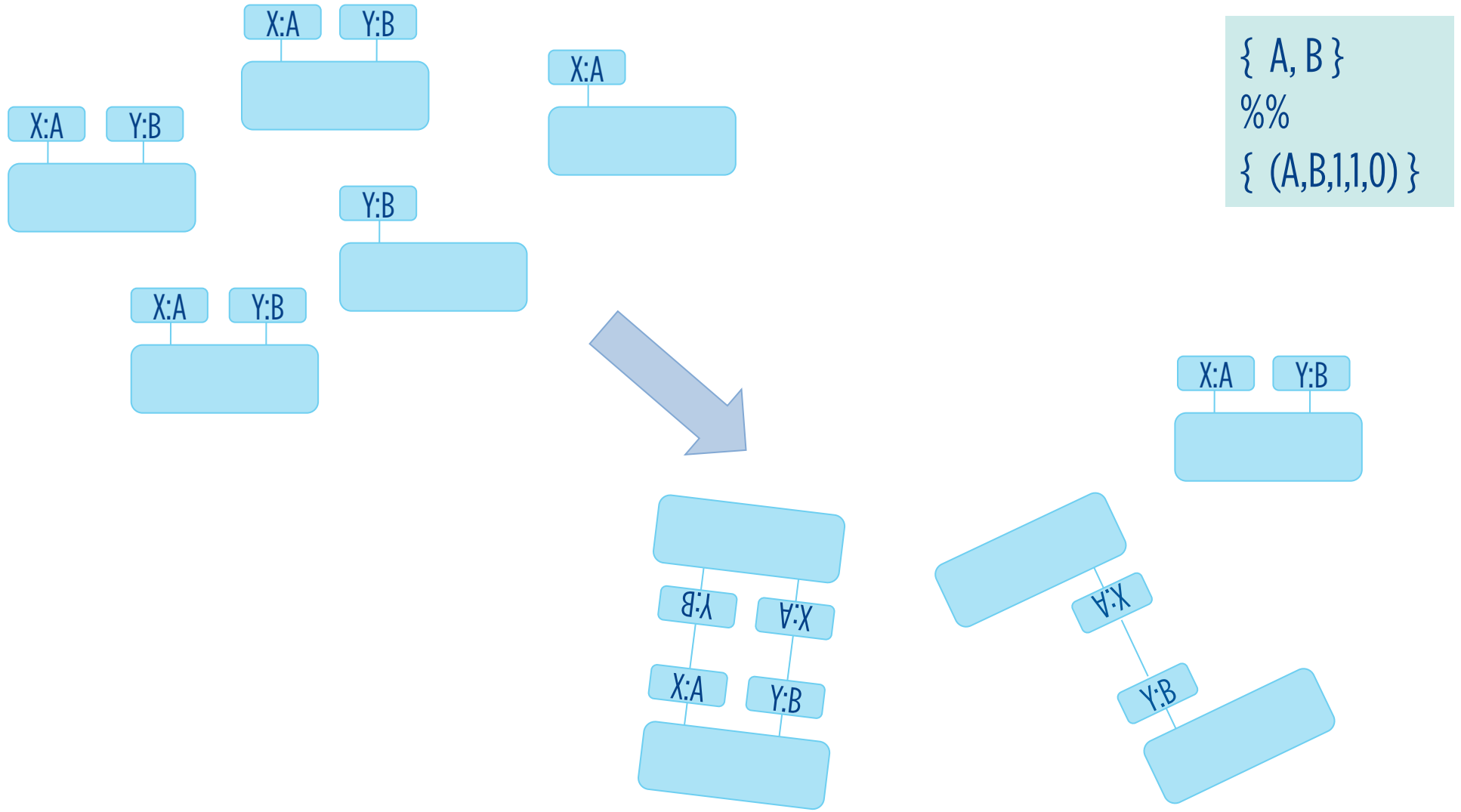
DYNAMICALLY CREATING STRUCTURES



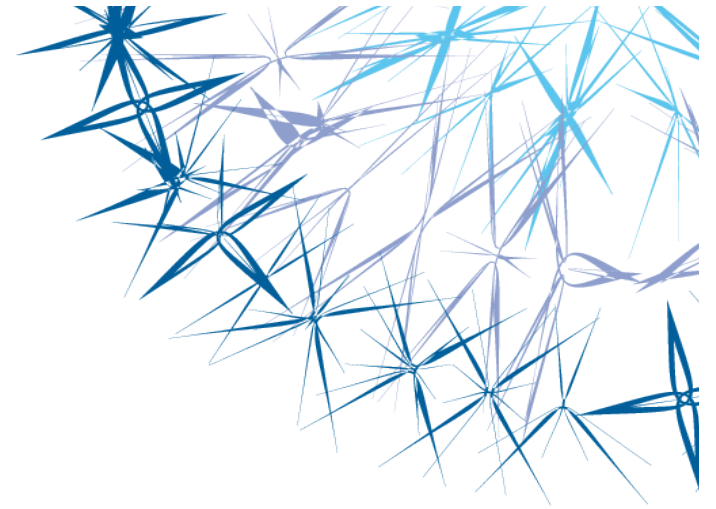
DYNAMICALLY CREATING STRUCTURES



DYNAMICALLY CREATING STRUCTURES



Example
SELF assembly of trees



SELF ASSEMBLY OF TREES

```
// Tree.prog
[ steps = 10, delta = 10 ]
<< BASERATE:inf, HIDE:inf, UNHIDE:inf >>

// Initiator Definition
let Initiator : bproc = #(out,I) [ out?().out!(root).nil ];

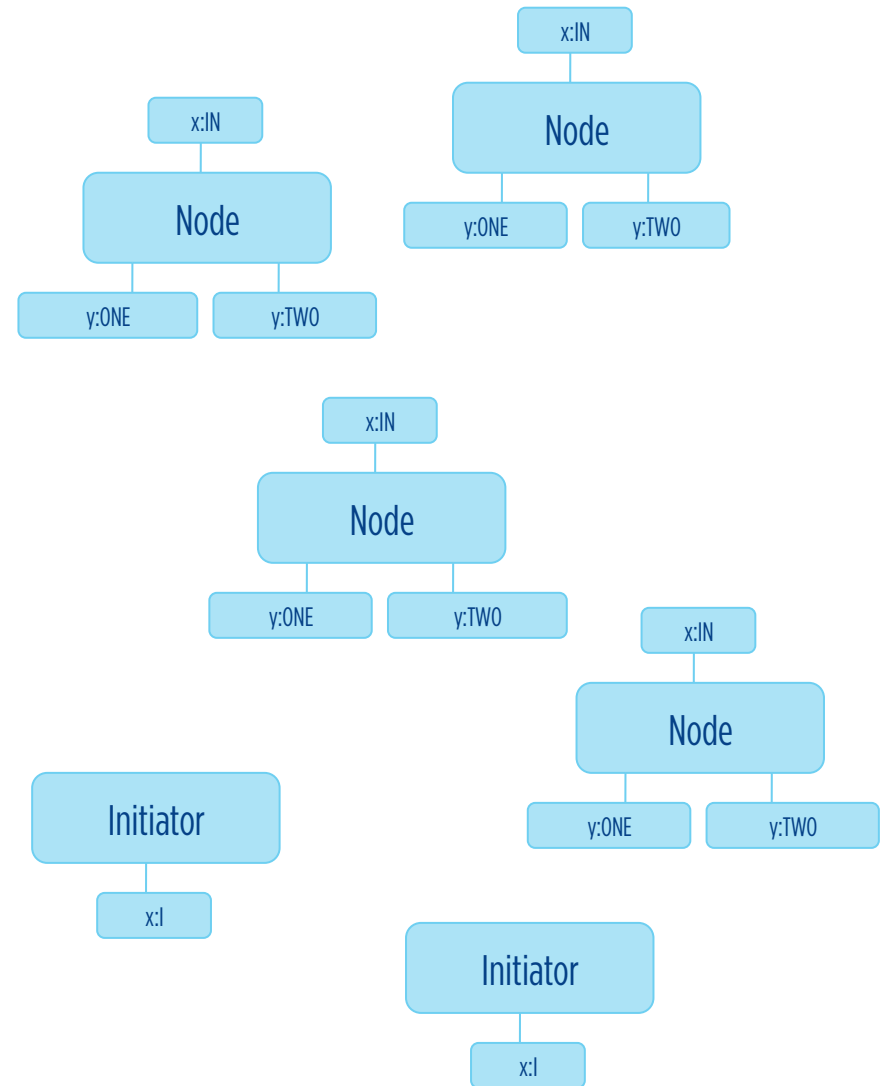
// Node Definition
let p1 : pproc = !(out?().out2?().out!(node).out2!(node).nil) ;
let p2 : pproc = !(out?().out2?().inp!().inp?(m).out1!(m).out2!(m).nil) ;
let nodeP : pproc =
  inp!().inp?(t).( t!() | (
    node?().unhide(out1).unhide(out2).p2 +
    root?().unhide(out1).unhide(out2).p1
  ) );

let Node : bproc = #(inp,IN),#h(out1,ONE),#h(out2,TWO)
  [ nodeP ];

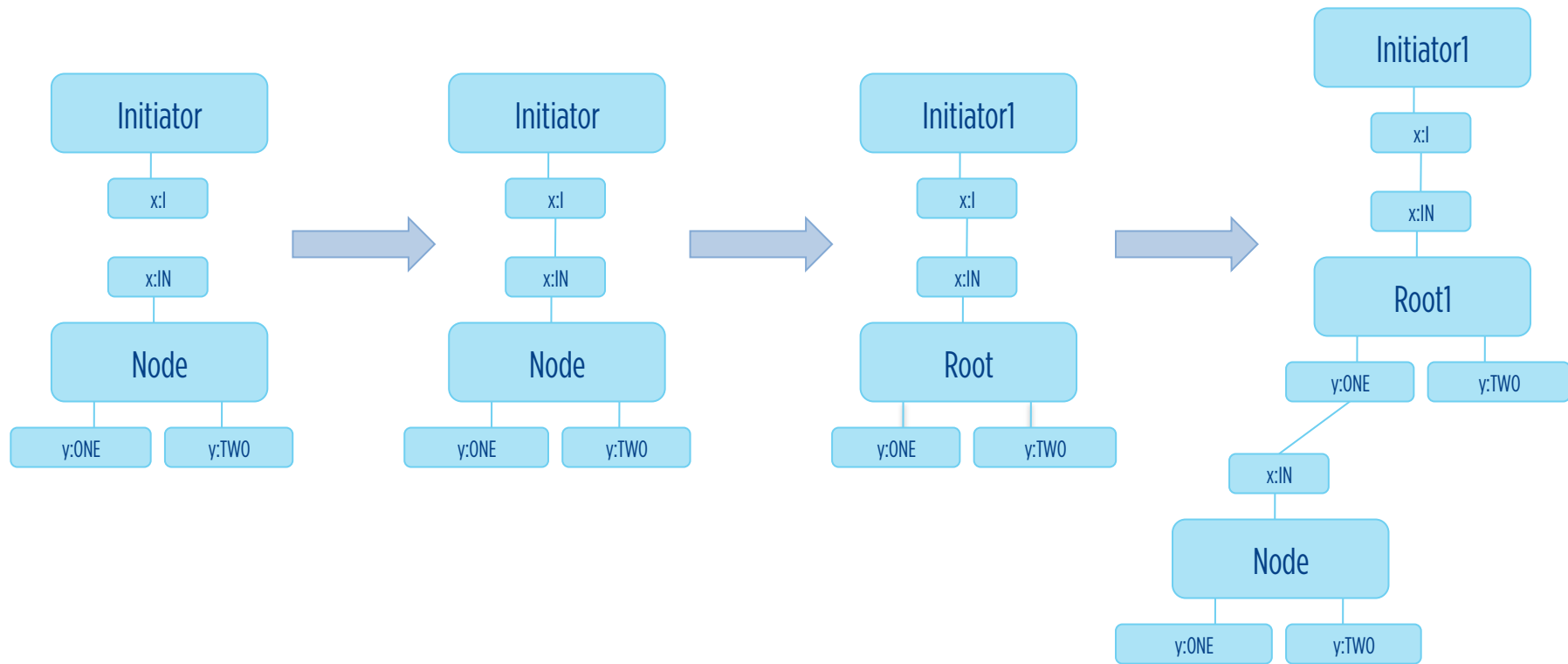
// Init
run 2 Initiator || 10 Node
```

```
// Tree.types
```

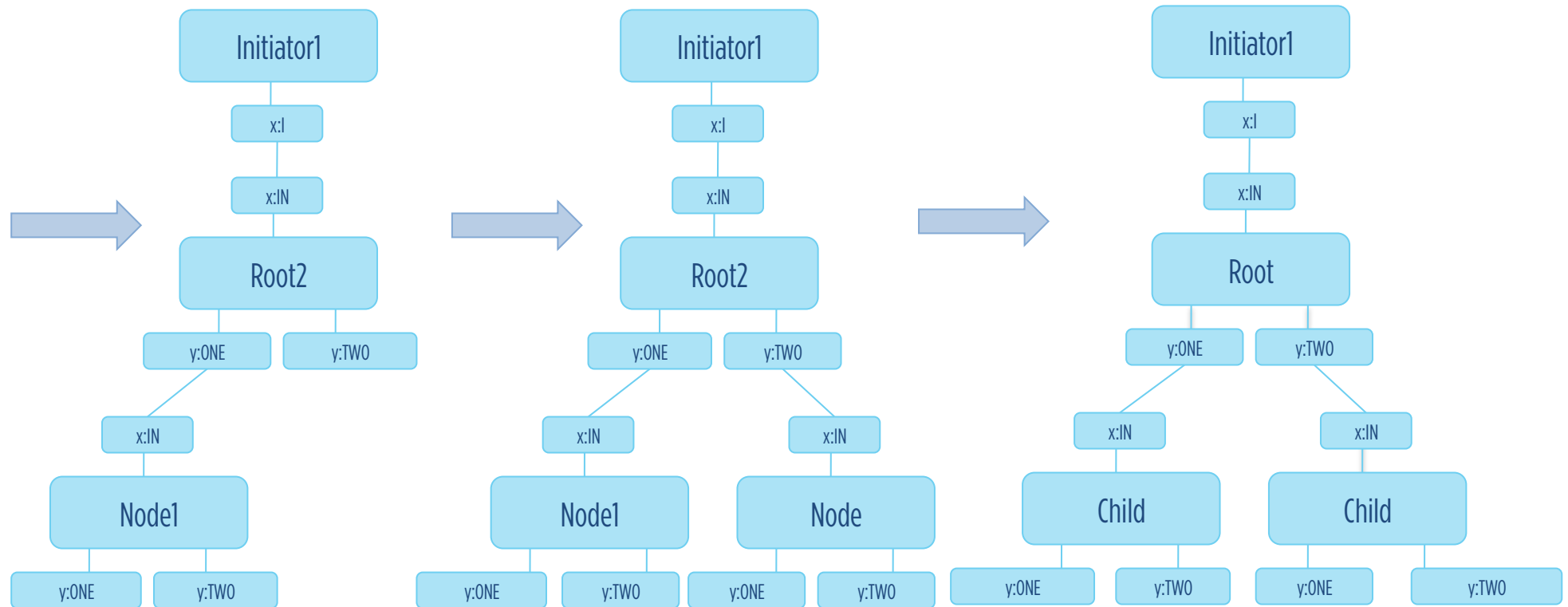
```
{ I, IN, ONE, TWO }
%%
{
  (I,IN,100,0,inf),
  (ONE,IN,1,0,inf),
  (TWO,IN,1,0,inf)
}
```



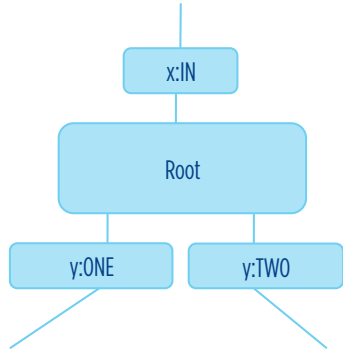
SELF ASSEMBLY OF TREES



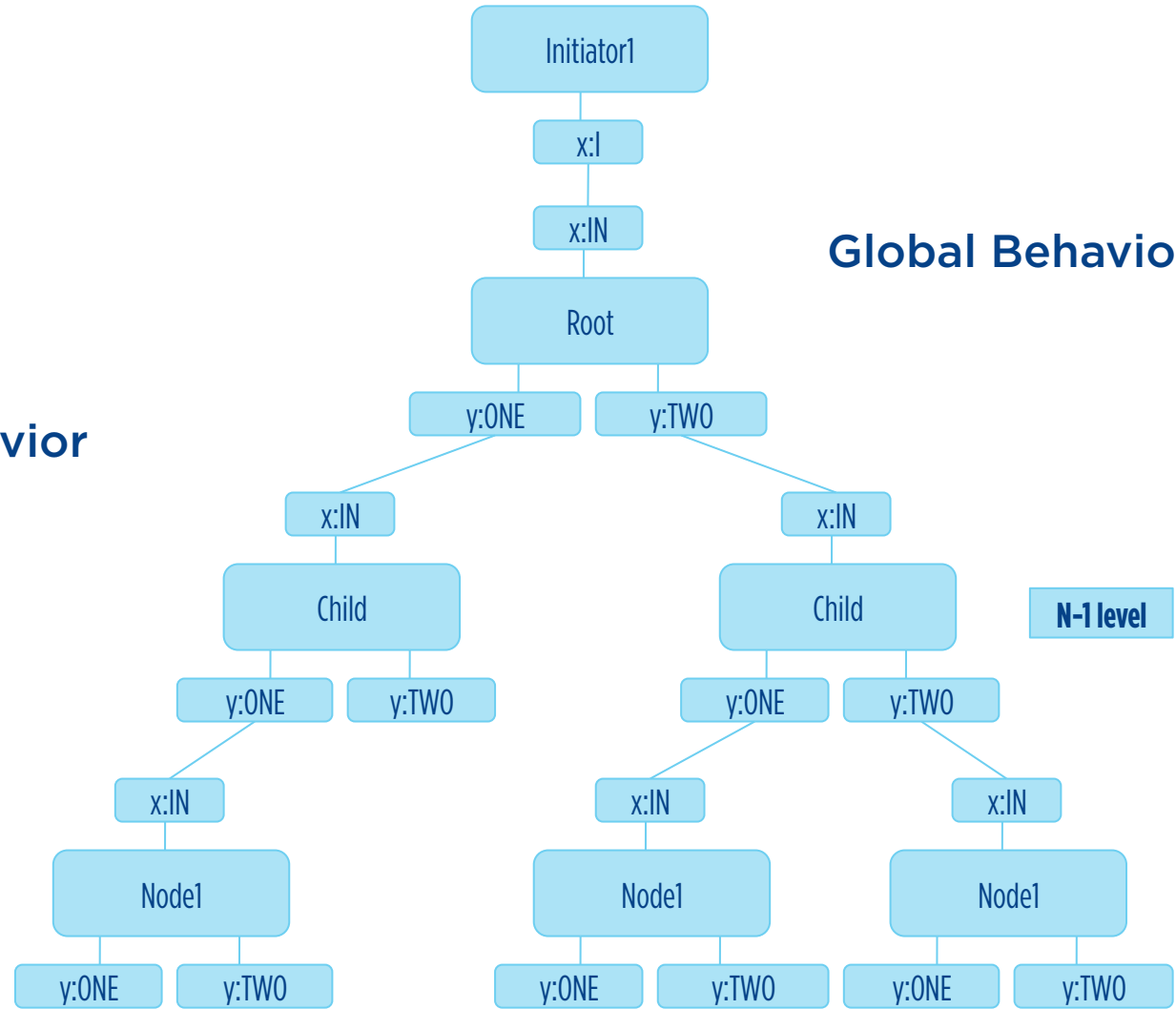
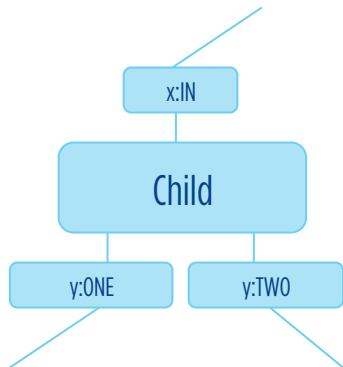
SELF ASSEMBLY OF TREES



SELF ASSEMBLY OF TREES

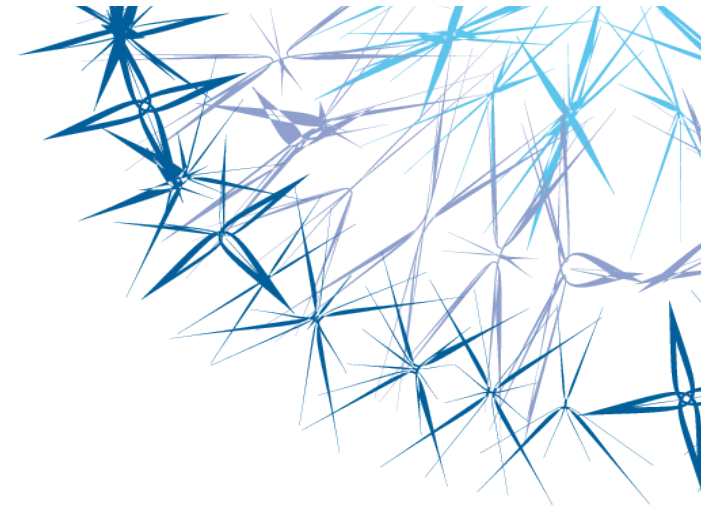


Local Behavior



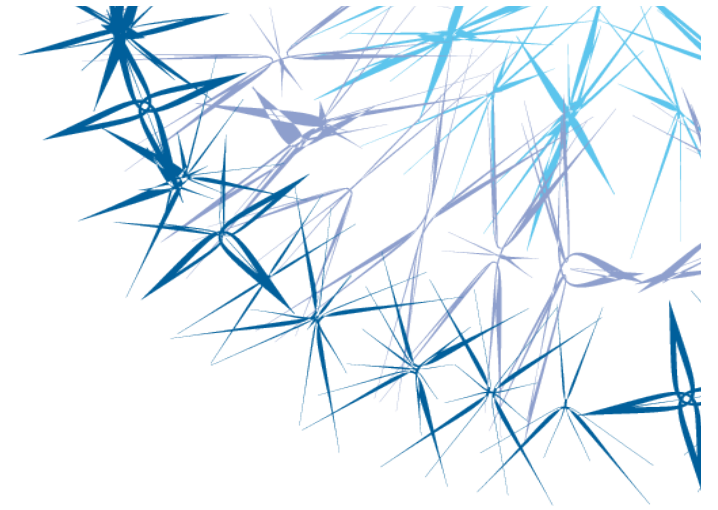
Global Behavior

END Example
SELF assembly of trees



BLENX

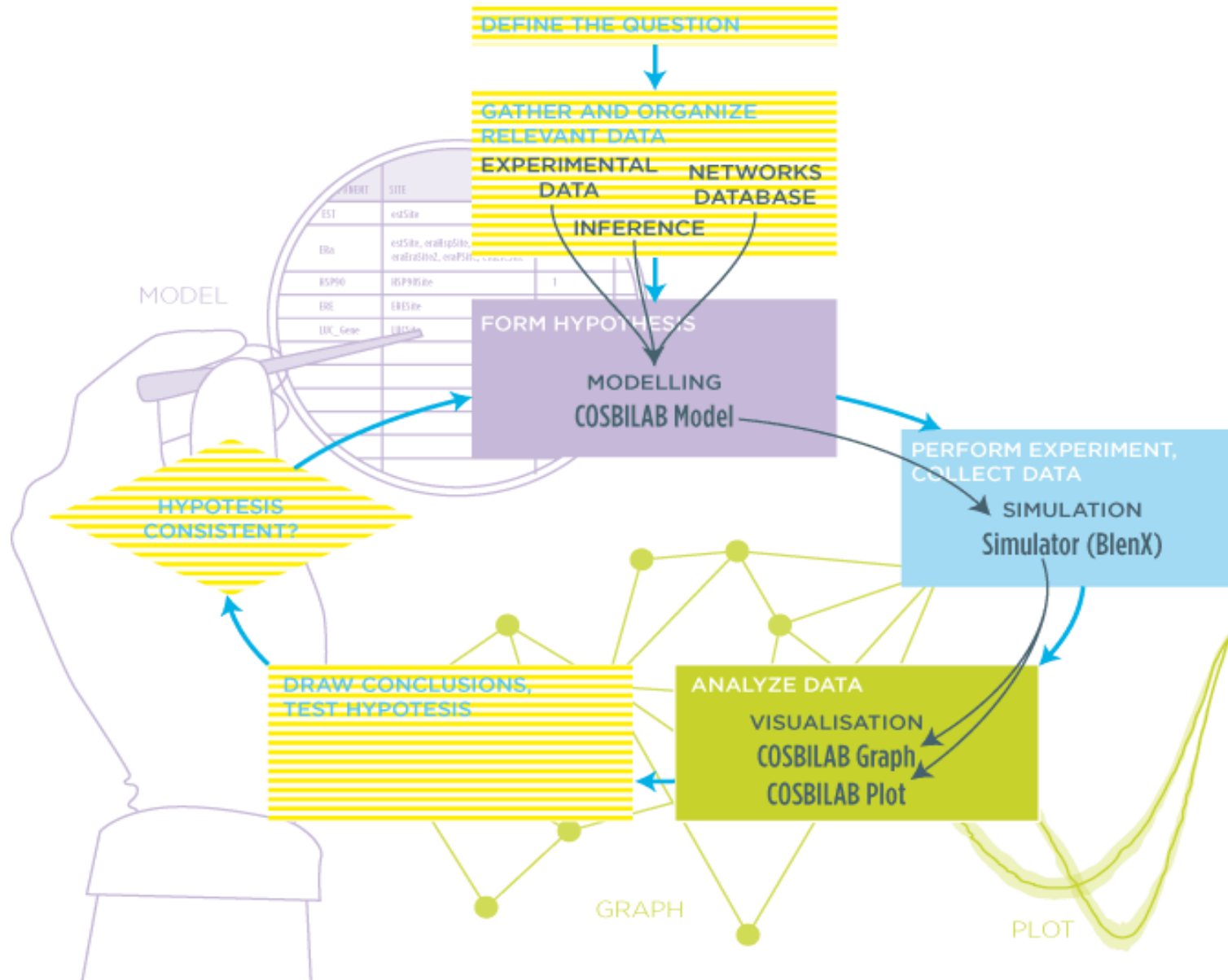
RECAP MAIN CONCEPTS

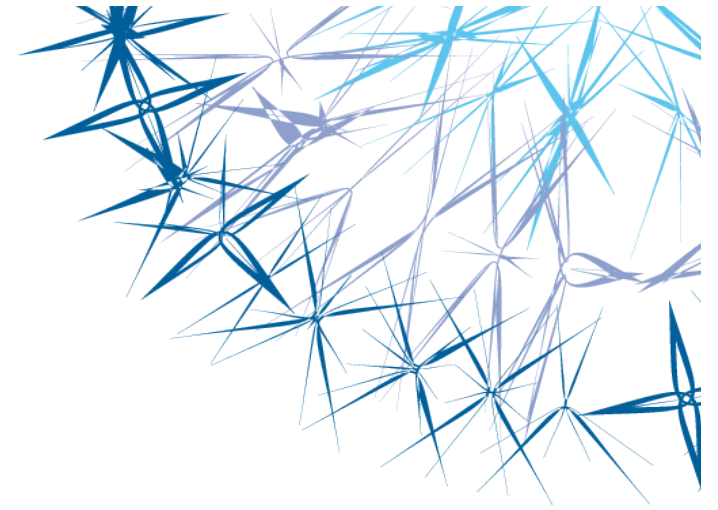


RECAP BLENX

- Simulation and analysis (causality, logical properties)
- Scalability, modularity, compositionality
- Different levels of abstraction and refinement
- Easy models, easy libraries due to combinatorial effects ruled out at modeling level
- Executable vs. solvable specifications, modules vs. variables, dynamic relations vs. static relations

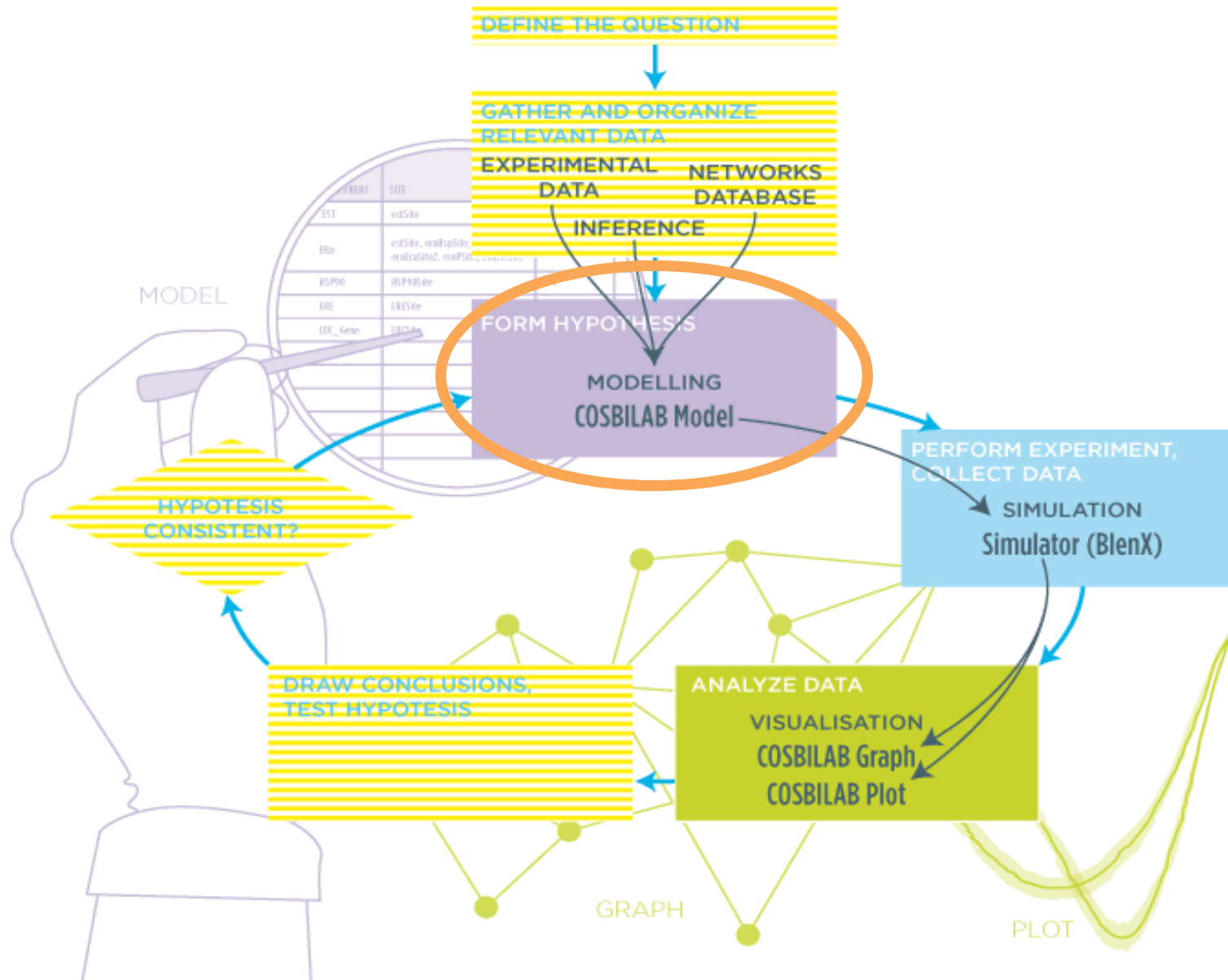
COSBI WORKING FLOW





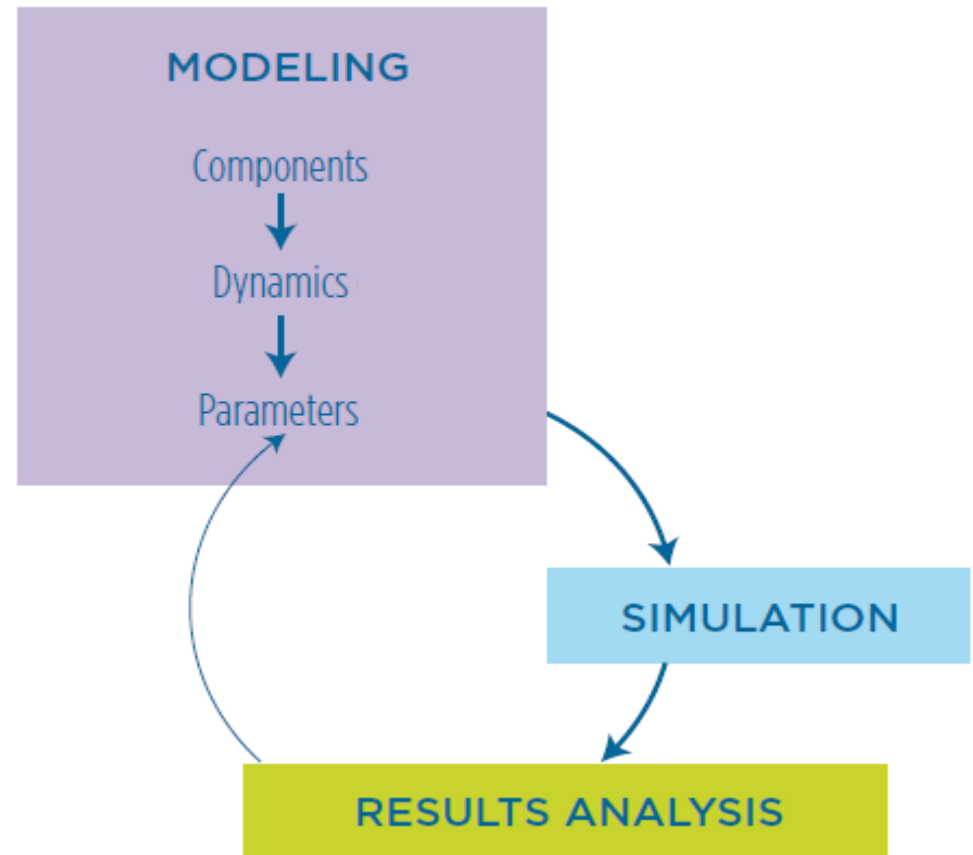
Usability of the framework

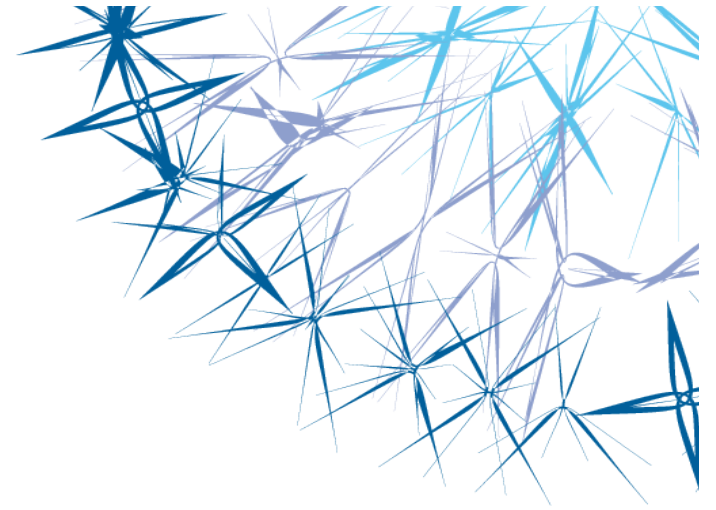
COSBI WORKING FLOW



MODELING WORKFLOW

- *Components* represent the biological entities acting in the scenario;
- Interaction rules are specified in an **intelligible, narrative language**;
- Kinetic *parameters* are summarized in a single page from where users can perform multiple **in silico experiments**;
- An integrated, stochastic simulation engine (based on the **Gillespie algorithm**) is included;
- Models are translated to BlenX, a lower level modeling language for biological systems.





Programming without programming

COSBILAB MODEL

High-level, tabular
interface for BlenX

Hiding programming
details from BlenX users

Strongly inspired to a
narrative language

M.L. Guerriero, J. Heath and C. Priami. *An automated translation from a narrative language for biological modelling into process algebra*. Proceedings of CMSB07, **LNBI 4695**, 136-151, Springer, 2007.

M.L. Guerriero, A. Dudka, N. Underhill-Day, J.K. Heath, C. Priami. *Narrative-based computational modelling of the Gp130/JAK/STAT signalling pathway*. **BMC Systems Biology**, **3**:40, 2009.

C. Priami, P. Ballarini and P. Quaglia *BlenX4Bio: BlenX for Biologists*. Proceedings of CMSB09, **LNBI 5688**, Springer, 2009.

COSBI MODELING PHILOSOPHY

Intelligible models:

- Understandable to non-expert modelers
- Easy to write, modify and reuse
- Manage combinatorial complexity of dynamics

INTELLIGIBILITY

- < Obscure syntax
- < No bio keywords



Narrative Language

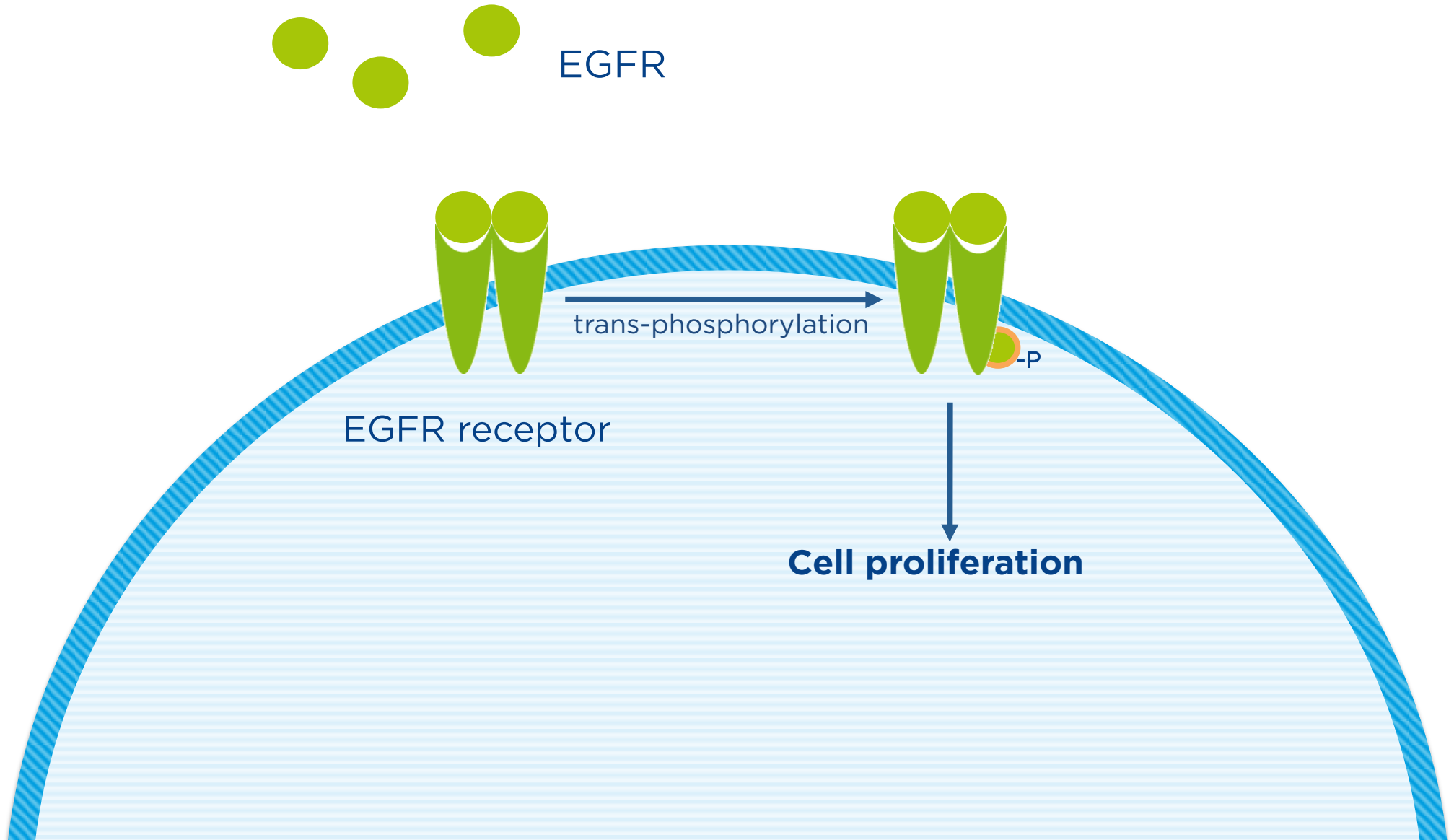


- < Large size
- < Complex mathematics

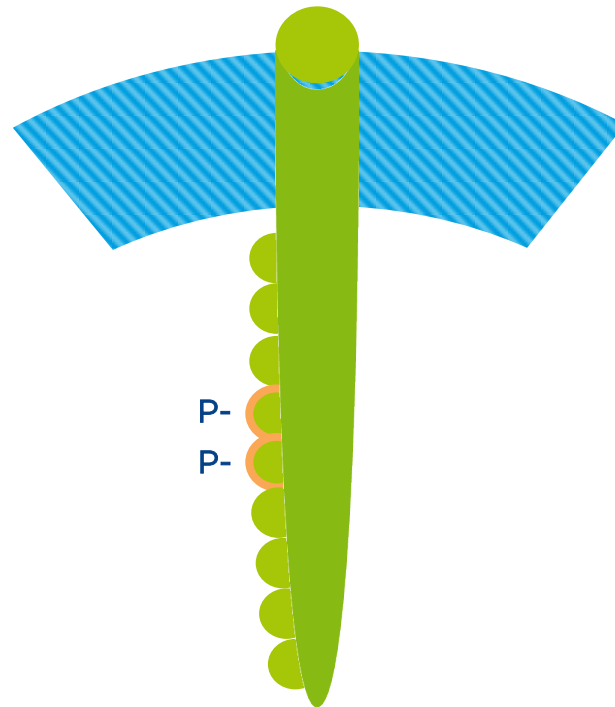


Minimize required modeling information

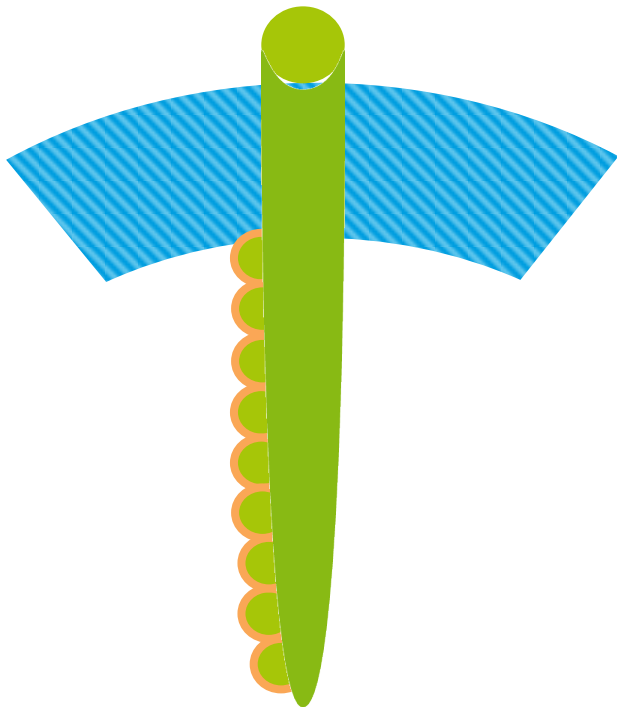
EPIDERMAL GROWTH FACTOR



EGFR MOLECULE

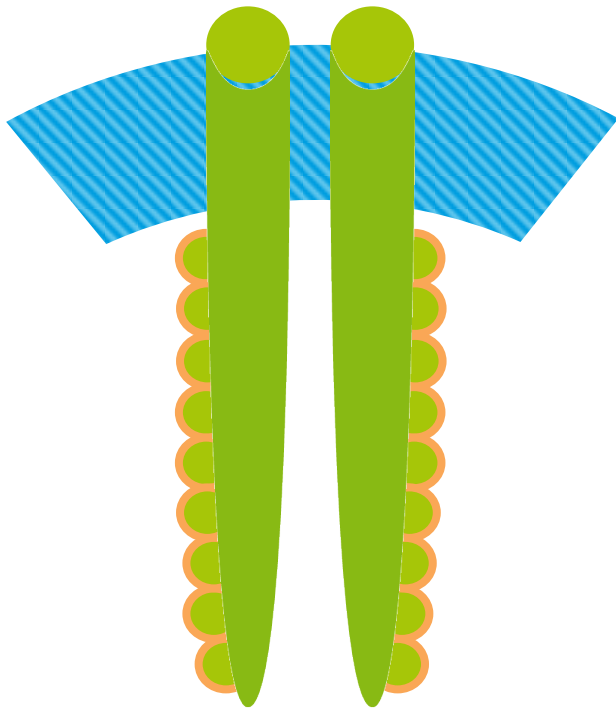


EGFR MOLECULE



2⁹ configuration of EGFR
that is **512** different states

EGFR HOMODIMERS



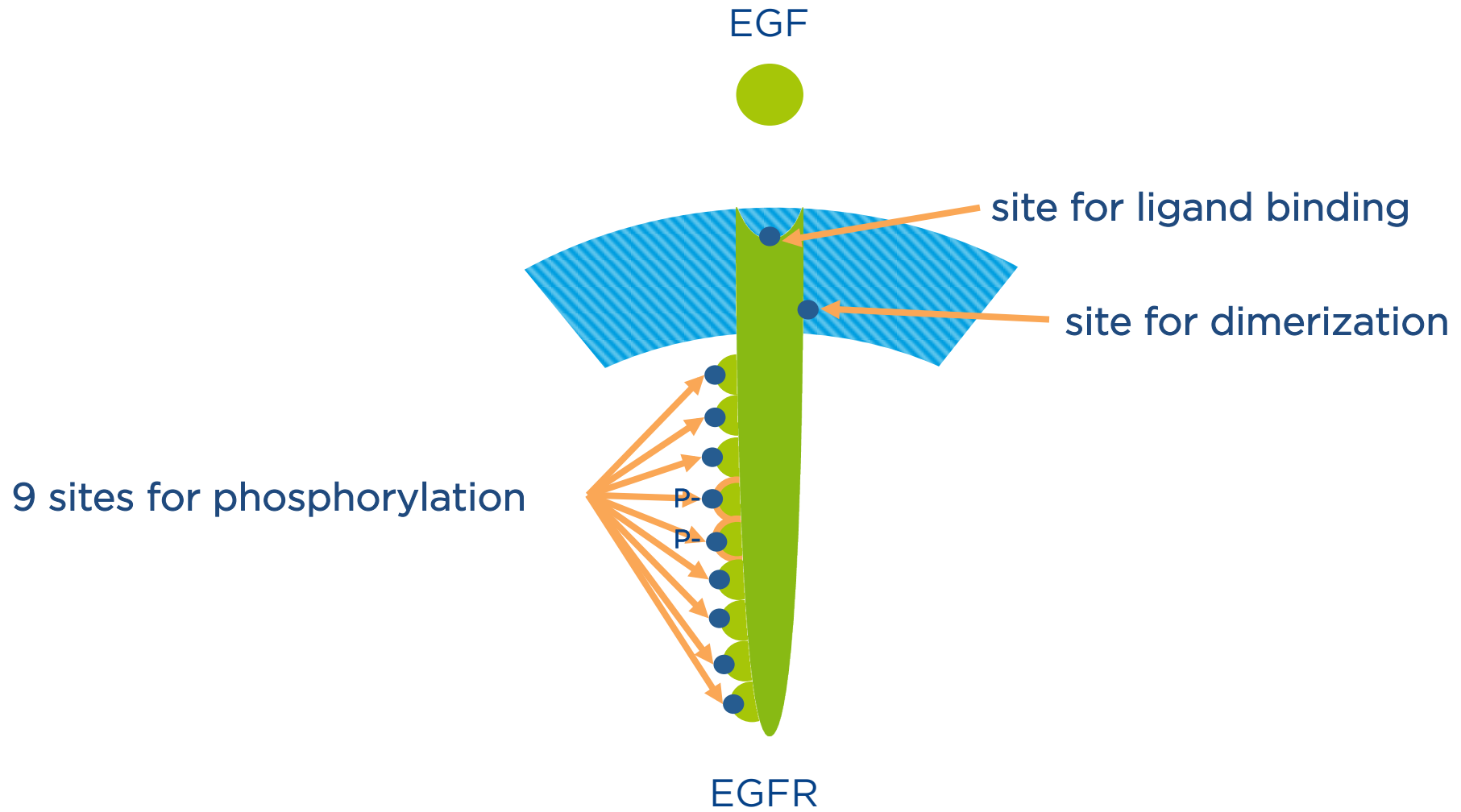
2^{18} configuration of EGFR

Homodimers

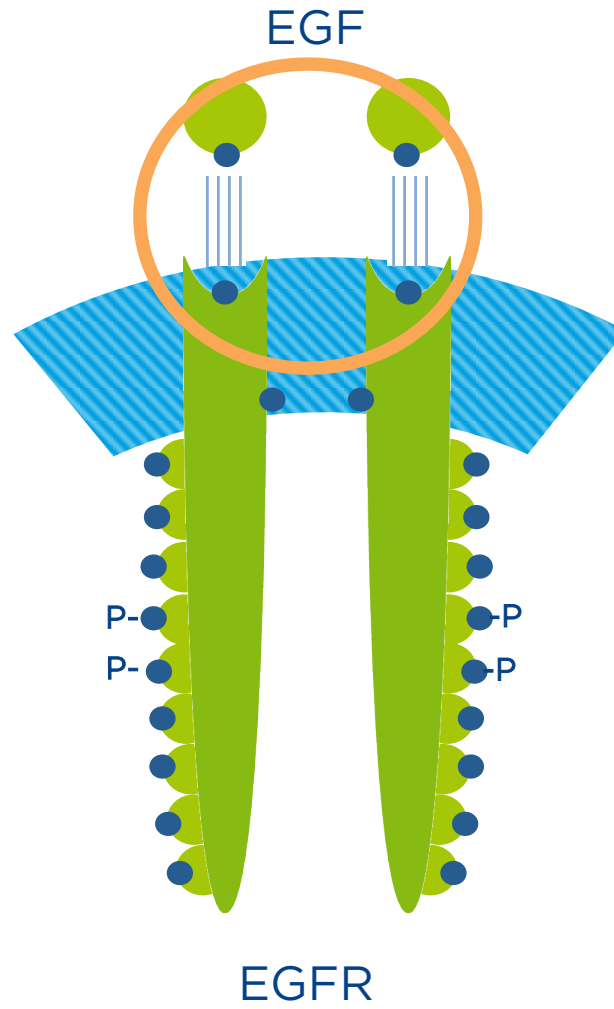
that is more than **250 000**

different states

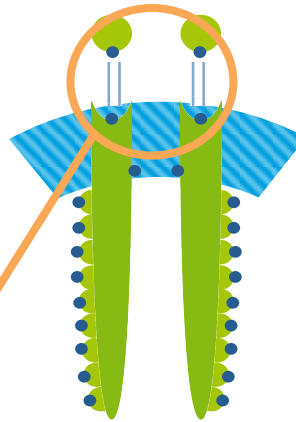
EGFR MOLECULE



INTERACTION BETWEEN EGFR AND EGF



INTERACTION BETWEEN EGFR AND EGF



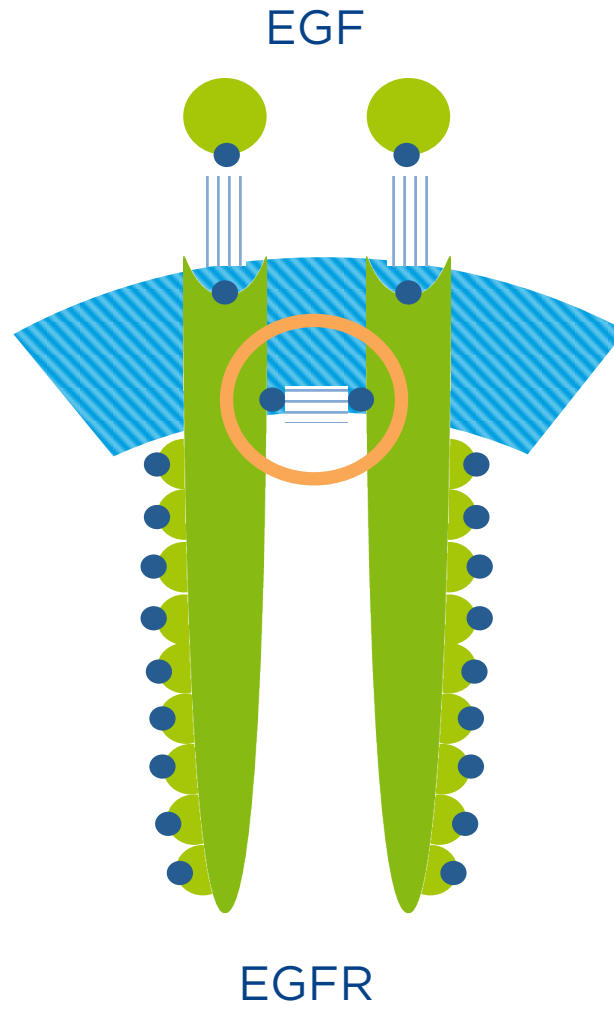
New - COSBI LAB Model 1.0.0.57

File Edit Simulation Help

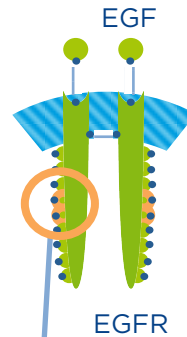
Components Complexes Global Dynamics Bimolecular Dynamics Binding Dynamics Translocations Compartments Parameters Initial State

	Binding	(Parameter) Binding Rate	(Parameter) Unbinding Rate	Condition
Click here to add new item				
<input type="checkbox"/>	EGFR@Membrane on s_arm binds EGF@System on s_lig	r_EGFR_EGF_bin	r_EGFR_EGF_unb	None
<input checked="" type="checkbox"/>	EGFR on s_dim binds EGFR on s_dim	r_EGFR_EGFR_bin	r_EGFR_EGFR_unb	Binding, Unbinding

INTERACTION BETWEEN EGF AND EGFR



SITE'S DYNAMIC FOR STATE CHANGE



Site's Dynamic for state change

New - COSBI LAB Model 1.0.0.4858 Continuous Integration [Development]_20110912.5

File Edit Simulation Help

Components Complexes Global Dynamics Bimolecular Dynamics Binding Dynamics Translocations Compartments Parameters Initial State

Component Name	Site	Configurations	Description
EGFR	s_arm, s_dim, s_ph1, s_ph2, s_ph3, s_ph4, s_ph5, s_ph6	1	

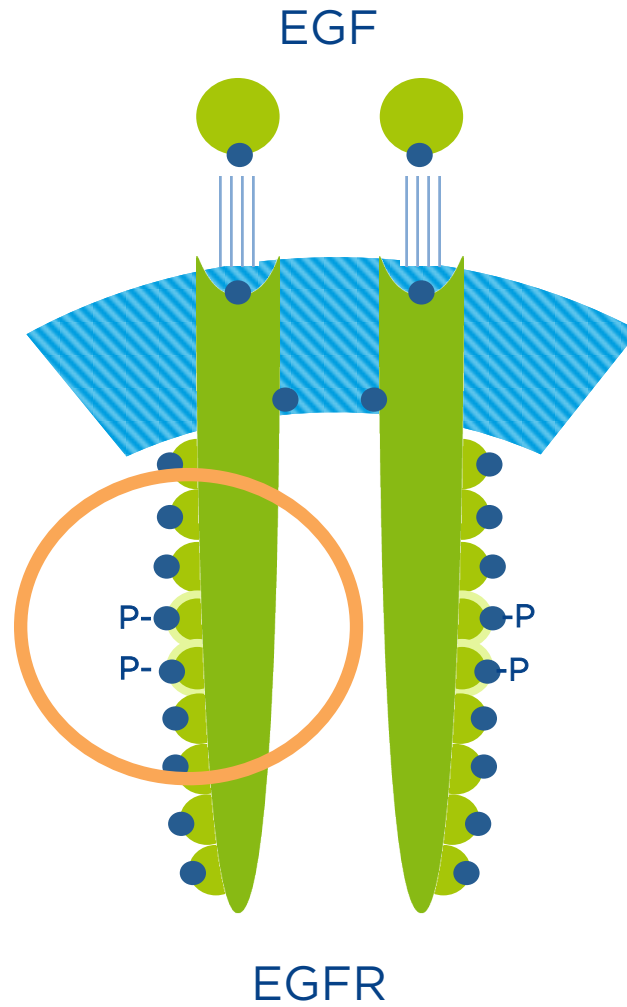
Click here to add new item

Component Site Definition Component Configurations Monomolecular Dynamics

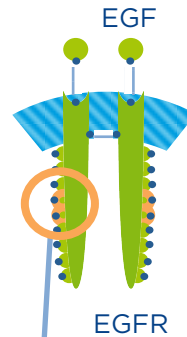
Action	(Parameter) Rate	Condition	Compartment	Description
Phosphorylated on s_ph1	r_EGFR_ph1	s_dim is Bound	Membrane	
Phosphorylated on s_ph2	r_EGFR_ph2	s_dim is Bound	Membrane	
Phosphorylated on s_ph3	r_EGFR_ph3	s_dim is Bound	Membrane	
Phosphorylated on s_ph4	r_EGFR_ph4	s_dim is Bound	Membrane	
Phosphorylated on s_ph5	r_EGFR_ph5	s_dim is Bound	Membrane	
Phosphorylated on s_ph6	r_EGFR_ph6	s_dim is Bound	Membrane	

EGF	s_lig		1	
SHC	s_ph		1	

SITE'S DYNAMIC FOR STATE CHANGE



SITE'S DYNAMIC FOR STATE CHANGE



Site's Dynamic for state change

New - COSBI LAB Model 1.0.0.4858 Continuous Integration [Development]_20110912.5

File Edit Simulation Help

Components Complexes Global Dynamics Bimolecular Dynamics Binding Dynamics Translocations Compartments Parameters Initial State

Component Name	Site	Configurations	Description
EGFR	s_arm, s_dim, s_ph1, s_ph2, s_ph3, s_ph4, s_ph5, s_ph6	1	

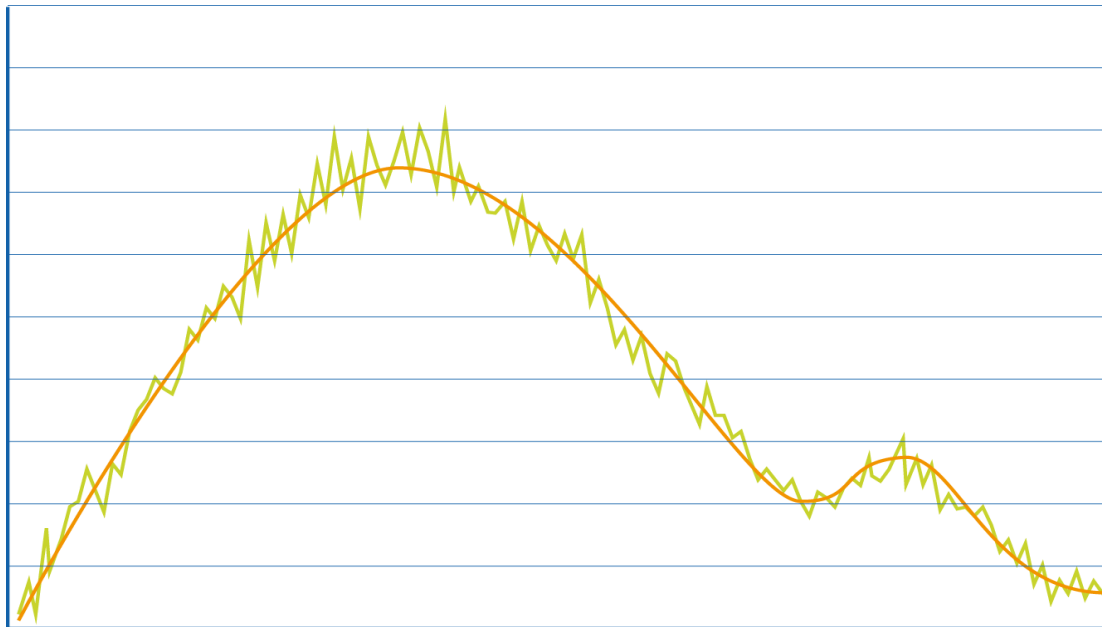
Click here to add new item

Component Site Definition Component Configurations Monomolecular Dynamics

Action	(Parameter) Rate	Condition	Compartment	Description
Phosphorylated on s_ph1	r_EGFR_ph1	s_dim is Bound	Membrane	
Phosphorylated on s_ph2	r_EGFR_ph2	s_dim is Bound	Membrane	
Phosphorylated on s_ph3	r_EGFR_ph3	s_dim is Bound	Membrane	
Phosphorylated on s_ph4	r_EGFR_ph4	s_dim is Bound	Membrane	
Phosphorylated on s_ph5	r_EGFR_ph5	s_dim is Bound	Membrane	
Phosphorylated on s_ph6	r_EGFR_ph6	s_dim is Bound	Membrane	

EGF	s_lig		1	
SHC	s_ph		1	

DISCRETE STOCHASTIC APPROACH

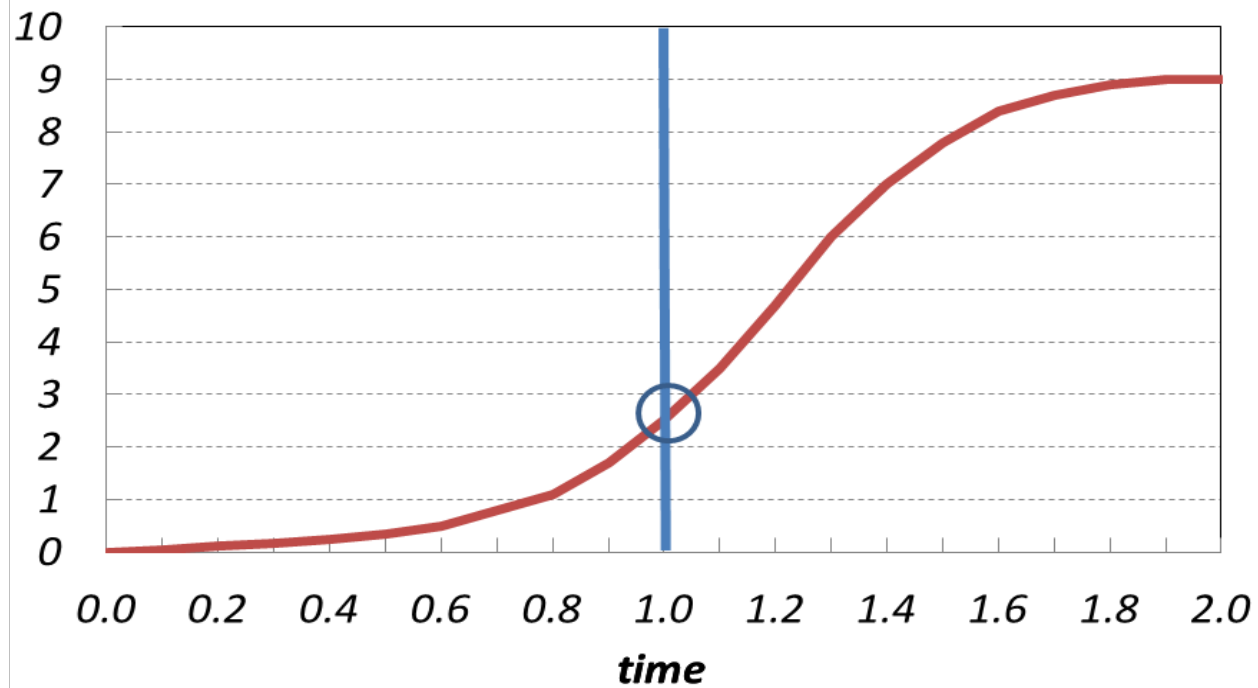


DISCRETE STOCHASTIC APPROACH

- Molecular interaction is discrete
- Receive more detailed information
- Able to produce more accurate predictions

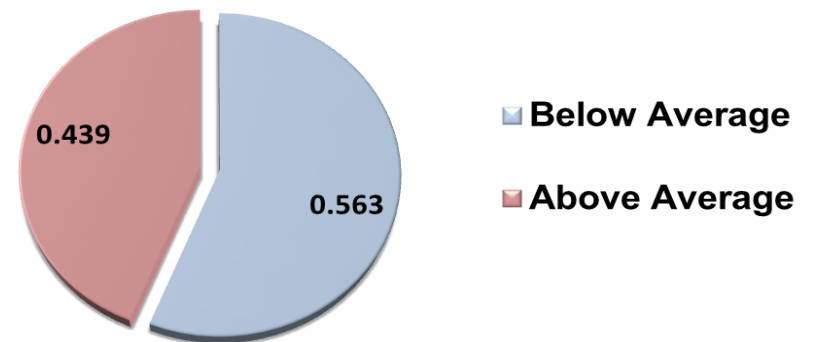
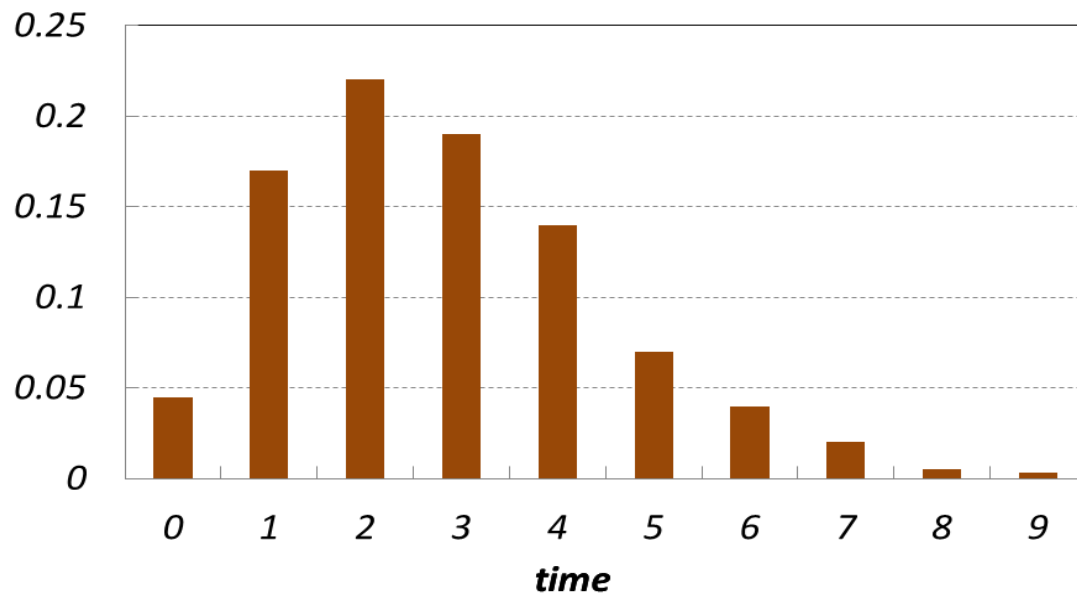
DISCRETE STOCHASTIC APPROACH EGFR EXAMPLE

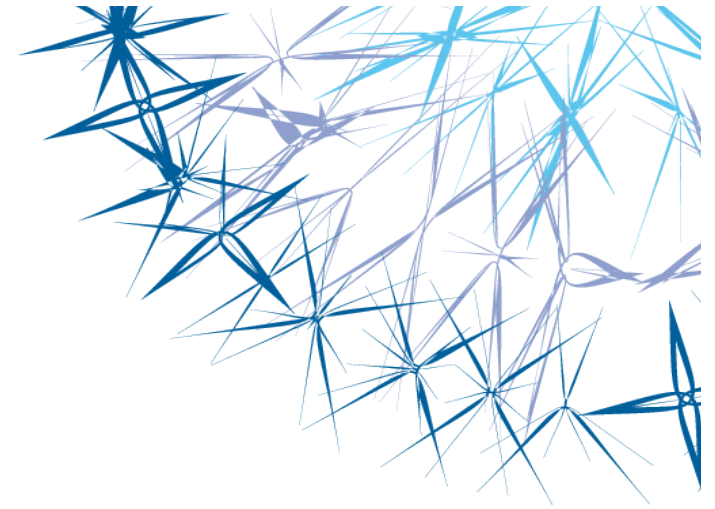
Average EGFR Phosphorylation



DISCRETE STOCHASTIC APPROACH EGFR EXAMPLE

Distribution at time t=1





COSBILAB Model

The gemcitabine example

P. Lecca, O. Kahramanogullari, D. Morpurgo, C. Priami, R. Soo. Modelling the tumor shrinkage pharmacodynamics with BlenX, **1st IEEE International Conference on Computational Advances in Bio and medical Sciences (ICCABS)**, 2011

P. Lecca, O. Kahramanogullari, D. Morpurgo, C. Priami, R. Soo. Modelling and estimating dynamics of tumor shrinkage with BlenX and Kinfer, **UKSim 2001 - 13th International Conference on Modelling and Simulation.**, 2011

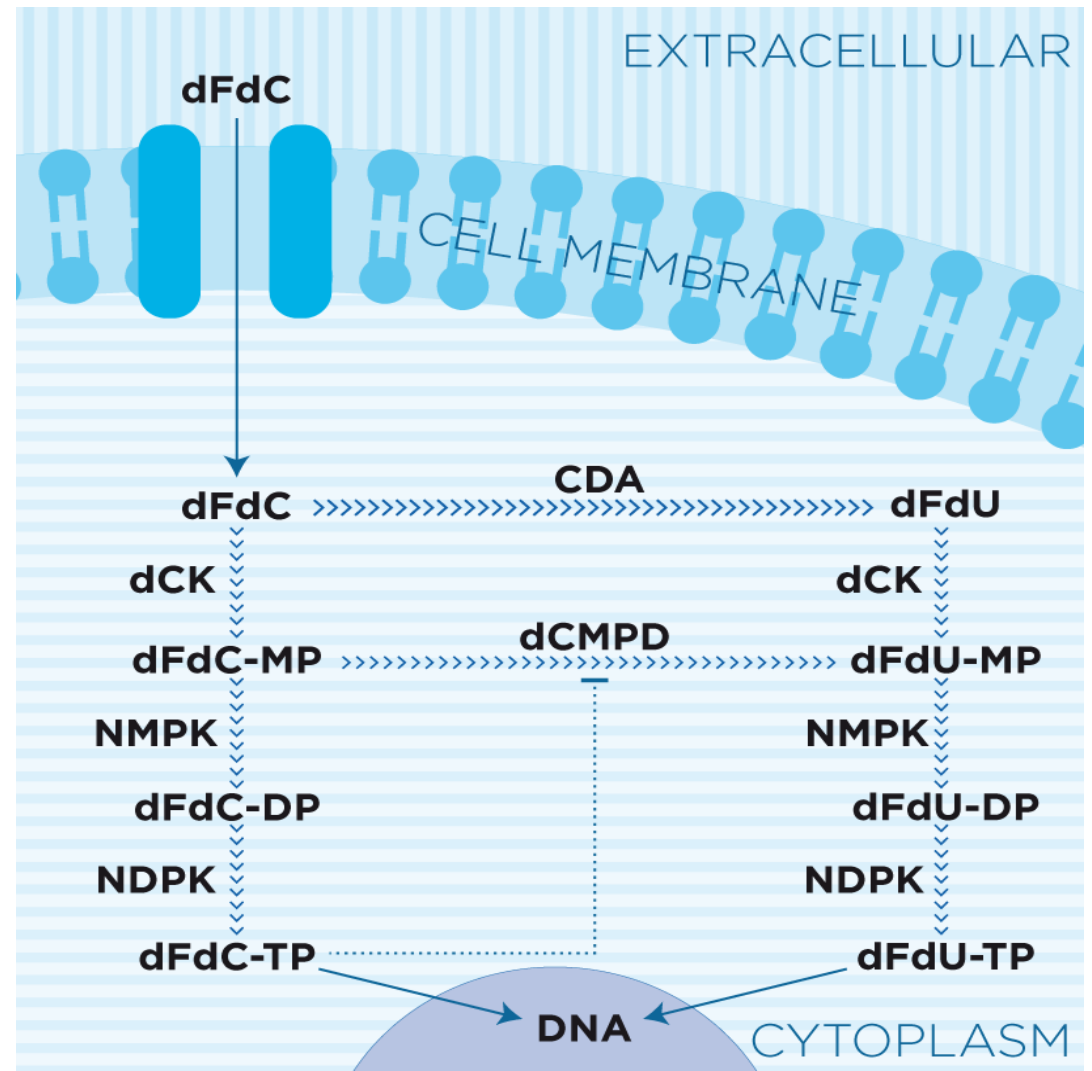
TUMOR SHRINKAGE BY GEMCITABINE

An algorithmic pharmacodynamics model for the time course of tumor shrinkage by gemcitabine in non small cell lung cancer patients with the following events:

- part of the injected gemcitabine degrades;
- the gemcitabine interacts with the tumor without any effect;
- The gemcitabine interacts with the tumor, is consumed and the tumor cell is killed;
- tumor grows.
- new doses of drug are regularly injected

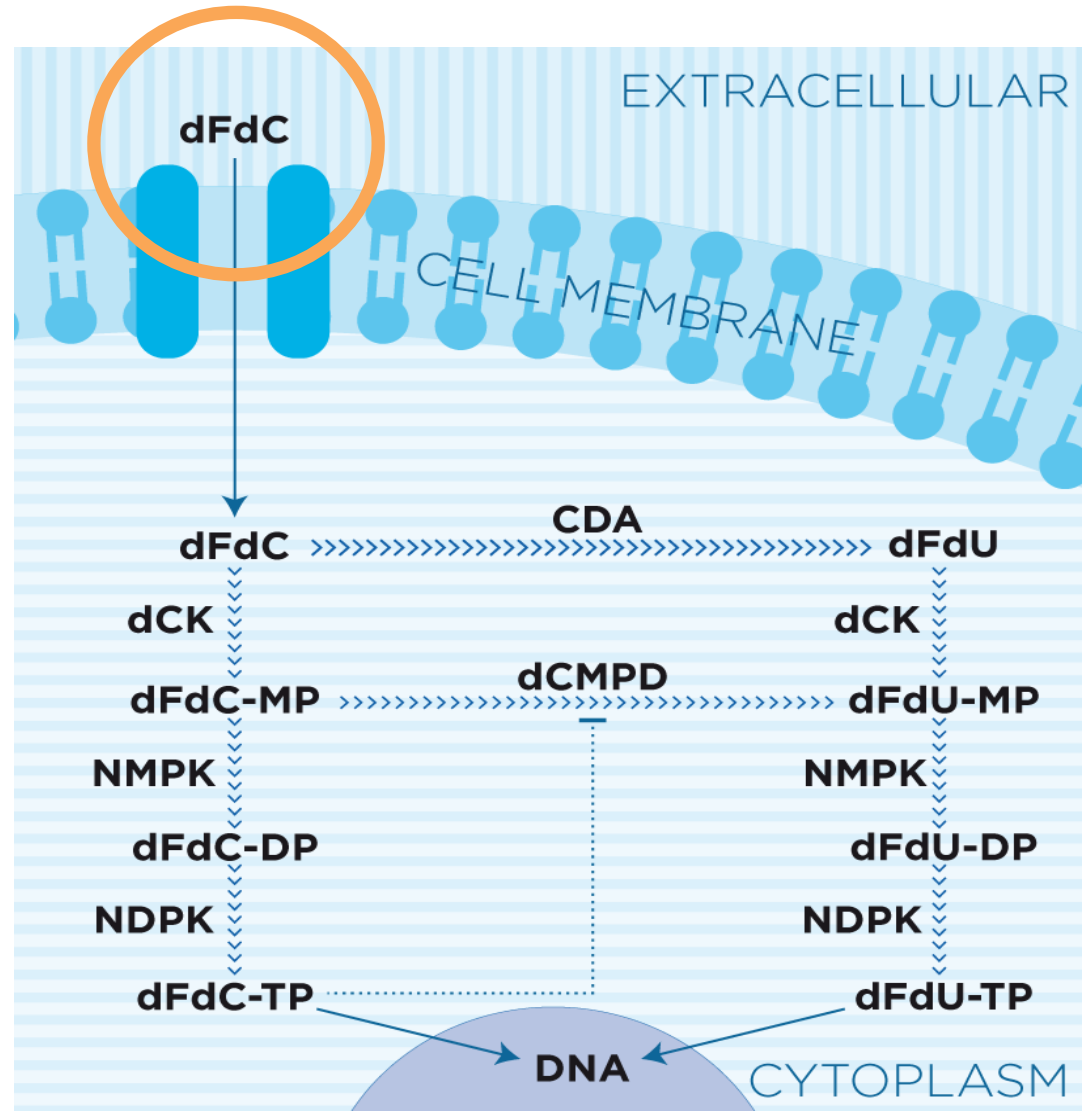
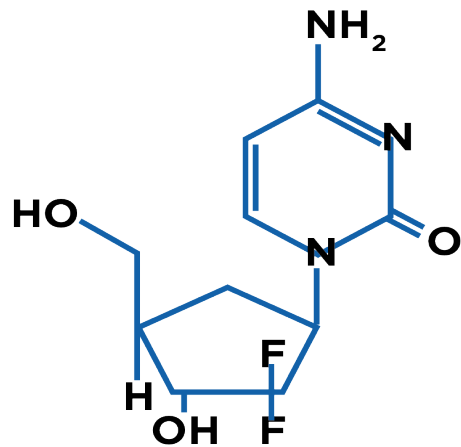
GEMCITABINE MECHANISM OF ACTION

- Gemcitabine (2'-2'-difluorodeoxycytidine, dFdC) is a nucleoside analog used in oncology to block DNA replication in tumor cells;
- Gemcitabine is transported from plasma into the cell and then it is subjected to deamination and/or multiple phosphorylation leading to its active triphosphate (dFdCTP and dFdUTP) metabolites;
- As an example, a simplified version of gemcitabine metabolic network will be modeled.



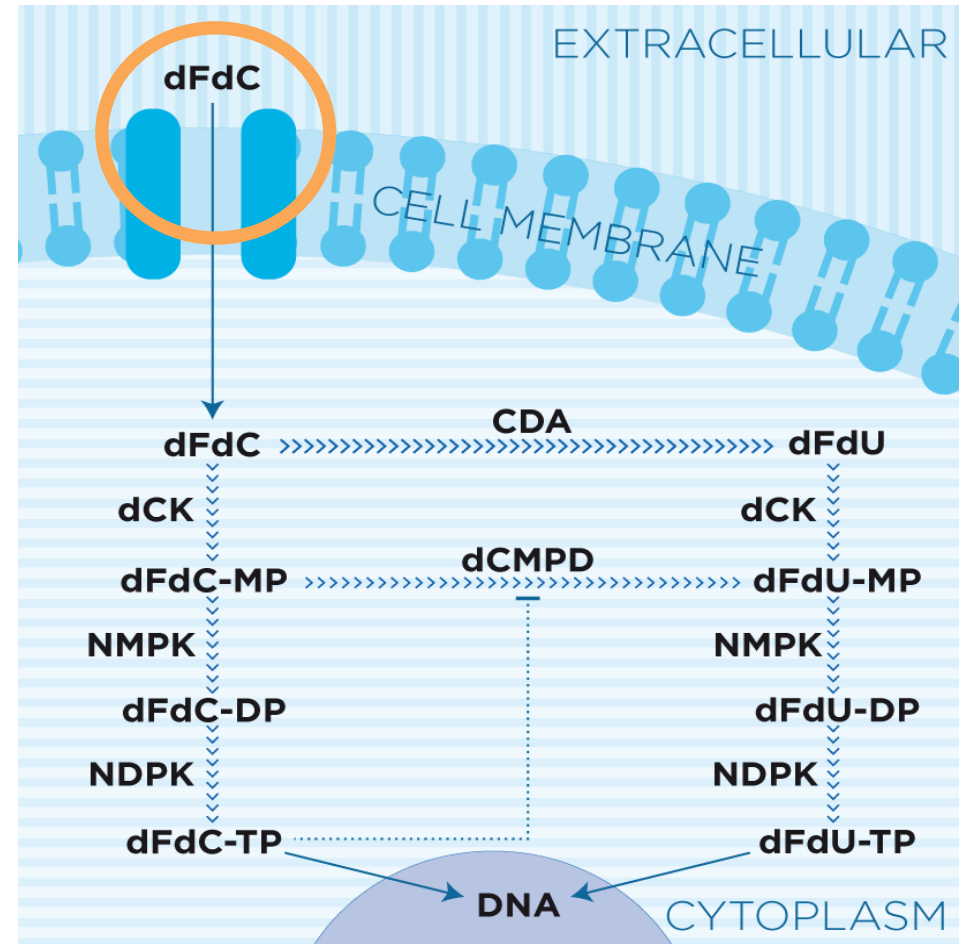
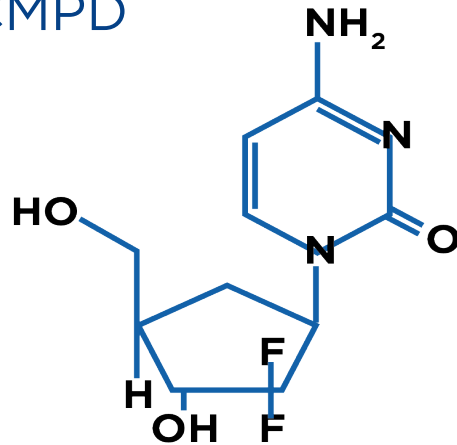
COMPONENTS

Gemcitabine (dFdC) has sites for deamination, phosphorylation, and inhibition of dCMPD enzyme.



COMPONENTS

Gemcitabine (dFdC) has sites for deamination, phosphorylation, and inhibition of dCMPD enzyme.



BlenX code:

```
let dFdcOut: bproc = #(s1, c_dFdcOut1), #(s2, c_dFdcOut2) [ p_main | rep start_p_main?().p_main ];
```

COMPONENTS

Components Complexes Global Dynamics Bimolecular Dynamics Binding Dynamics Translocati

Component Name	Site	Configurations	Description
Click here to add new item			
dFdC	nh, ph1, ph2, ph3, bin	1	

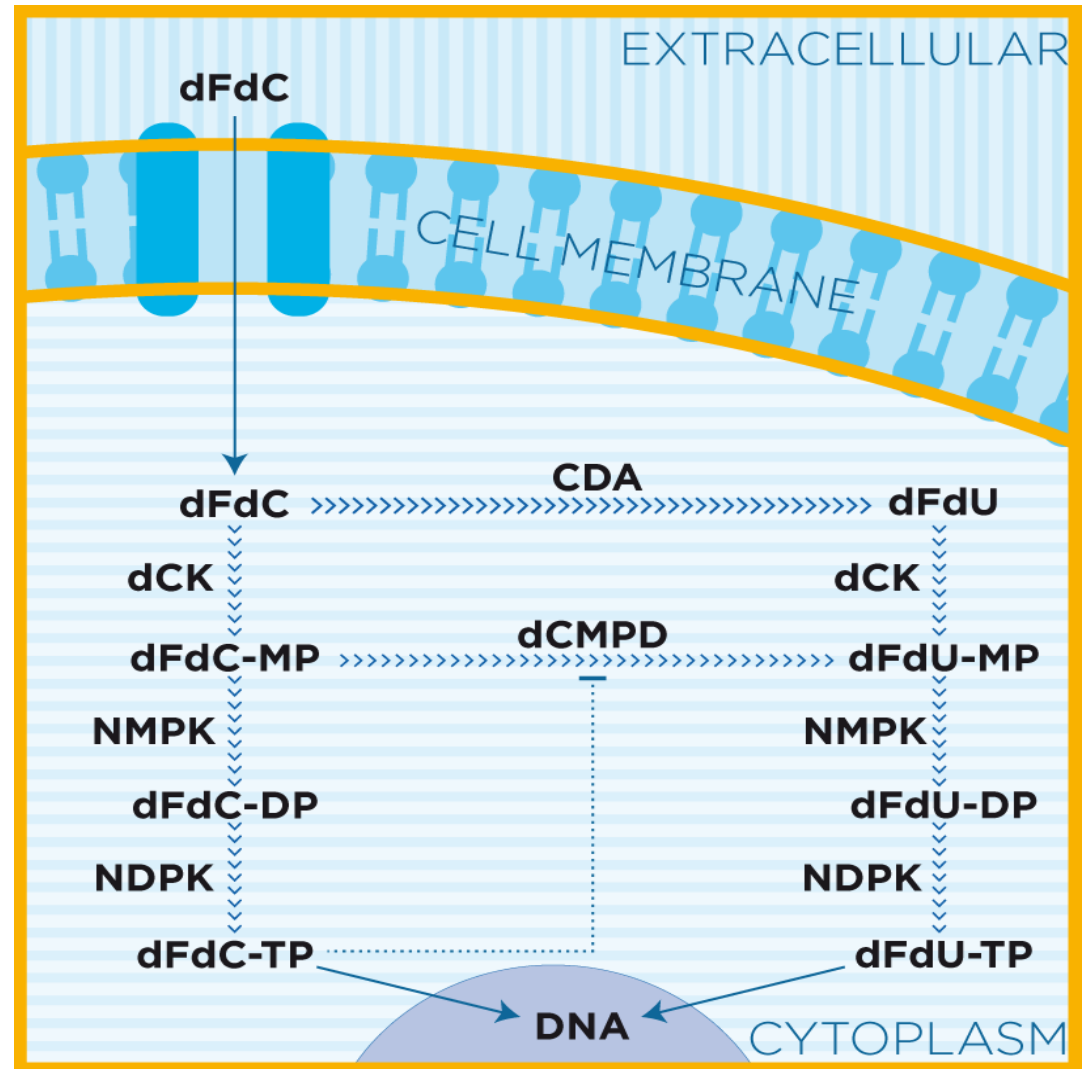
Component Site Definition Component Configurations Monomolecular Dynamics

Site Name	Description
Click here to add new item	
nh	Deamination
ph1	First phosphorylation
ph2	Second phosphorylation
ph3	Third phosphorylation
bin	Inhibition of dCMPD

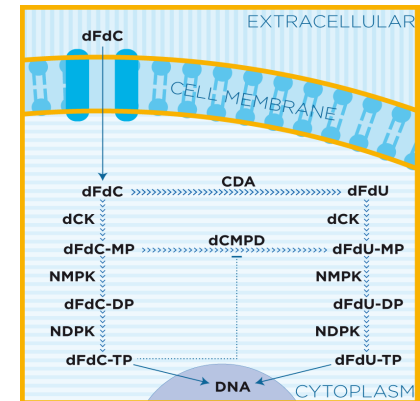
Diagram illustrating the transport and phosphorylation of dFdC across a cell membrane. The diagram shows the extracellular space, cell membrane, and cytoplasm. dFdC is shown in the extracellular space, crossing the cell membrane into the cytoplasm. In the cytoplasm, dFdC is phosphorylated by NMPK to dFdC-MP, then by NDPK to dFdC-DP, and finally to dFdC-TP. dFdC-TP is shown binding to DNA. The diagram also shows the conversion of dFdC to dFdu by CDA, and the subsequent phosphorylation of dFdu to dFdu-MP, dFdu-DP, and dFdu-TP by NMPK and NDPK.

COMPARTMENTS

The extracellular medium, the cellular membrane, and the intracellular medium (cytosol) are considered;



COMPARTMENTS

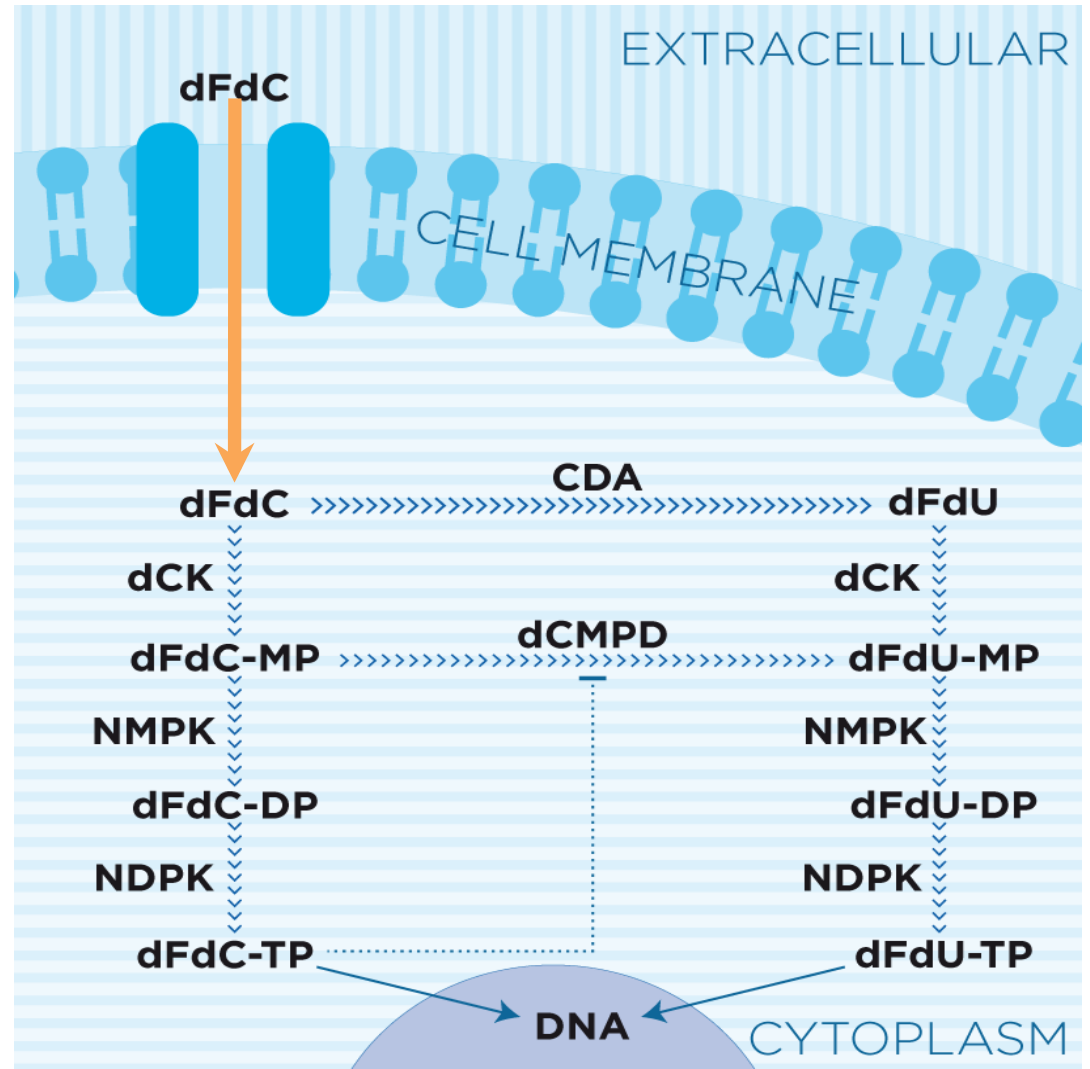


Options	Name	Type	Enclosed in	Volume	Description
▲ [+]	System	Compartment		System_Volume	
▲ [✕] [+]	Membrane	Membrane	System	Membrane_Volume	
> [✕] [+]	Cytosol	Compartment	Membrane	Cytosol_Volume	

Type	Kinetic Law	Name	Value	Unit of Measure	Description
+ Click here to add new item					
[✕] Volume		System_Volume	50	fl (10 ⁻⁹ mm ³)	
[✕] Volume		Membrane_Volume	0.1	fl (10 ⁻⁹ mm ³)	
> [✕] Volume		Cytosol_Volume	17	fl (10 ⁻⁹ mm ³)	

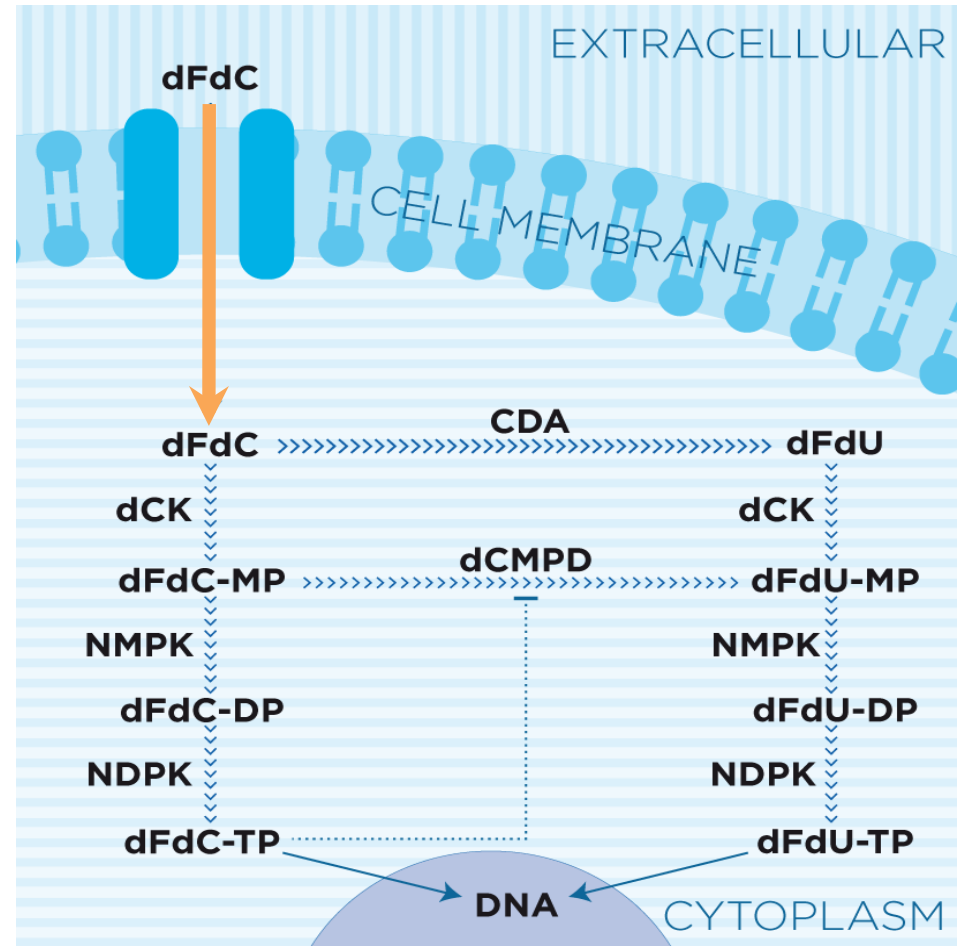
TRANSLOCATIONS

Gemcitabine moves from plasma into the cell passing through the cellular membrane;



TRANSLOCATIONS

Gemcitabine moves from plasma into the cell passing through the cellular membrane;



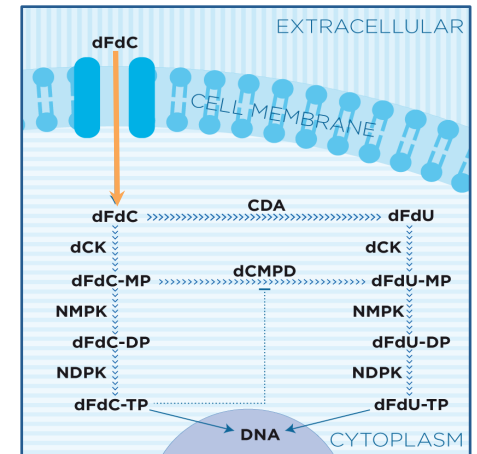
BlenX code:

```
let p_main : pproc =
```

```
  if (s1, c_dFdCout1) then ch(rate(r_dFdCout_in), s1, c_dFdC1).ch(s2, c_dFdC2).start_p_main!()
  endif + ...
```

```
let dFdCout: bproc = #(s1, c_dFdCout1), #(s2, c_dFdCout2) [ p_main | rep start_p_main?().p_main ];
```

TRANSLOCATIONS

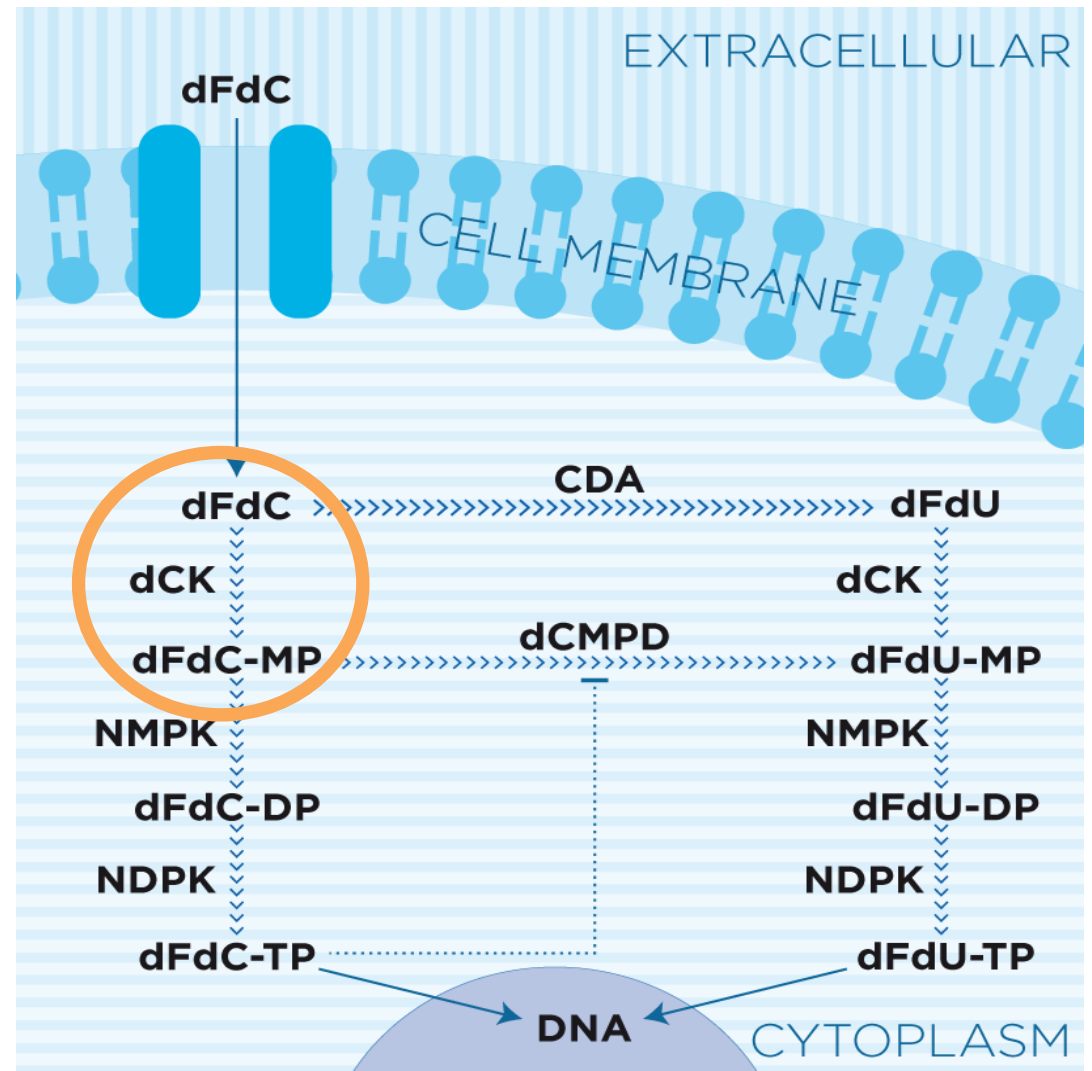


Components	Complexes	Global Dynamics	Bimolecular Dynamics	Binding Dynamics	Translocations	Compartments	Parameters	Initial Sta
		Action		(Parameter) Rate	(Parameter) Back Rate	Description		
Click here to add new item								
>	✖	dFdC_init moves from System to Cytoplasm		r_dFdCout_in				

Components	Complexes	Global Dynamics	Bimolecular Dynamics	Binding Dynamics	Translocations	Compartments	Parameters	Initial
		Type	Kinetic Law	Name	Value	Unit of Measure	Description	
Click here to add new item								
	✖	Volume		System_Volume	50	fl (10 ⁻⁹ mm ³)		
	✖	Volume		Membrane_Volume	0.1	fl (10 ⁻⁹ mm ³)		
	✖	Volume		Cytosol_Volume	17	fl (10 ⁻⁹ mm ³)		
>	✖	Rate	Mass Action	r_dFdCout_in	9.97	1/hours		

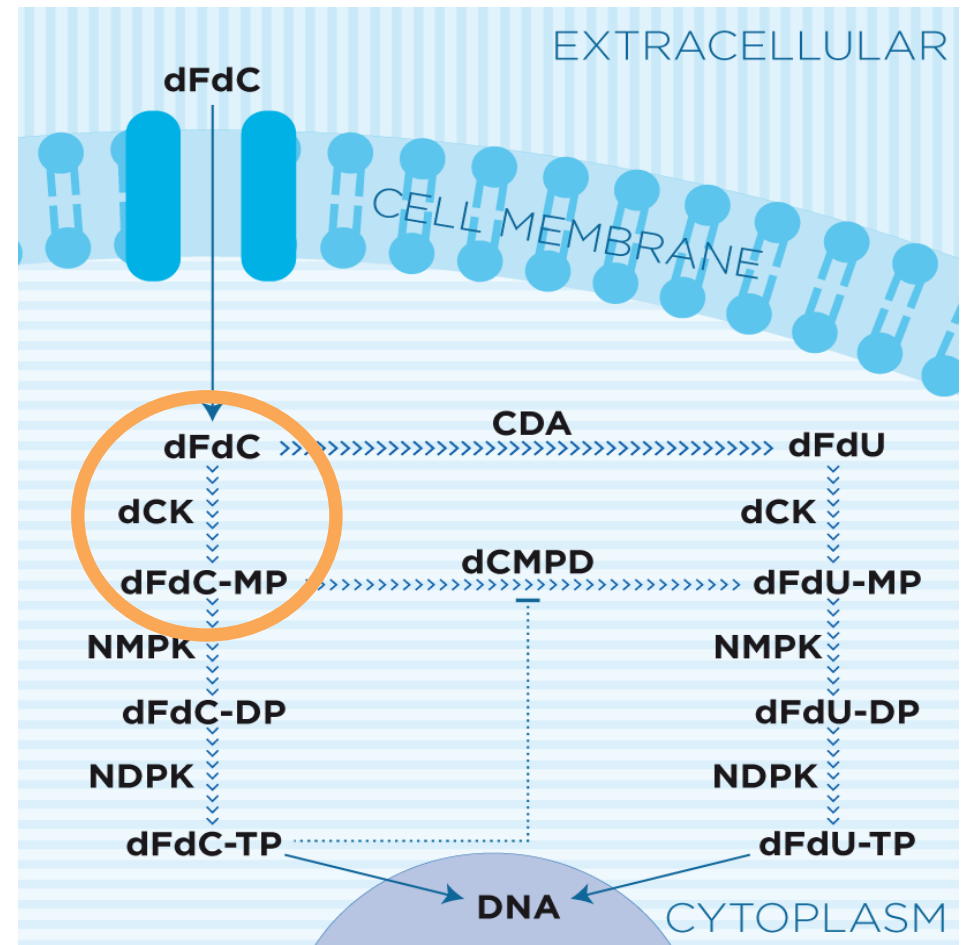
BIMOLECULAR DYNAMICS

- Once inside the cytoplasm, dFdc is phosphorylated by deoxycytidine kinase (dCK);
- Built-in kinetic laws are available (Mass-Action, Michaelis-Menten, Hill) as well as User Defined;



BIMOLECULAR DYNAMICS

- Once inside the cytoplasm, dFdC is phosphorylated by deoxycytidine kinase (dCK);
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BlenX code:

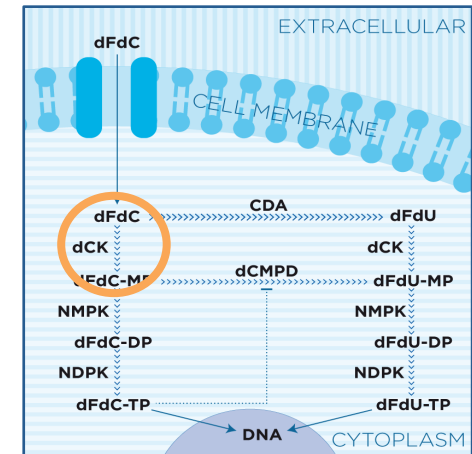
```
let p_main : pproc = ...+if (s1, c_dFdC1) then s1?().ch(s1, c_dFdCMP1).ch(s2, c_dFdCMP2).start_p_main!() +
    endif + ...
```

```
let dFdC : bproc = #(s1, c_dFdC1), #(s2, c_dFdC2) [ p_main | rep start_p_main?().p_main ];
```

```
let dCK : bproc = #(s, c_dCK) [ s!().startdCK!() | rep startdCK?().s!().startdCK!() ];
```

Affinity: (c_dCK, c_dFdC1, rate(r_dFdC_dCK))

BIMOLECULAR DYNAMICS

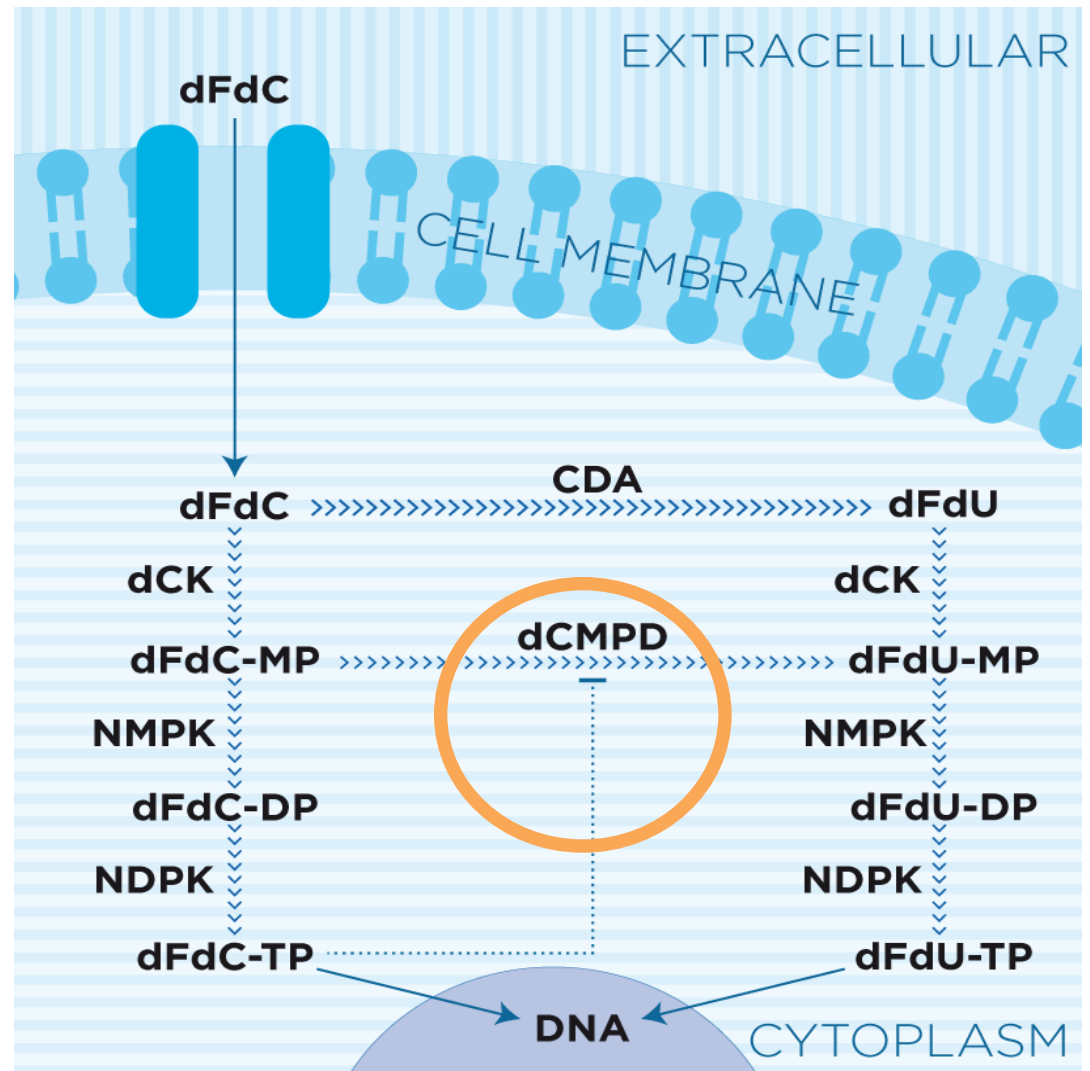


Components	Complexes	Global Dynamics	Bimolecular Dynamics	Binding Dynamics	Translocations	Compartments	Parameters	Initial S
Action		(Parameter) Rate	Active Condition	Passive Condition				
Click here to add new item								
>	<input type="checkbox"/>	dCK makes dFdc Phosphorylated on ph1	r_dFdc_dCK		(nh is Aminated and ph1 is Free)			

Components	Complexes	Global Dynamics	Bimolecular Dynamics	Binding Dynamics	Translocations	Compartments	Parameters	Ini
Type	Kinetic Law	Name	Value	Unit of Measure	Description			
Click here to add new item								
<input type="checkbox"/>	Volume		System_Volume	50	fl (10 ⁻⁹ mm ³)			
<input type="checkbox"/>	Volume		Membrane_Volume	0.1	fl (10 ⁻⁹ mm ³)			
<input type="checkbox"/>	Volume		Cytosol_Volume	17	fl (10 ⁻⁹ mm ³)			
<input type="checkbox"/>	Rate	Mass Action	r_dFdcOut_in	9.97	1/hours			
>	<input type="checkbox"/>	Rate	Mass Action	r_dFdc_dCK	0.01	1/(Units*hours)		

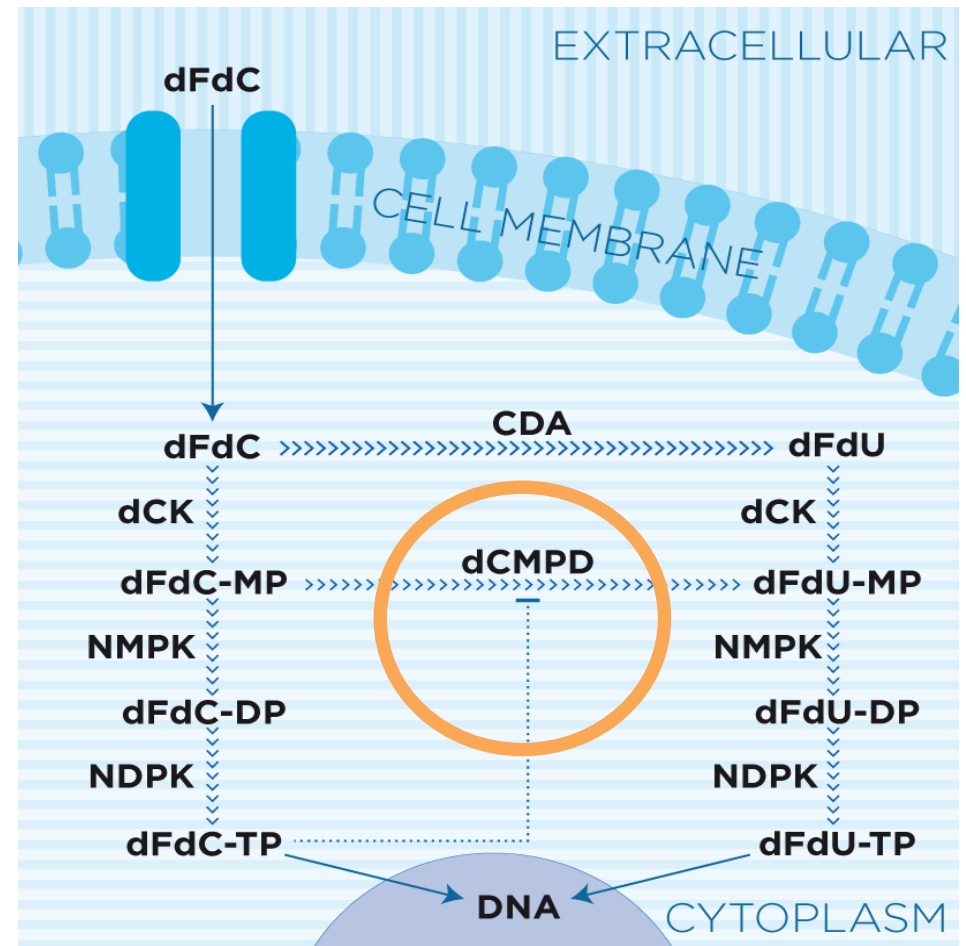
BINDING DYNAMICS

- Inhibition mechanisms can be modeled in different ways, in this case we considered an inhibitory complexation between dFdCTP and dCMPD;
- Binding conditions are specified to identify the target configuration.



BINDING DYNAMICS

- Inhibition mechanisms can be modeled in different ways, in this case we considered an inhibitory complexation between dFdCTP and dCMPD;
- Binding conditions are specified to identify the target configuration.



BlenX code:

```
let dFdCTP : bproc = #(s1, c_dFdCTP1), #(s2, c_dFdCTP2) [ p_main | rep start_p_main?().p_main ];
```

```
let dCMPD : bproc = #(s, c_dCMPD) [ s!().startdCMPD!() | rep startdCMPD?().s!().startdCMPD!() ];
```

```
Affinity: (c_dFdCTP1, c_dCMPD, rate(r_dFdCTP_dCMPD_bin), rate(r_dFdCTP_dCMPD_unb), 0)
```

BINDING DYNAMICS

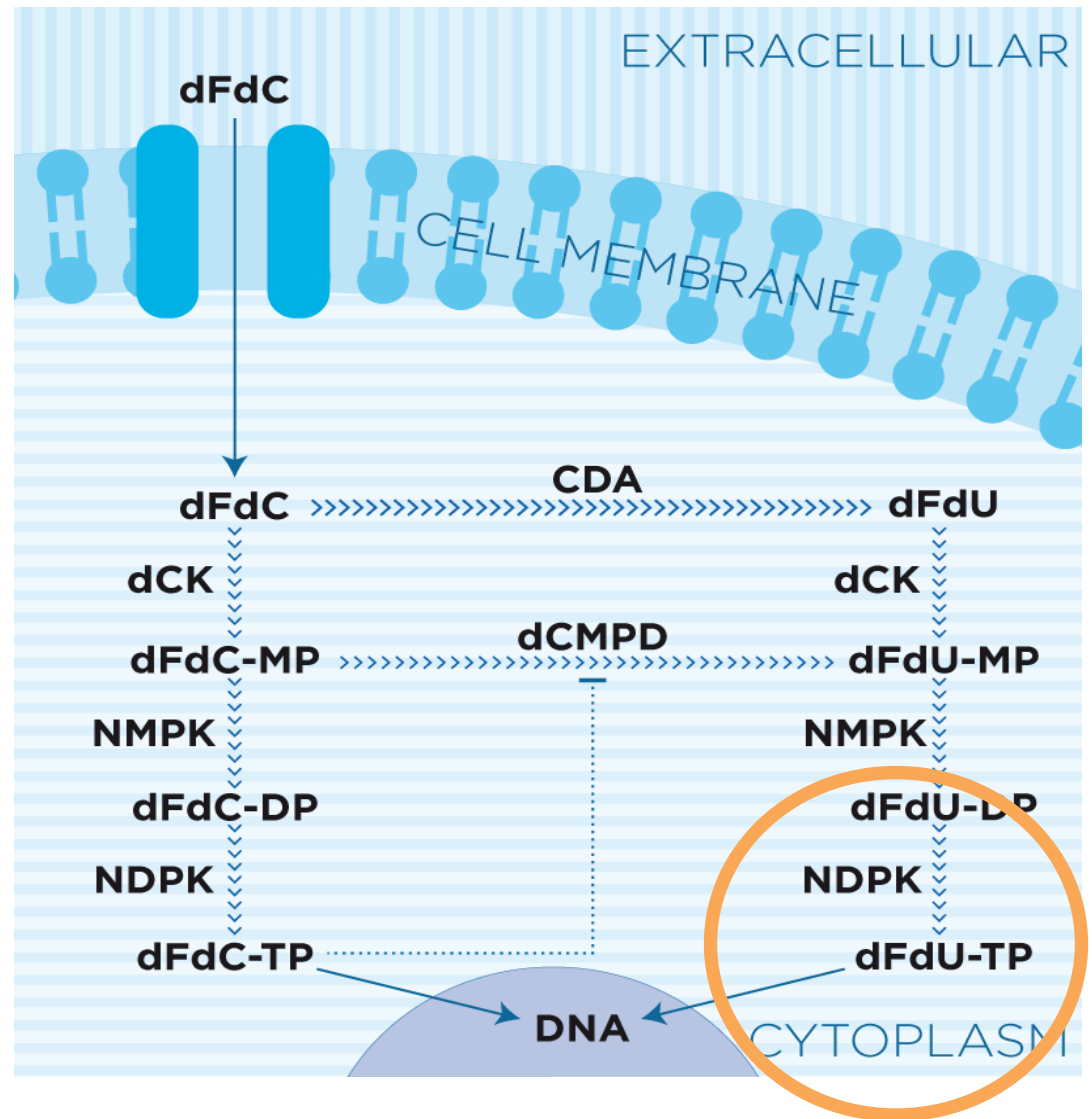
Components	Complexes	Global Dynamics	Bimolecular Dynamics	Binding Dynamics	Translocations	Compartments	Parameters	Initial State	
		Component 1	Site 1	Component 2	Site 2	(Parameter) Binding Rate	(Parameter) Unbinding Rate	Condition	Comp
Click here to add new item									
		dFdC	bin	dCMPD	bin	r_dFdCTP_dCMPD_bin	r_dFdCTP_dCMPD_unb	Binding	Cytos
Conditions					Modifiers				
Component 1					Component 2				
Binding Condition for Component 1					Binding Condition for Component 2				
(nh is Aminated and ph3 is Phosphorylated)									
Unbinding Condition for Component 1					Unbinding Condition for Component 2				

Components	Complexes	Global Dynamics	Bimolecular Dynamics	Binding Dynamics	Translocations	Compartments	Parameters	Initial State
Type	Kinetic Law		Name	Value	Unit of Measure	Description		
Click here to add new item								
	Volume		System_Volume	50	fl (10 ⁻⁹ mm ³)			
	Volume		Membrane_Volume	0.1	fl (10 ⁻⁹ mm ³)			
	Volume		Cytosol_Volume	17	fl (10 ⁻⁹ mm ³)			
	Rate	Mass Action	r_dFdCout_in	9.97	1/hours			
	Rate	Mass Action	r_dFdC_CDA	0.000005	1/(Units*hours)			
	Rate	Mass Action	r_dFdCMP_dCMPD	0.00005	1/(Units*hours)			
	Rate	Mass Action	r_dFdU_dCK	0.01	1/(Units*hours)			
	Rate	Mass Action	r_dFdUMP_NMPK	0.001	1/(Units*hours)			
	Rate	Mass Action	r_dFdUDP_NDPK	0.002	1/(Units*hours)			
	Rate	Mass Action	r_dFdCTP_dCMPD_bin	1E-02	1/(Units*hours)			
	Rate	Mass Action	r_dFdCTP_dCMPD_unb	1E-08	1/(Units*hours)			

The diagram illustrates the binding dynamics of dFdC. It shows the EXTRACELLULAR space above the CELL MEMBRANE and the CYTOPLASM below. dFdC molecules are shown binding to receptors on the membrane. This triggers the activation of CDA (Cytidine Deaminase), which converts dFdU to dFdU-MP, then dFdU-DP, and finally dFdU-TP. dFdU-TP then binds to DNA in the cytoplasm. The diagram also shows the conversion of dFdC to dFdC-MP and dFdC-DP, and the conversion of dFdU to dFdU-MP and dFdU-DP. The diagram is labeled with 'EXTRACELLULAR', 'CELL MEMBRANE', 'CYTOPLASM', and 'DNA'.

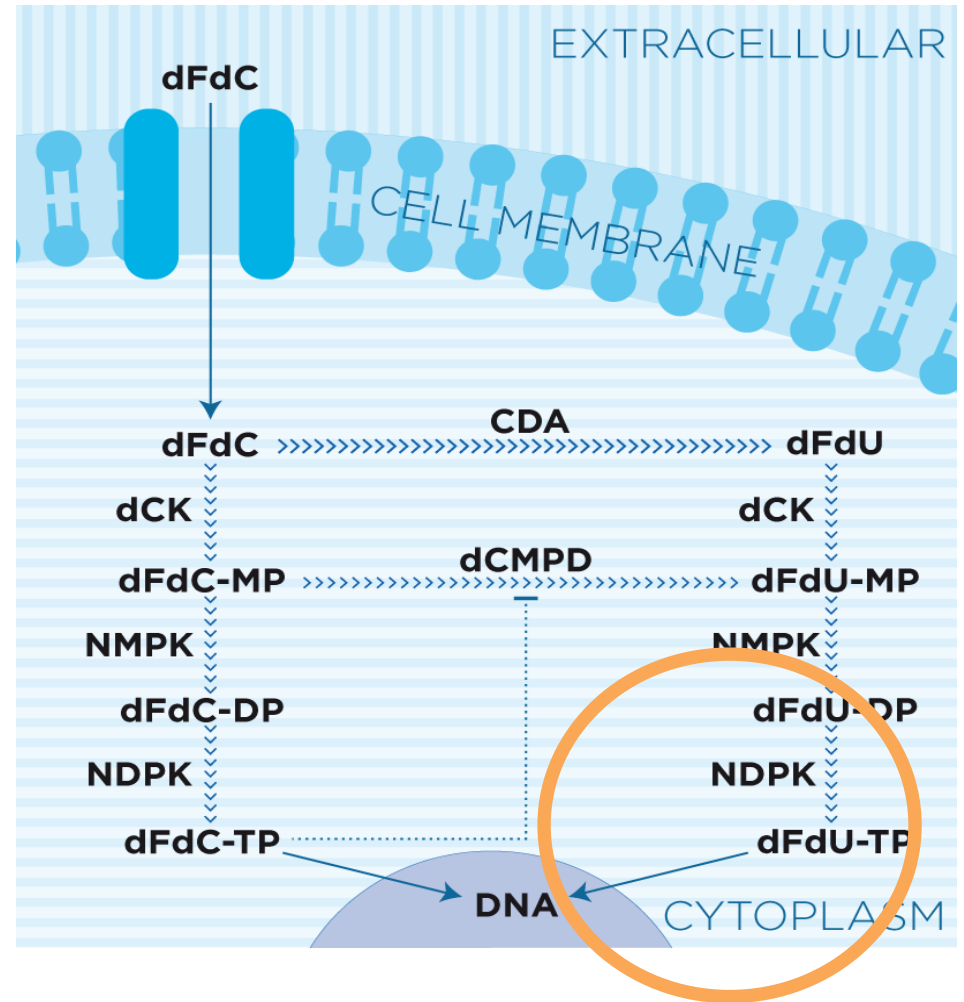
GLOBAL DYNAMICS

Gemcitabine triphosphate (dFdCTP and dFdUTP) is incorporated into DNA causing chain termination.



GLOBAL DYNAMICS

Gemcitabine triphosphate (dFdCTP and dFdUTP) is incorporated into DNA causing chain termination.

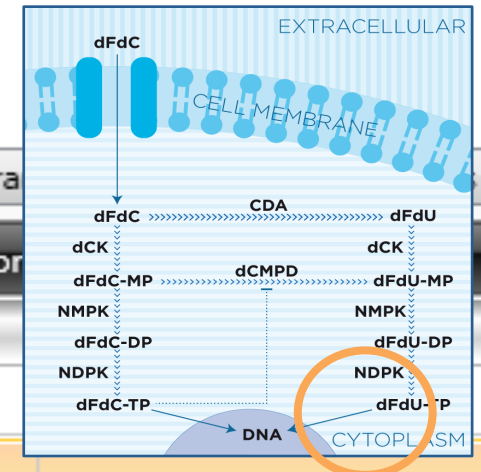


BlenX code:

```
let dFdUTP : bproc = #(s1, c_dFdUTP1), #(s2, c_dFdUTP2) [ p_main | rep start_p_main?().p_main ];  
when (dFdUTP :: rate(r_dFdUTP_DNA)) split(DNA, Nil);
```

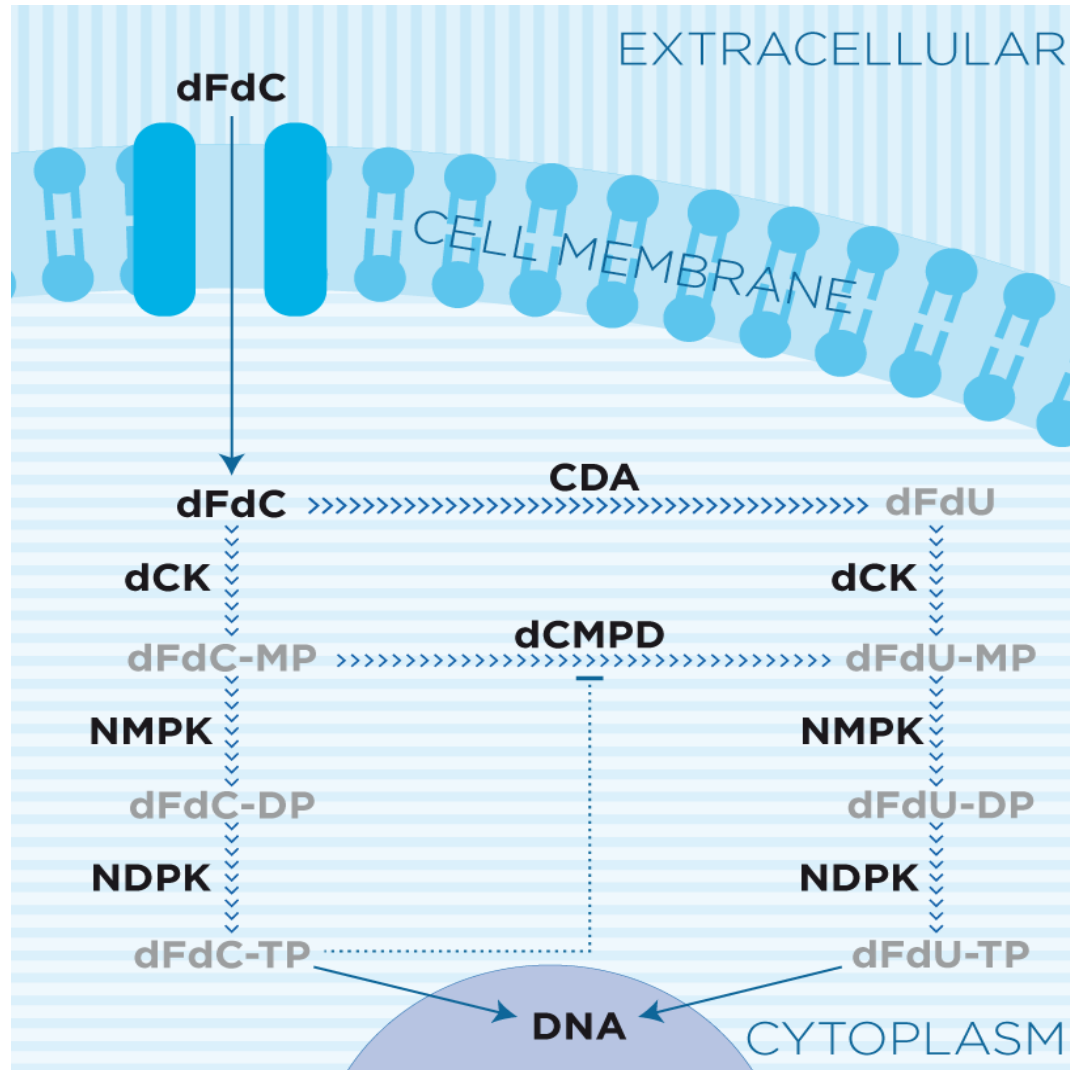
GLOBAL DYNAMICS

Components		Complexes		Global Dynamics		Bimolecular Dynamics		Binding Dynamics		Tra	
	Action	(Parameter) Rate	Condition	Compartment	Description						
+>	Click here to add new item										
	<input type="checkbox"/> substitute dFdCTP with DNA	r_dFdCTP_DNA		Cytosol							
>	<input checked="" type="checkbox"/> substitute dFdUTP with DNA	r_dFdUTP_DNA		Cytosol							

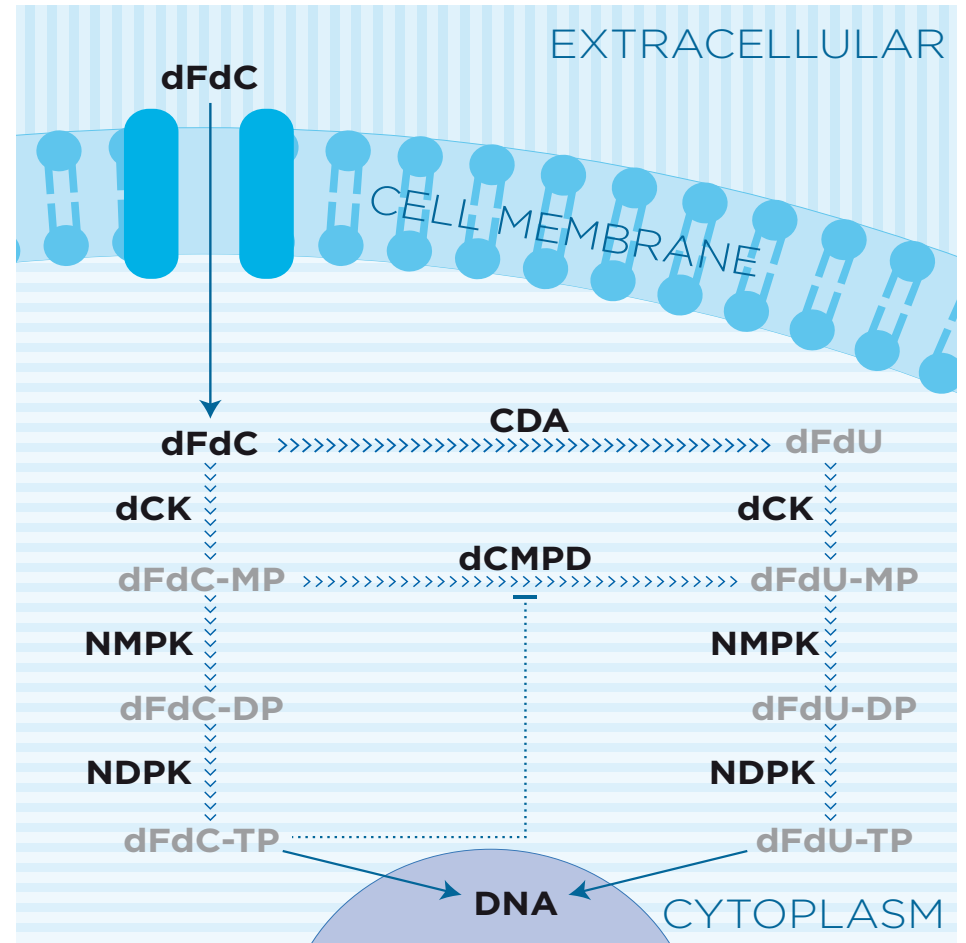


<input checked="" type="checkbox"/>	Volume			Membrane_Volume	0.1	fl (10 ⁻⁹ mm ³)	
<input checked="" type="checkbox"/>	Volume			Cytosol_Volume	17	fl (10 ⁻⁹ mm ³)	
<input checked="" type="checkbox"/>	Rate	Mass Action		r_dFdCout_in	9.97	1/hours	
<input checked="" type="checkbox"/>	Rate	Mass Action		r_dFdC_CDA	0.000005	1/(Units*hours)	
<input checked="" type="checkbox"/>	Rate	Mass Action		r_dFdCMP_dCMPD	0.00005	1/(Units*hours)	
<input checked="" type="checkbox"/>	Rate	Mass Action		r_dFdU_dCK	0.01	1/(Units*hours)	
<input checked="" type="checkbox"/>	Rate	Mass Action		r_dFdUMP_NMPK	0.001	1/(Units*hours)	
<input checked="" type="checkbox"/>	Rate	Mass Action		r_dFdUDP_NDPK	0.002	1/(Units*hours)	
<input checked="" type="checkbox"/>	Rate	Mass Action		r_dFdCTP_dCMPD_bin	1E-02	1/(Units*hours)	
<input checked="" type="checkbox"/>	Rate	Mass Action		r_dFdCTP_dCMPD_unb	1E-08	1/hours	
>	<input checked="" type="checkbox"/> Rate	Mass Action		r_dFdUTP_DNA	0.001	1/hours	

INITIAL STATE



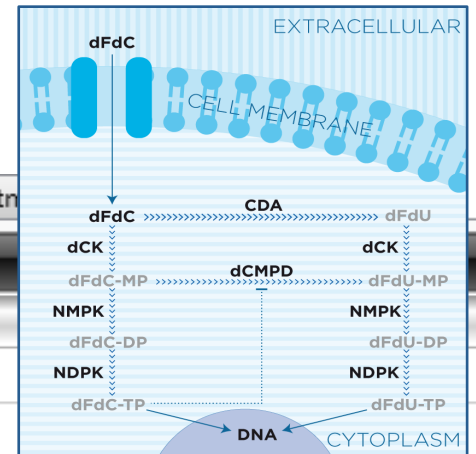
INITIAL STATE



BlenX code:

```
run 10000 dFdCout || 100 dCK || 100 NMPK || 100 NDPK || 100 CDA || 100 dCMPD
```

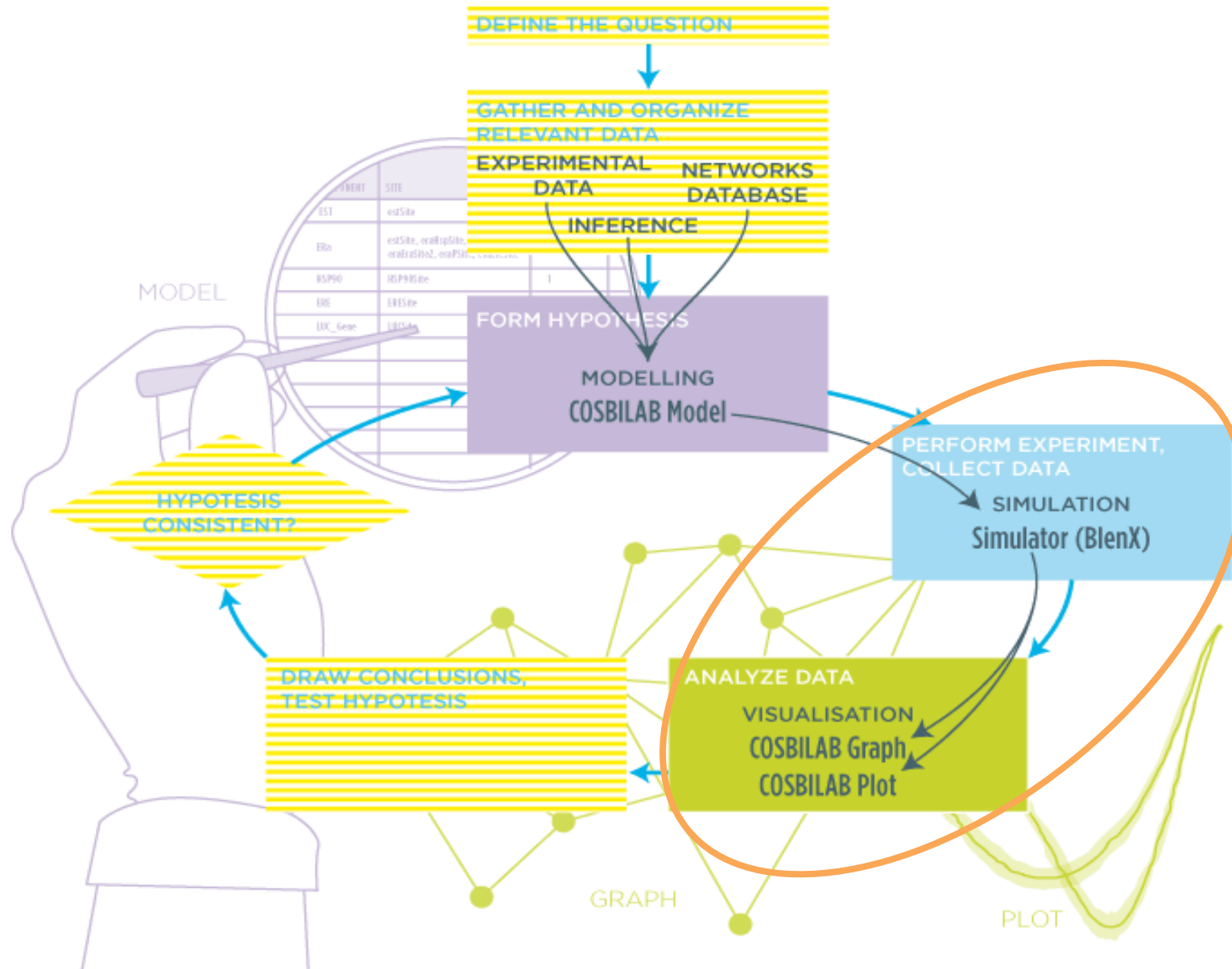
INITIAL STATE



Components	Complexes	Global Dynamics	Bimolecular Dynamics	Binding Dynamics	Translocations	Compartments
Component/Complex Configuration	Type	(Parameter) Quantity	Compartment	Description		
Click here to add new item						
<input type="checkbox"/>	dFdC_init	Component	q_dFdC_out	System		
<input type="checkbox"/>	dCK_init	Component	q_enzyme	Cytosol		
<input type="checkbox"/>	NMPK_init	Component	q_enzyme	Cytosol		
<input type="checkbox"/>	NDPK_init	Component	q_enzyme	Cytosol		
<input type="checkbox"/>	CDA_init	Component	q_enzyme	Cytosol		
<input checked="" type="checkbox"/>	dCMPD_init	Component	q_enzyme	Cytosol		

Components	Complexes	Global Dynamics	Bimolecular Dynamics	Binding Dynamics	Translocations	Compartments	Parameters	Initial State
Type	Kinetic Law	Name	Value	Unit of Measure	Description			
Click here to add new item								
<input type="checkbox"/>	Volume		System_Volume	50	fl (10 ⁻⁹ mm ³)			
<input type="checkbox"/>	Rate	Mass Action	r_dFdCTP_dCMPD_bin	1E-02	1/(Units*hours)			
<input type="checkbox"/>	Rate	Mass Action	r_dFdCTP_dCMPD_unb	1E-08	1/hours			
<input type="checkbox"/>	Rate	Mass Action	r_dFdCTP_DNA	0.05	1/hours			
<input type="checkbox"/>	Rate	Mass Action	r_dFdUTP_DNA	0.001	1/hours			
<input type="checkbox"/>	Quantity		q_dFdC_out	10000	Units			
<input checked="" type="checkbox"/>	Quantity		q_enzyme	100	Units			

COSBI WORKING FLOW



SIMULATION

COSBI LAB Simulation Wizard 1.0.0

Select Parameter to Scan

Simulation Path
Simulation Arguments
Parameter Scan
Configure Parameters
Set Environment
Number of Runs
Run Simulation

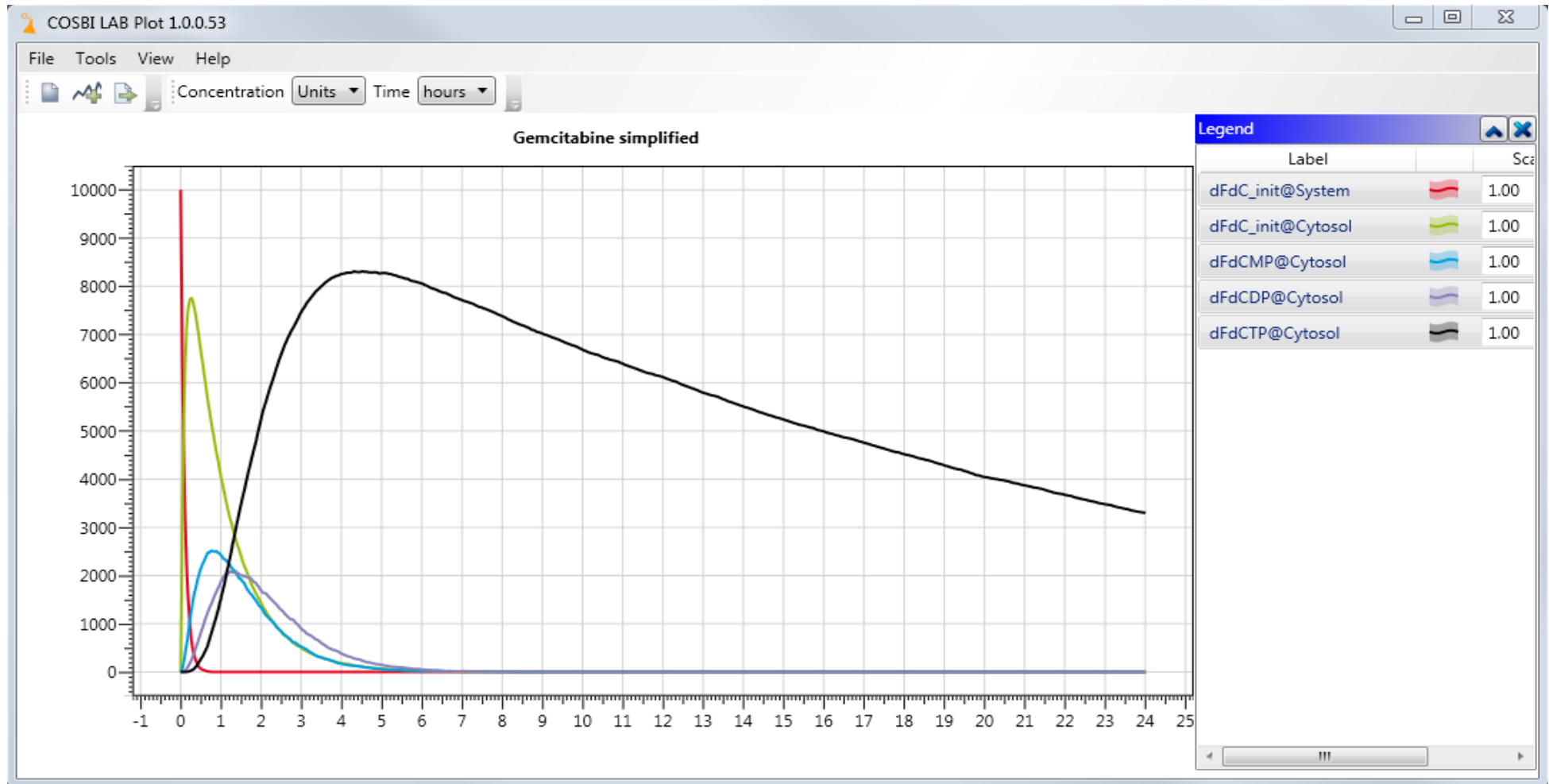
Scan	Parameter	Type	Unit of Measure
<input type="checkbox"/>	System_Volume	Volume	fl (10 ⁻⁹ mm ³)
<input type="checkbox"/>	Membrane_Volume	Volume	fl (10 ⁻⁹ mm ³)
<input type="checkbox"/>	Cytosol_Volume	Volume	fl (10 ⁻⁹ mm ³)
<input checked="" type="checkbox"/>	r_dFdCout_in	Rate	1/hours
<input type="checkbox"/>	r_dFdC_dCK	Rate	1/(Units*hours)
<input type="checkbox"/>	r_dFdCMP_NMPK	Rate	1/(Units*hours)
<input type="checkbox"/>	r_dFdCDP_NDPK	Rate	1/(Units*hours)
<input type="checkbox"/>	r_dFdC_CDA	Rate	1/(Units*hours)

1/hours]

Back Next Cancel

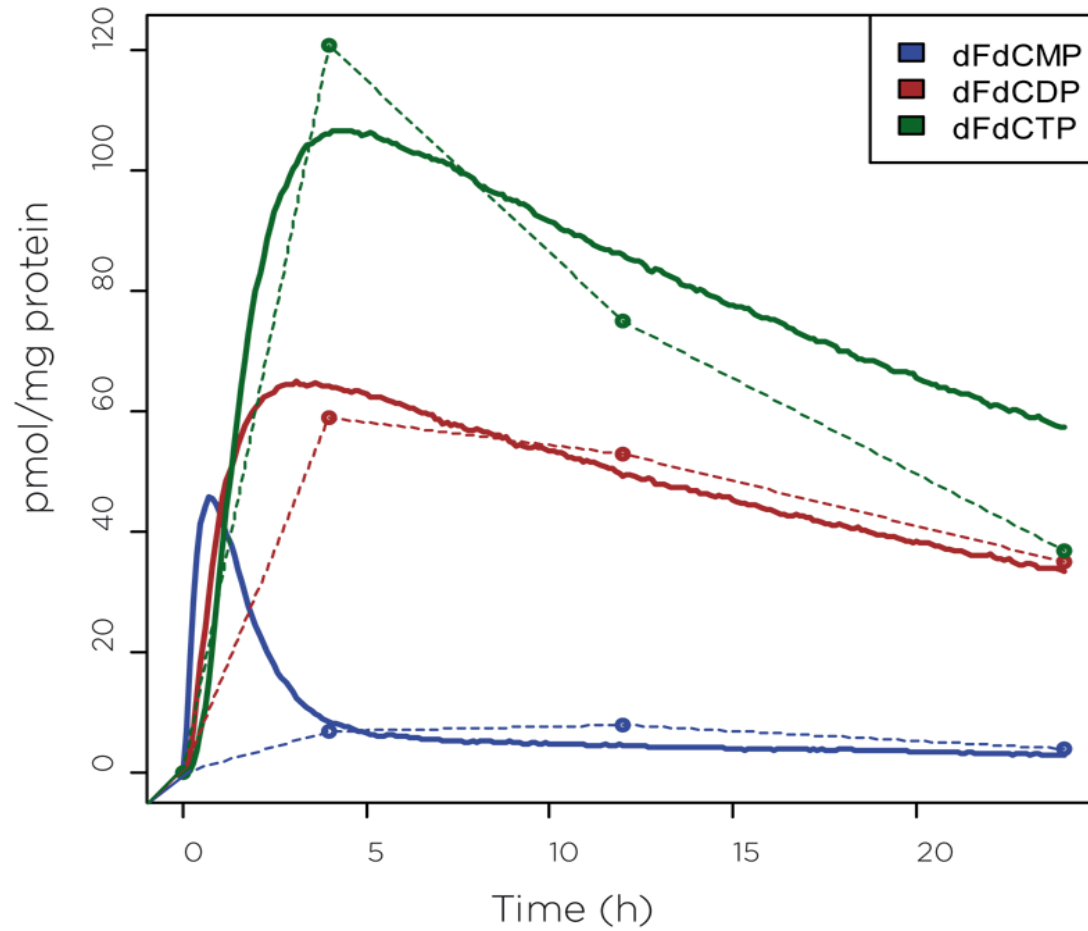
Back Next Cancel

VISUALIZE RESULTS



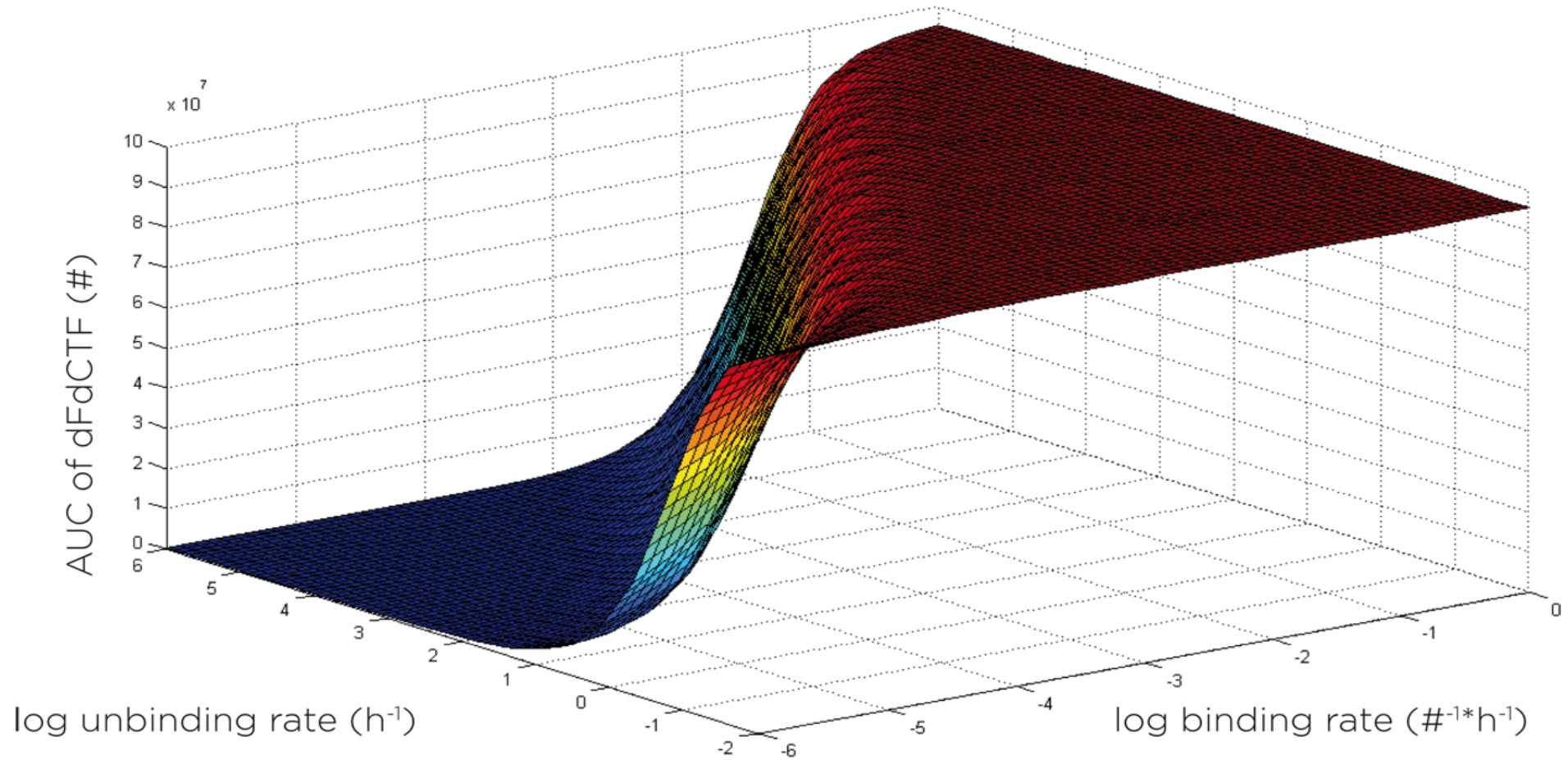
SIMULATION RESULTS PLOT

Intracellular concentration versus time

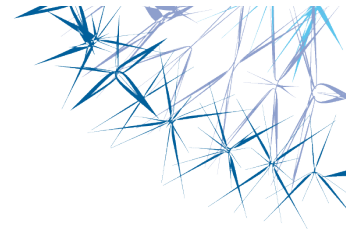


SENSITIVITY ANALYSIS

Accumulation of dFdCTP w.r.t. dCTP inhibition on dCK



COSBILAB MODEL: MODELING & SIMULATING COMPLEX SYSTEMS



Create models via tabular interface.
No expertise in programming nor math needed

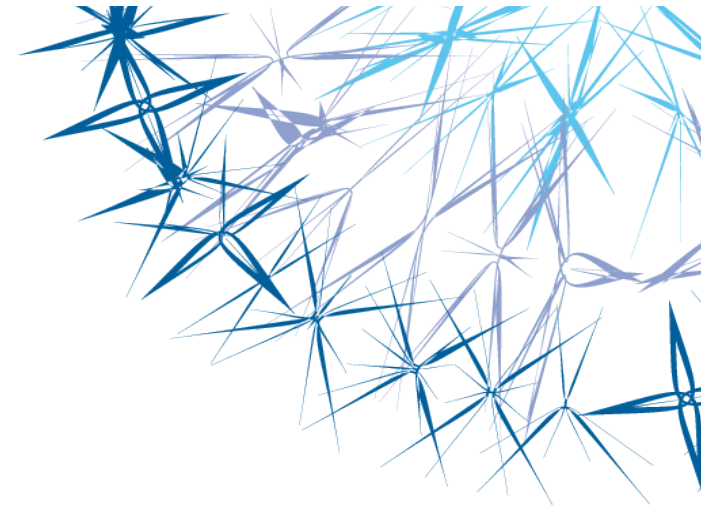
Easily manage experiments: knock-down genes, change rates, add a “virtual” drug.

Share models & results with colleagues, wherever they are.

Allow life scientist analyze models

In silico science: adding knowledge, saving time.

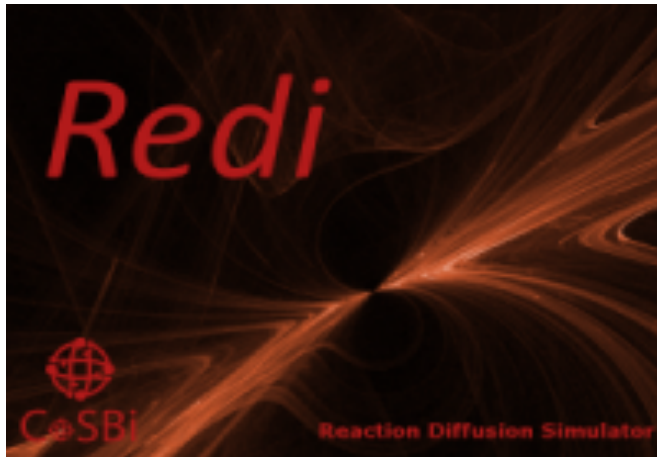
Accelerate discovery process, add value to your work.



Spatial simulation

P. Lecca, L. Dematte', A. Ihekwaba, C. Priami. Redi: a simulator of stochastic biochemical reaction-diffusion systems, *The Second International Conference on Advances in System Simulation (SIMUL 2010)*, 2010.

SPACE AND DIFFUSION

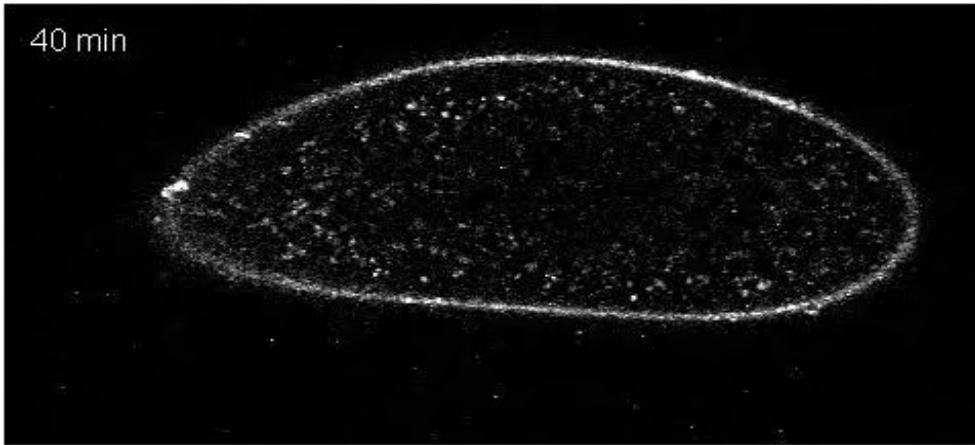


Reaction-diffusion simulator at the mesoscopic interaction scale.

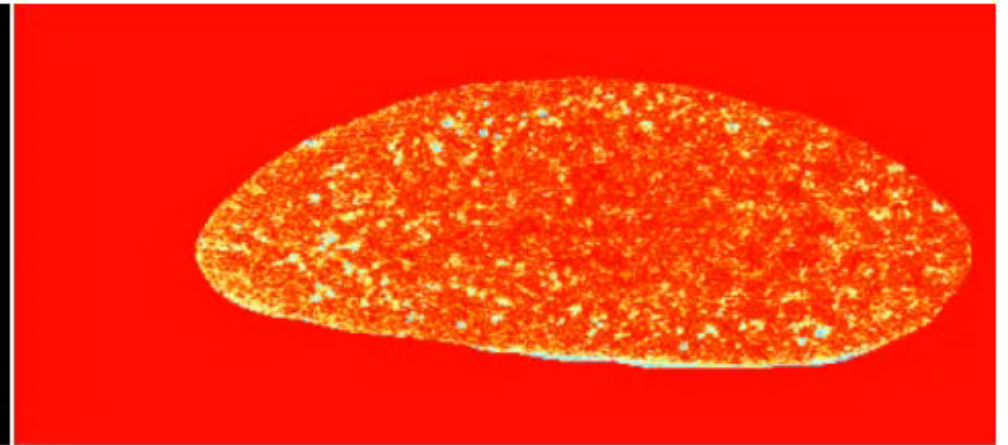
Space discretised variant of the Gillespie SSA.

Dynamically state-dependent diffusion coefficient.

BICOID SIMULATION

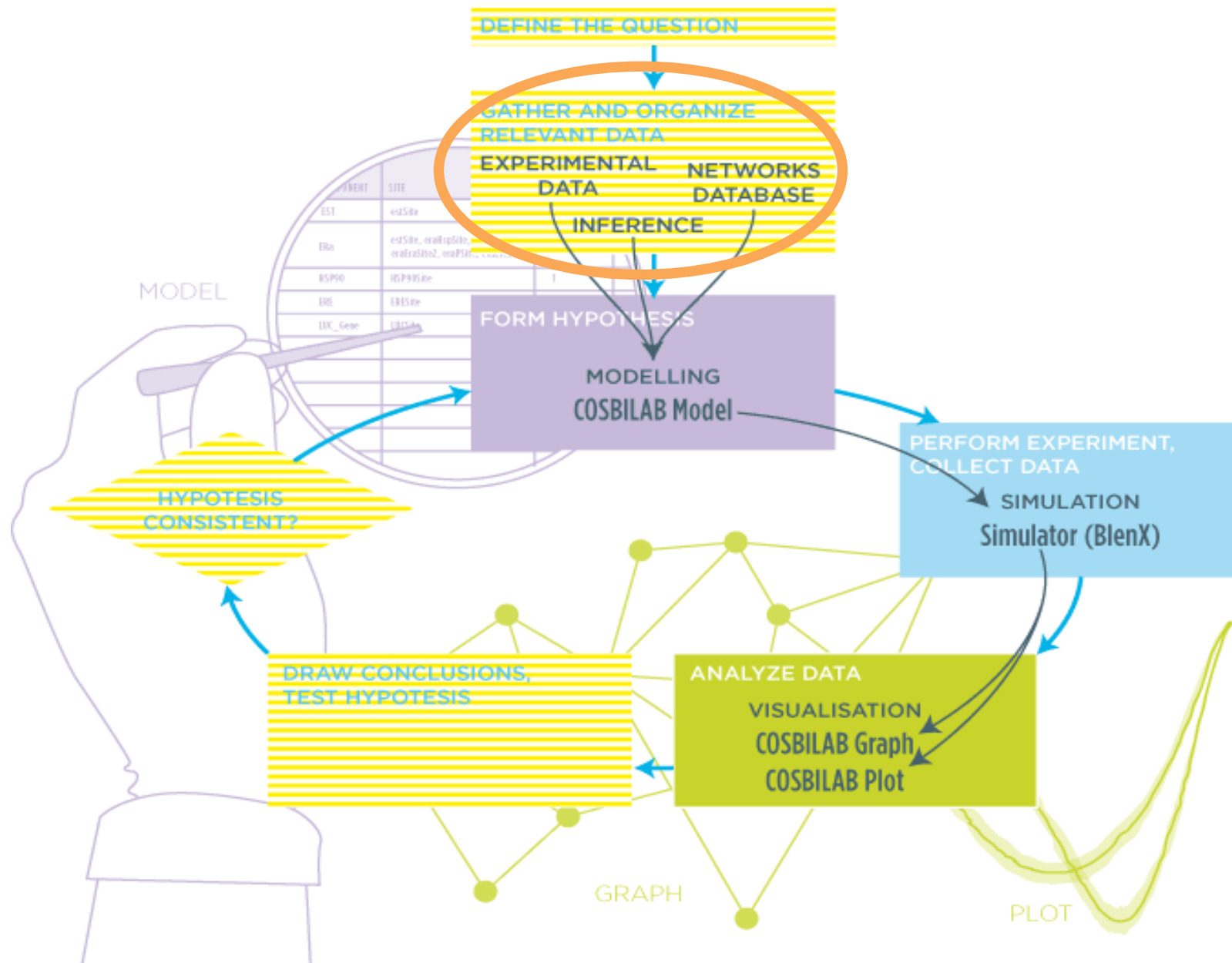


Experimental



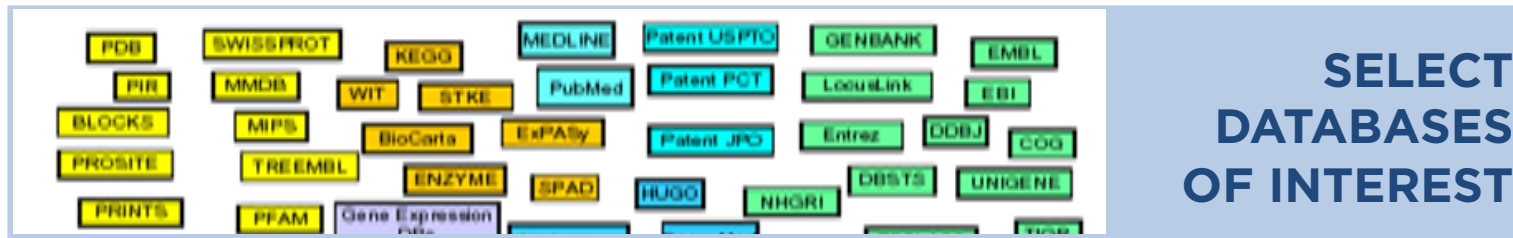
Simulated

COSBI WORKING FLOW



KNOWLEDGE EXTRACTION AND MODELING

Define the problem you want to address



Clinical - *omics* experiments - Ontological

DATA MINING

Organize, integrate and analyze data

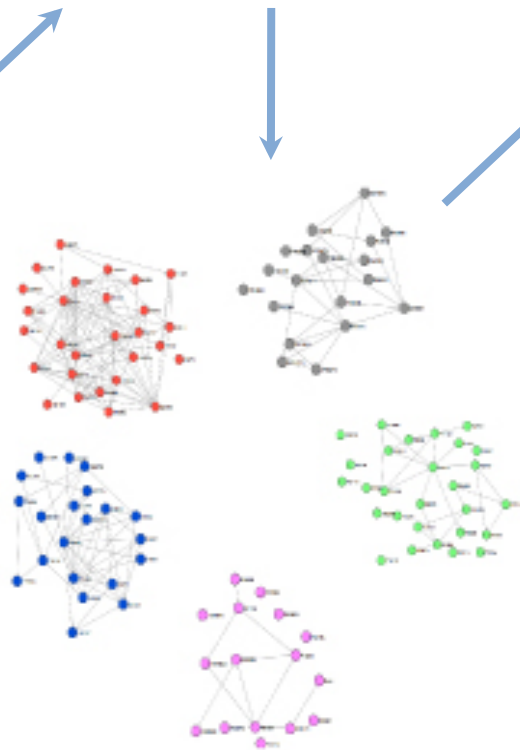
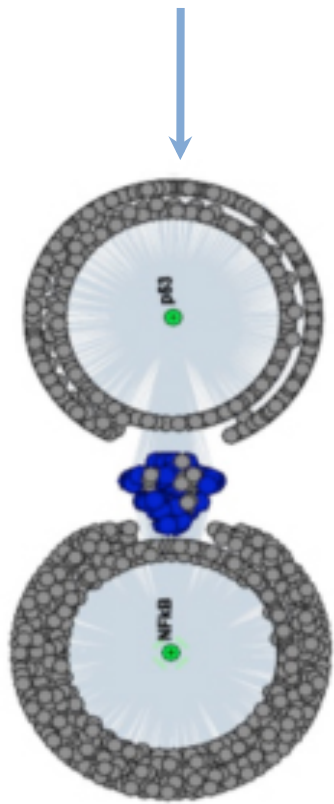
KNOWLEDGE EXTRACTION

KNOWLEDGE EXTRACTION AND MODELING

Model identification
Network inference

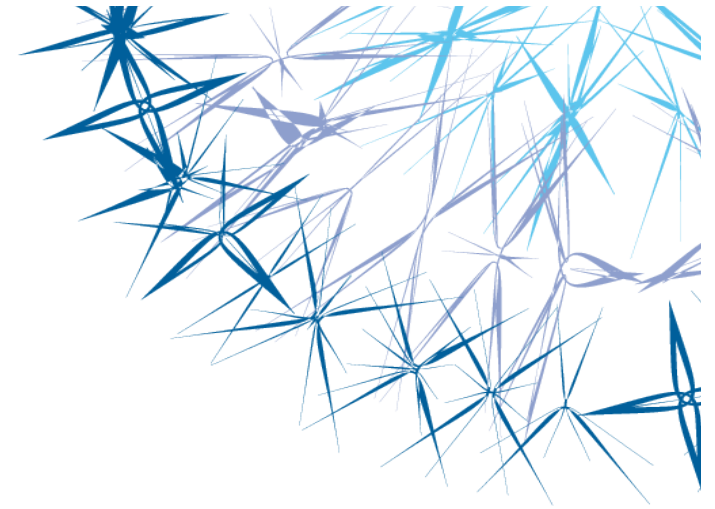
Model analysis
Network analysis

Model calibration
Parameter inference

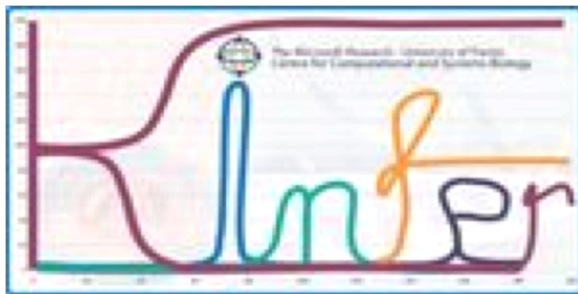


Parameter	Value ($\sigma = 0.008$)	Value ($\sigma = 0.01$)	Value ($\sigma = 0.04$)	Value ($\sigma = 0.19$)	Value ($\sigma = 1.66$)
k_1	0.0945	0.0954	0.0955	1.104	0.012
k_{-1}	0.164	0.146	0.140	0.024	0.726
k_2	3.609	3.640	3.0556	1.176	1.0526
k_{-2}	0.499	0.439	0.442	0.129	1.281
k_3	0.498	0.497	0.498	0.549	0.194
k_{-3}	0.127	0.123	0.144	0.254	0.074
k_4	0.554	0.557	0.577	0.552	0.444
k_{-4}	0.00183	0.00186	0.00256	0.0287	0.00668

Algorithmic
modeling



Knowledge inference



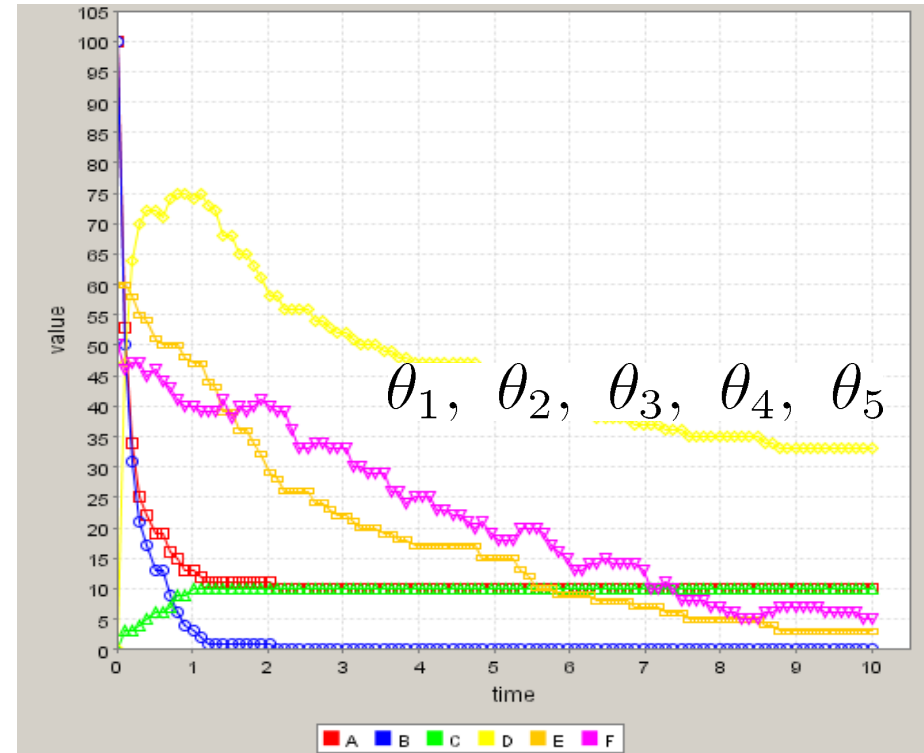
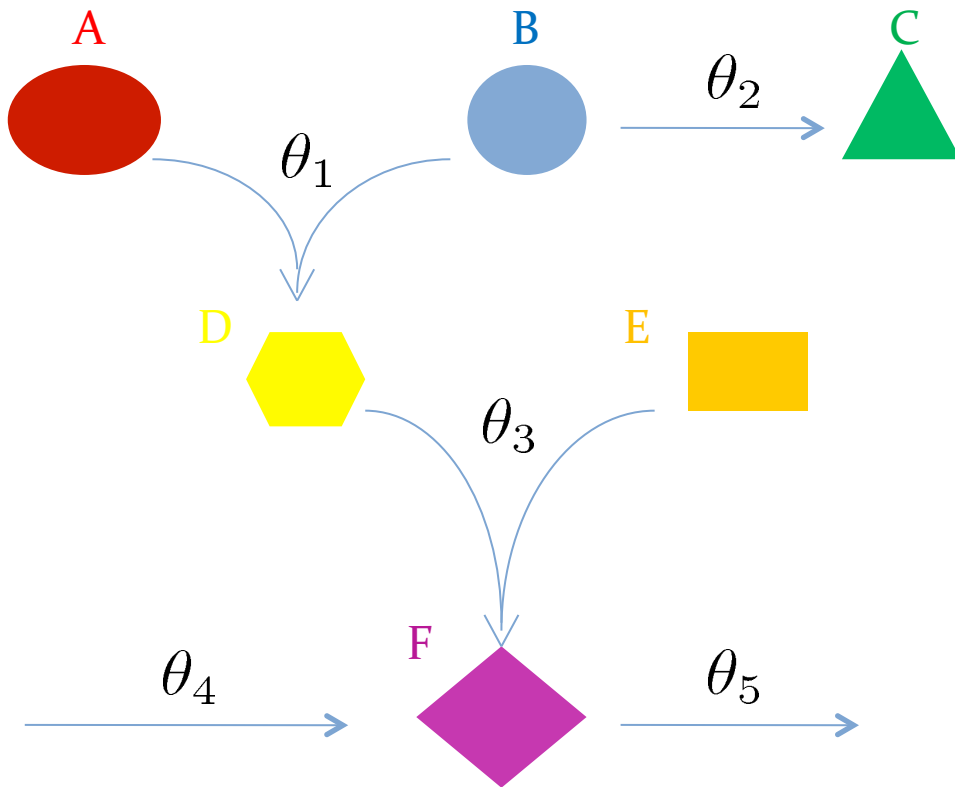
P. Lecca, A. Palmisano, C. Priami, G. Sanguinetti. *A new probabilistic generative model of parameter inference in biochemical networks*. Proceedings of the ACM Symposium on Applied Computing, 2009.

P. Lecca, A. Palmisano, A. Ihekweba, C. Priami. *Calibration of dynamic models of biological systems with KInfer*. **European Biophysics Journal**, 39:1019-1039, 2010

THE SYSTEM AND THE DATA

Experimentally observed time course data of the reactants concentrations.

From these behaviors we want to estimate



COMPUTATIONAL SUPPORT: KINFER

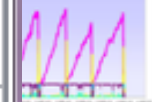
Loading time series of species concentrations

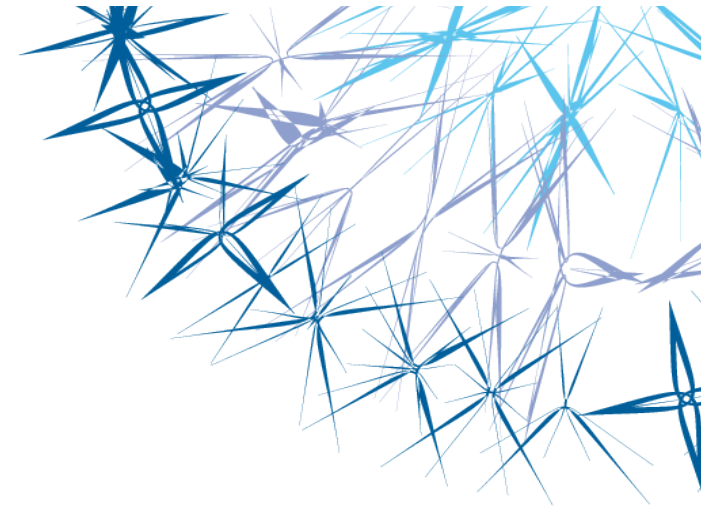
Estimation of initial guesses for parameters

The screenshot shows the KInfer software interface. The 'Initial values' tab is active, displaying a table of parameter estimates. The table has columns for parameter names and their estimated values. The parameters listed are k3, k4, sigma, k1, and k2. The estimated values are 12.9263, 0.006542645010828389, 0.749, 6.949, and 0.81774799, respectively. The 'Calculate!' button is visible next to the 'Error on concentration measure' field.

Parameter	Estimated Value
k3	12.9263
k4	0.006542645010828389
sigma	0.749
k1	6.949
k2	0.81774799

Time behaviour of the system



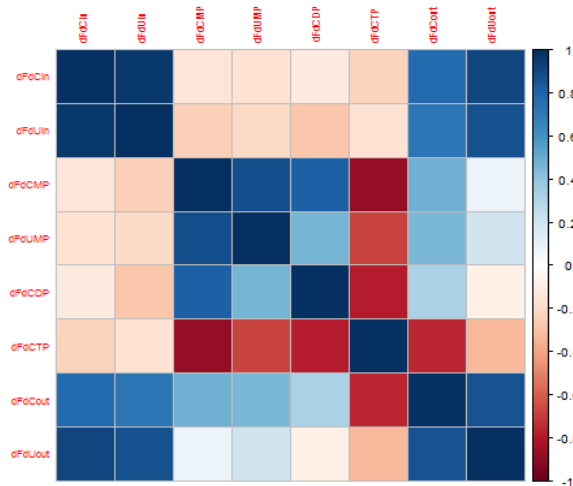


Network inference

P. Lecca, P. Nguyen, C. Priami, P. Quaglia. Network inference from Time-Dependent Omics Data, Bioinformatics for Omics data: Methods and Protocols in Molecular Biology Springer Science+Business Media, LLC 2011, 2011

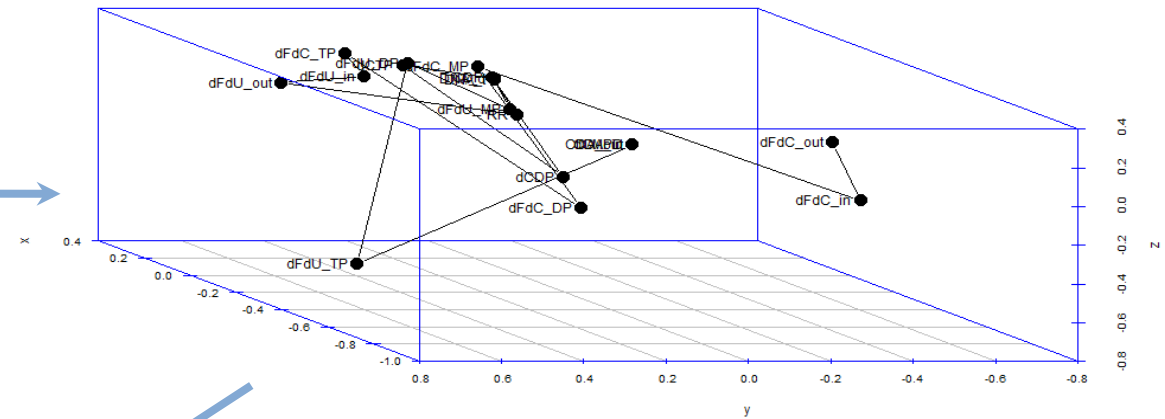
NETWORK INFERENCE

Lagged correlations

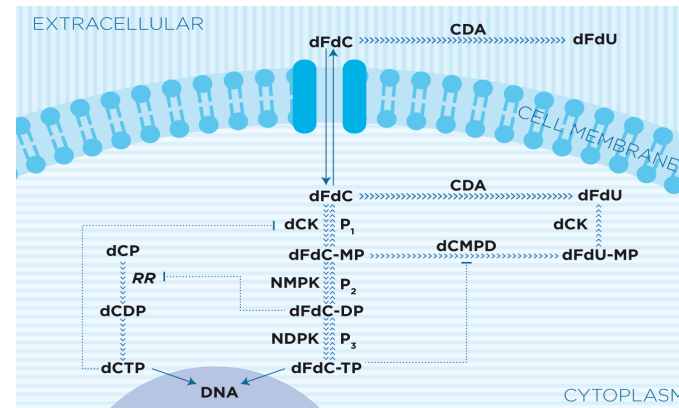


Distance clustering

Multidimensional Scaling 3-D plot of gemcitabine metabolites



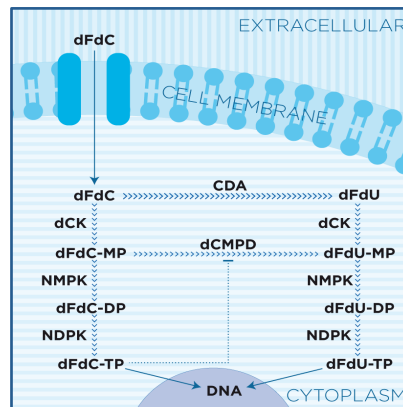
Network calibration



IDENTIFICATION OF METABOLIC NETWORKS

Gemcitabine metabolism: input data from

Volker Heinemann, Yi-Zheng Xu, Sherri Chubb, Alina Sen, Larry W. Hertel, Gerald B. Grindey, and William Plunkett
 Cellular Elimination of 2',2'-Difluorodeoxycytidine 5'-Triphosphate: A Mechanism of Self-Potential
 CANCER RESEARCH 52, 533-539, February 1, 1992

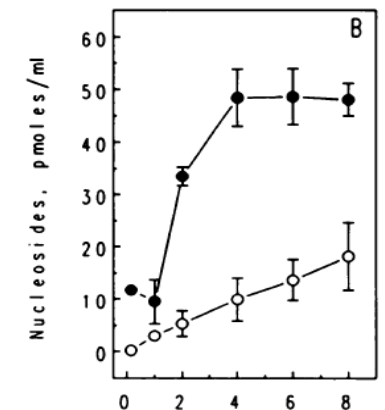
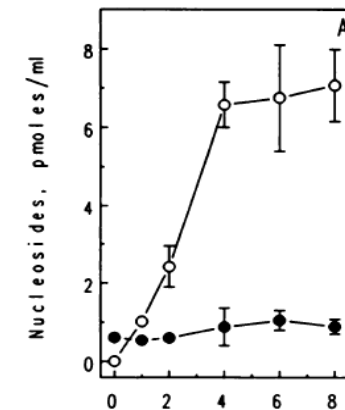
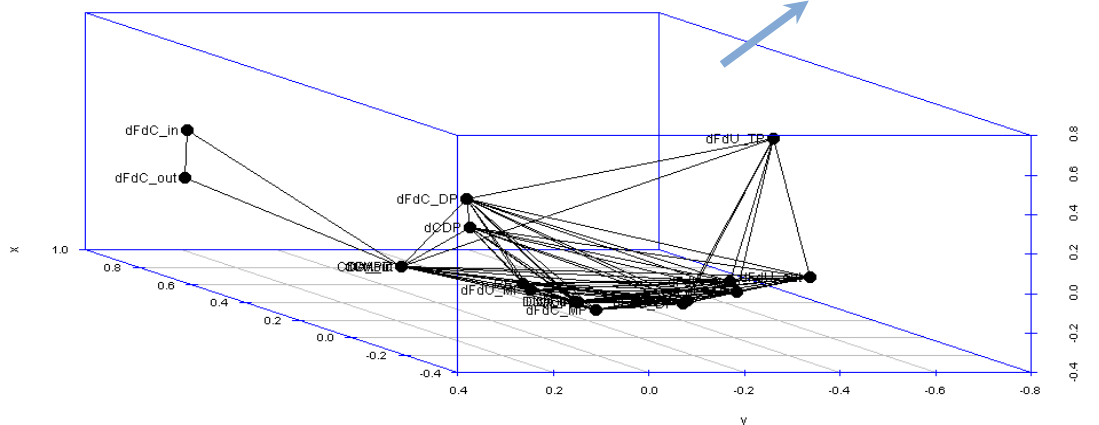


CEM cells were incubated for 2 h with either 10 μM [^{14}C]dFdC or 0.1 μM [^{14}C]dFdC. The cellular dFdCTP levels were 464 μM and 38 μM , respectively. Cells were then washed into fresh medium and portions of each culture were extracted at the indicated times and analyzed for intracellular dFdC metabolites.

	% of total metabolites				
	0 h	2 h	4 h	6 h	8 h
10 μM dFdC metabolite					
dFdC	0.3	0.6	0.6	1.2	
dFdU	0.4	0.8	1.4	4.4	3.6
dFdCMP	1.4	3.1	4.2	1.9	2.2
dFdUMP	15.3	15.7	17.7	15.2	16.0
dFdCDP	1.8	6.1	4.9	3.0	3.0
dFdCTP	80.9	73.6	71.2	74.3	76.3
0.1 μM dFdC metabolite					
dFdC	ND ^a	ND	ND	ND	ND
dFdU	0.9	1.4	3.2	4.9	8.3
dFdCMP	0.4	1.4	2.4	3.4	2.4
dFdUMP	11.9	13.9	17.5	17.4	20.5
dFdCDP	2.3	2.9	3.4	2.4	1.6
dFdCTP	84.5	80.5	73.5	71.8	67.2

^a ND, not detected.

Multidimensional Scaling 3-D plot



MODEL PARAMETERS

The parameters are the specific speed of the following events:

- Degradation rate of gemcitabine (k_1)
- Drug inefficiency rate (k_2)
- Drug efficacy (i. e. constant-cell-kill rate) (k_3)
- Tumor growth rate (k_4)
- k_5 : injection is modeled as single instantaneous event having infinite rate constant

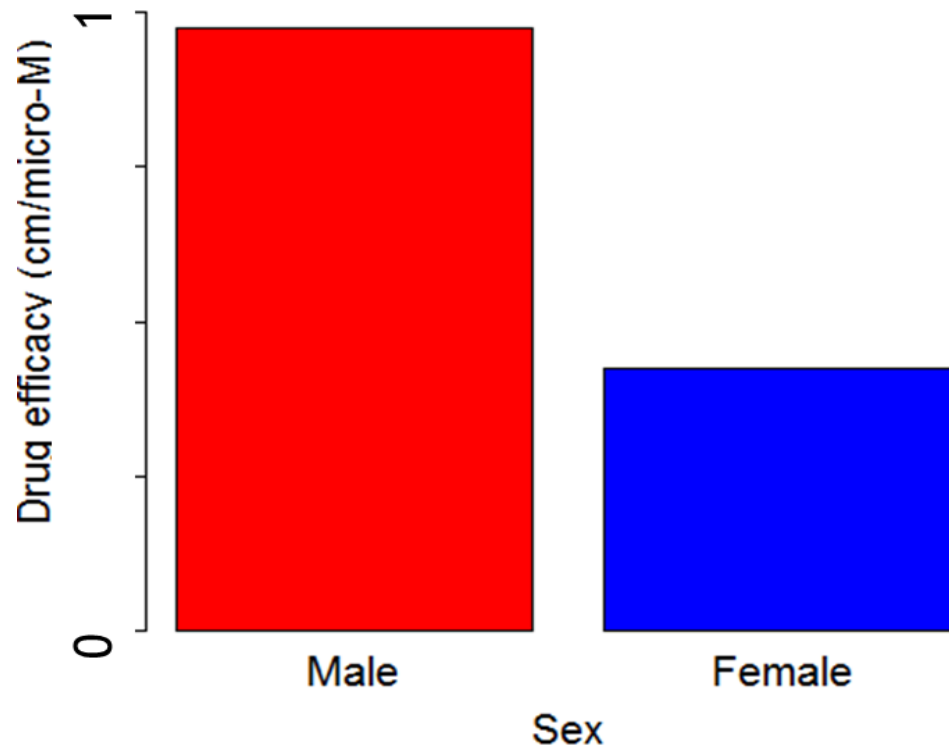
METHODS AND DATA

- We used KInfer on the time courses of tumor shrinkage and gemcitabine dosage on 56 patients to infer the parameters of the model.
- Patients have been divided with respect to sex, age and smoke history.
- The experimental curves of tumor shrinkage have been provided by R. A. Soo of the Department of Hematology-Oncology, National University Hospital, Singapore.

DRUG EFFICACY AND CLASSES OF PATIENTS

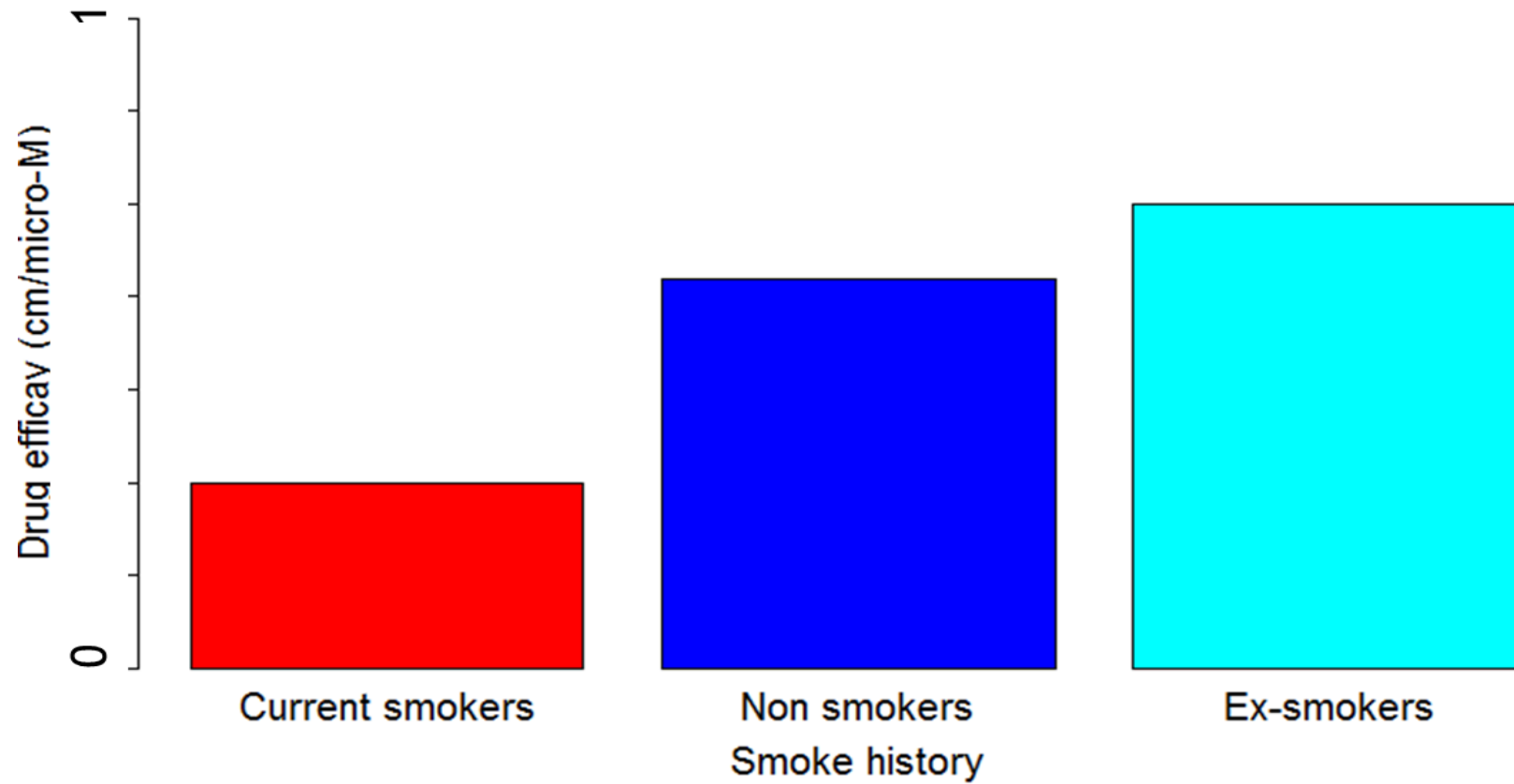
- We found that the drug efficacy (k_3) is correlated to the patient sex, smoke history, and tumor stage.
- No correlation seems to exist between drug efficacy and patient age.

EFFICACY AND GENDER



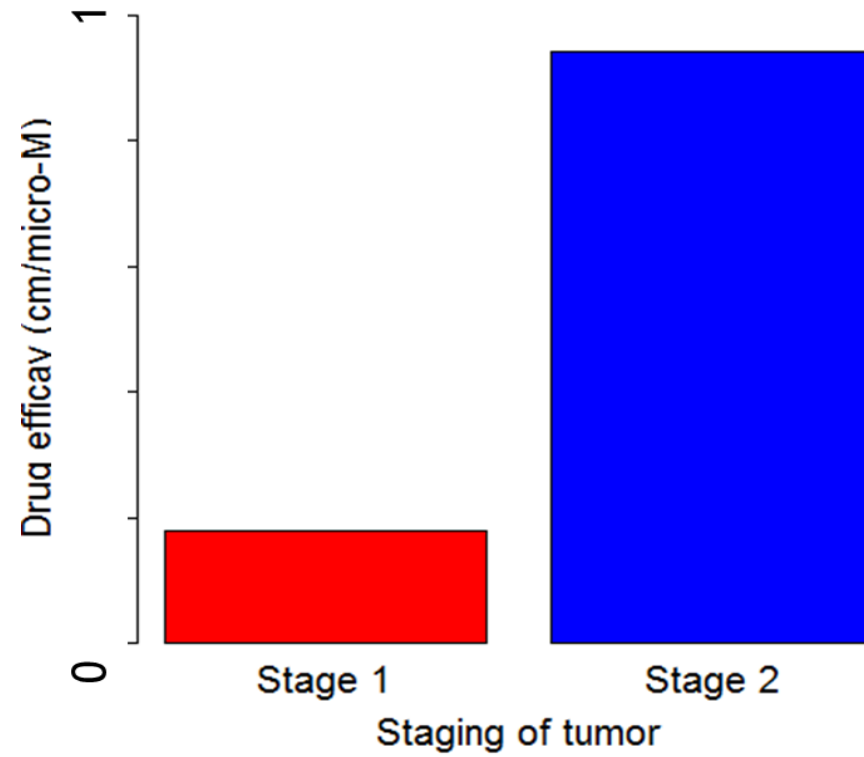
Normalized between 0 and 1

EFFICACY AND SMOKE HISTORY

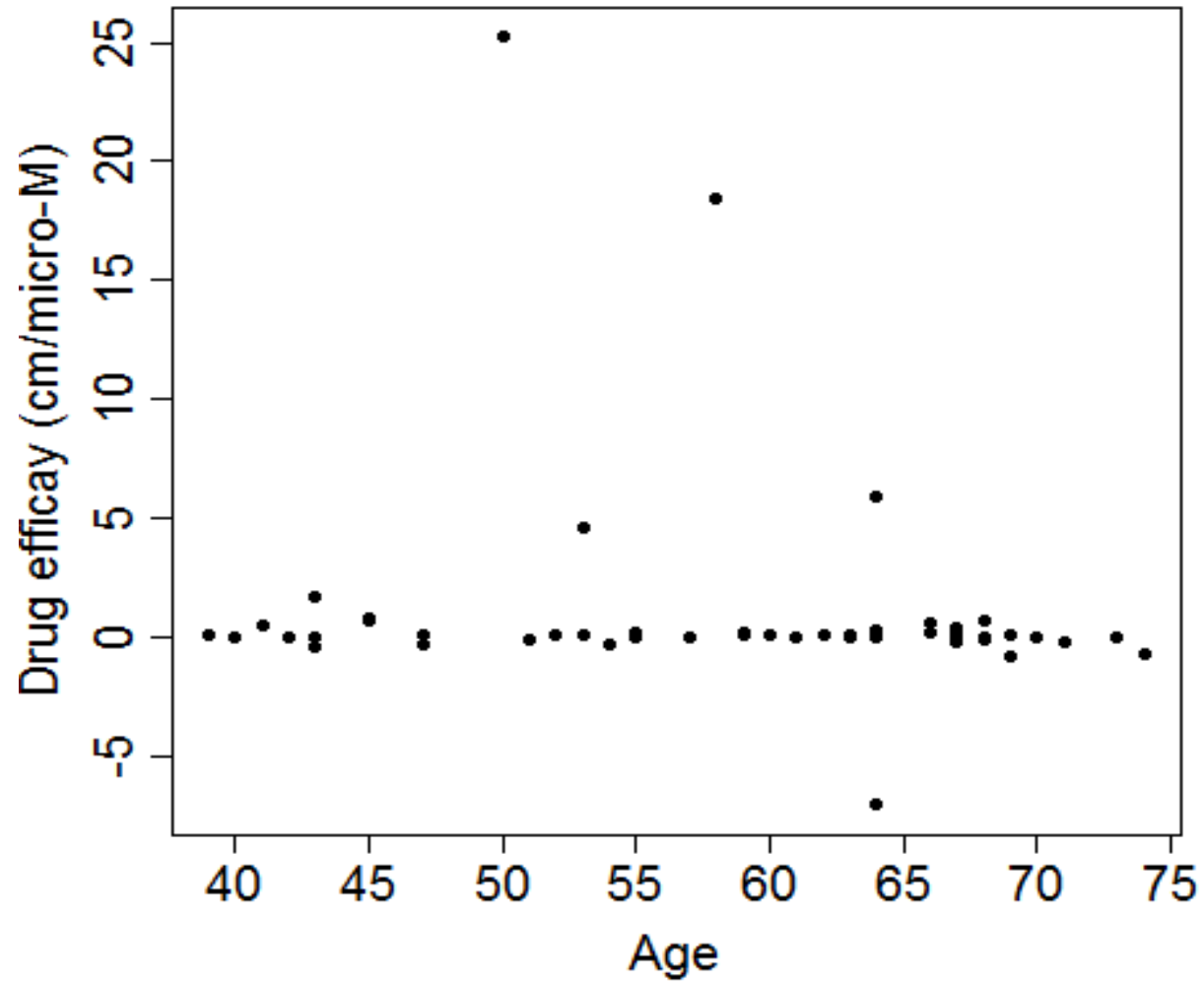


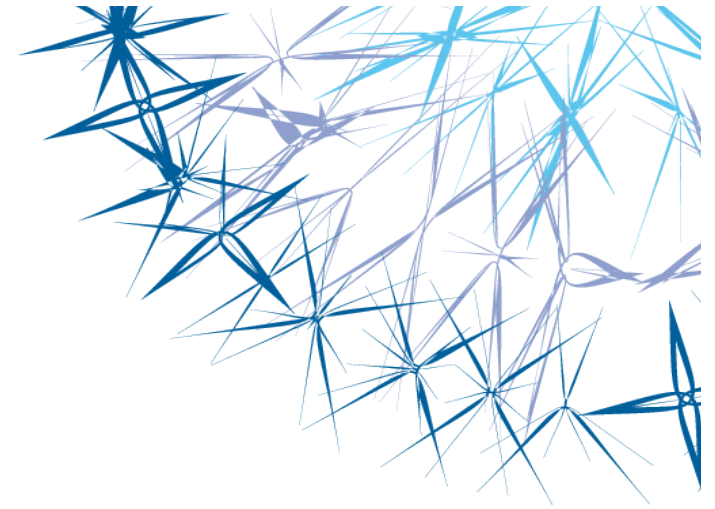
Normalized between 0 and 1

EFFICACY AND TUMOR STAGE



DRUG EFFICACY VS PATIENT AGE





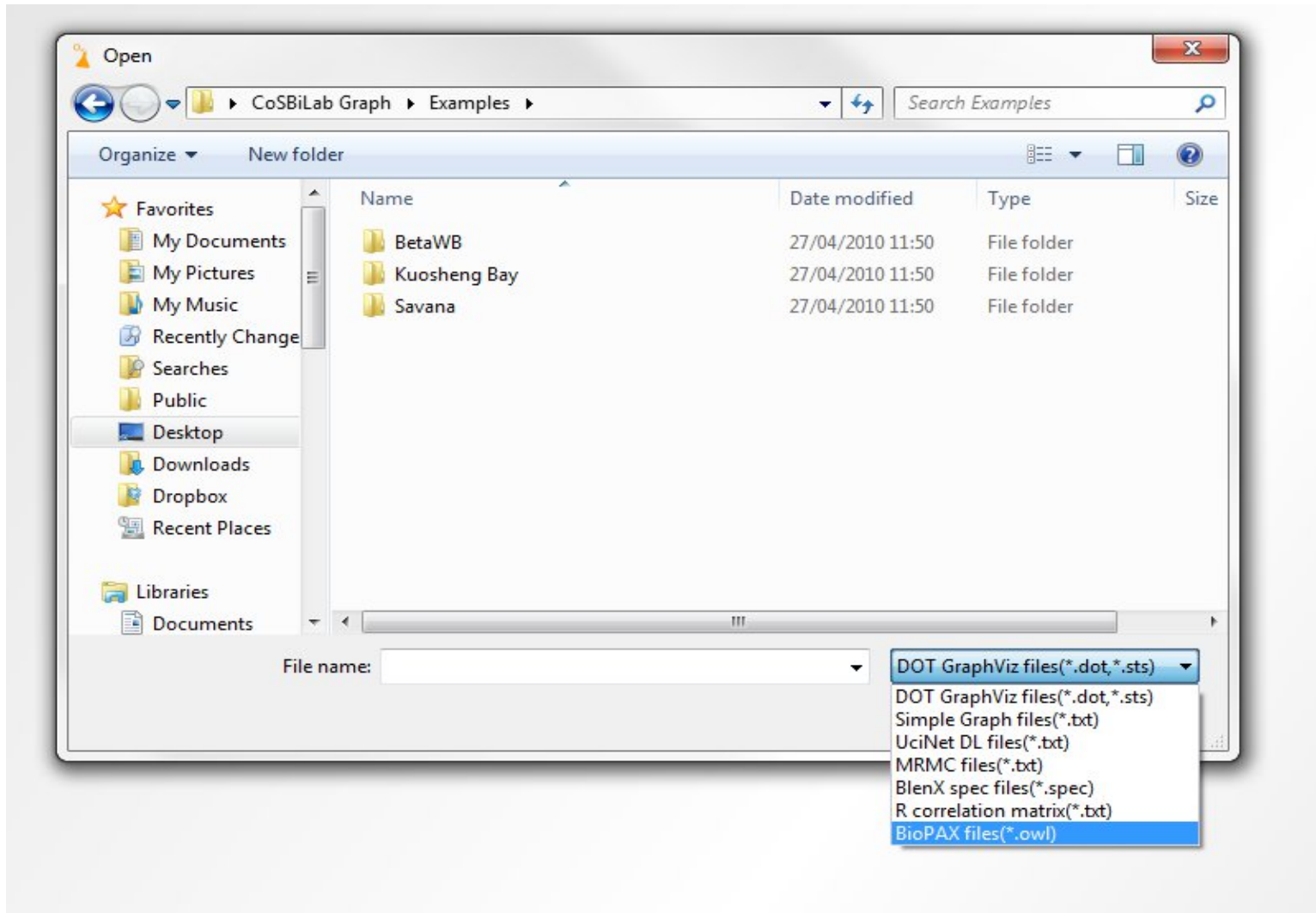
NETWORK Analysis and visualization



R. Valentini, F. Jordan. *CoSBI Lab Graph: the network analysis module of CoSBI Lab*. **Environmental Modelling and Software**, 25:886-888, 2010.

Download:
<http://www.cosbi.eu/index.php/research/prototypes/graph>

INPUT FORMATS



GRAPH IMPORTED

The screenshot displays the CoSbiLab Graph application interface. The main window shows a circular graph with 50 nodes and 100 edges. The nodes are arranged in a circle, and the edges connect them in a complex, dense pattern. The interface includes a menu bar (File, Graph, Selection, Windows, Tools, Layout, Actions, Help) and a toolbar with various icons for file operations and graph manipulation. A Properties Inspector panel on the right provides details about the graph, including its name (Graph1), node count (50), and edge count (100). Below the Properties Inspector is a Bookmarks panel.

CoSbiLab Graph - v.1.5 Dev

File Graph Selection Windows Tools Layout Actions Help

Graph1

Properties Inspector

Graph Graph1

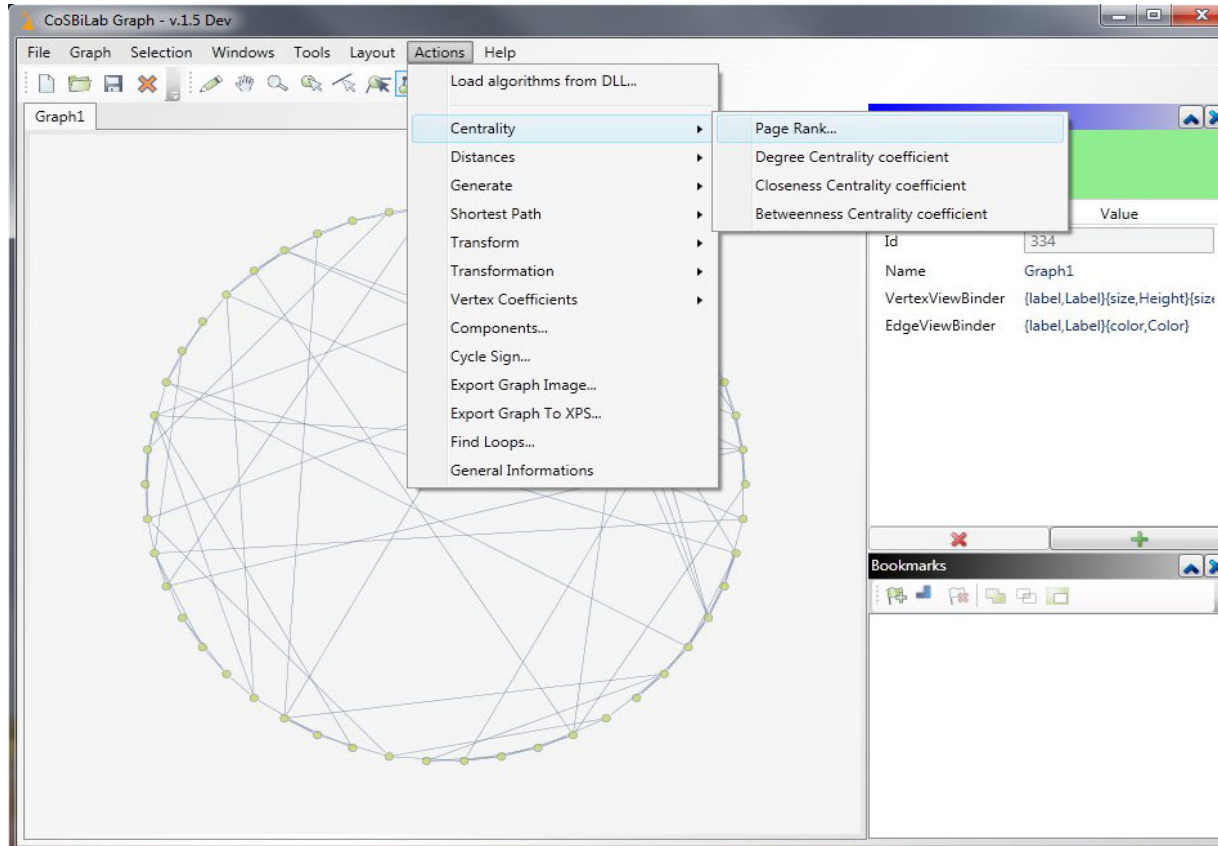
Nodes: 50

Edges: 100

Name	Value
Id	334
Name	Graph1
VertexViewBinder	{label,Label}{size,Height}{siz
EdgeViewBinder	{label,Label}{color,Color}

Bookmarks

ALGORITHMS FOR NETWORK ANALYSIS



Centrality:

Page Rank

Degree Centrality coefficient

Closeness Centrality coefficient

Distances:

Diameter

Compactness Index

Center of Gravity

Average Distance

Generate:

Watts-Strogatz Random Graph

Erdos-Reyni Random Graph

Barabasi-Albert Random Graph

Shortest Path:

Shortest Path

Shortest Path Matrix

Dijkstra Shortest Path

BellmanFord Shortest Path

Vertex Coefficients:

Status, Contrastatus, Netstatus

K-Index

Degree

Clustering Coefficients

LAYOUTS: FRUCHTERMAN REINGOLD

The screenshot displays the CoSbiLab Graph - v.1.5 Dev application interface. The main window shows a graph visualization with a circular layout. A menu is open, listing various layout options: Random, Balloon Tree, Tree, Fruchterman, Spring, Grid, H Sinusoid, V Sinusoid, Spiral, and Circle. The Fruchterman layout is currently selected. The graph visualization shows a complex network of nodes and edges, with a circular arrangement of nodes on the left side. A Properties Inspector panel is visible on the right, showing the Selection tab with the following information:

Selection
Number of vertices: 5
Number of edges: 0
Selected Vertices
Selected Edges

Name	Value
PageRank	Different values

Below the Properties Inspector, there is a Bookmarks panel showing a search result: "=PageRank>1.4".

PROPERTY INSPECTOR

The screenshot displays the CoSBI Lab Graph software interface. The main window, titled "CoSBI Lab Graph - v.1.5 Dev", features a menu bar with "File", "Graph", "Selection", "Windows", "Tools", "Layout", "Actions", and "Help". Below the menu is a toolbar with various icons for file operations and graph manipulation, including a search icon. The central workspace, labeled "Graph1", shows a network graph with numerous nodes and edges. One node is highlighted in blue, and its incident edges are also highlighted in blue. To the right of the graph is the "Properties Inspector" panel, which displays the following information:

Node
Size: (20, 20)
Location: (672, -374)
Degree: 5
Incident Edges

Name	Value
Id	372
Name	
PageRank	1.31

Below the Properties Inspector is a "Bookmarks" panel, which is currently empty. The interface includes standard window controls (minimize, maximize, close) and a search bar.

SEARCH: PAGE RANK > 1.4

The screenshot displays the CoSBI Lab Graph - v.1.5 Dev application window. The main area shows a network graph with numerous nodes and edges. Five nodes are highlighted in green, representing the search results for the query "=PageRank>1.4". The search results are also visible in the Properties Inspector and Bookmarks panels on the right.

Properties Inspector

Selection

Number of vertices: 5
Number of edges: 0

- Selected Vertices
- Selected Edges

Name	Value
PageRank	Different values

Bookmarks

Search result: "=PageRank>1.4"

SHORTEST PATH TOOL

The screenshot displays the CoSBI Lab Graph - v.1.5 Dev application. The main window shows a network graph with numerous nodes and edges. A specific path of four edges is highlighted in green, representing the shortest path. The interface includes a menu bar (File, Graph, Selection, Windows, Tools, Layout, Actions, Help) and a toolbar with various icons. A search filter is applied: `=PageRank>1.4`. On the right side, there are two panels: the Properties Inspector and the Bookmarks panel.

Properties Inspector

Selection

Number of vertices: 2
Number of edges: 4

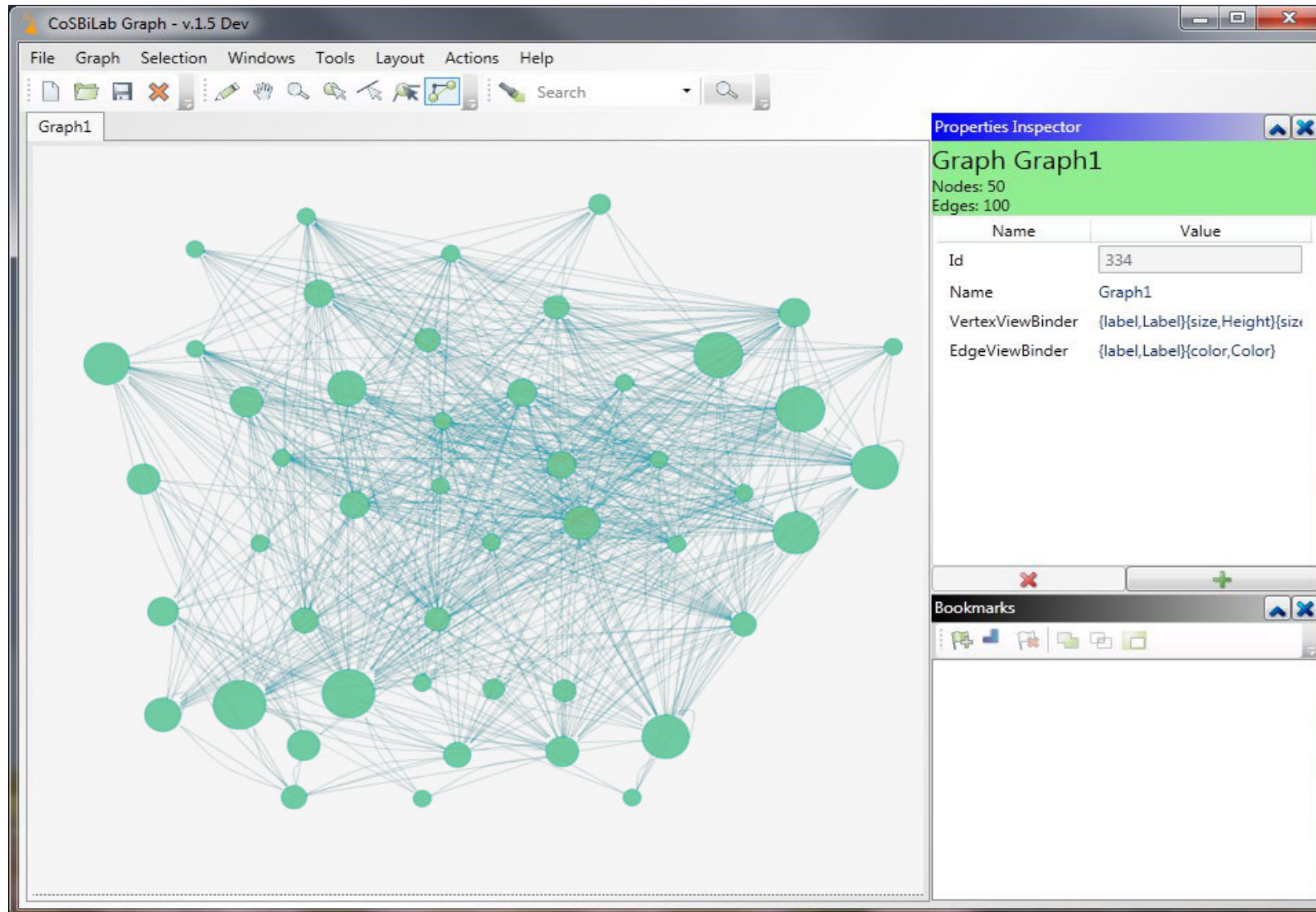
- Selected Vertices
- Selected Edges

Name	Value
PageRank	Different values

Bookmarks

Search result: `"=PageRank>1.4"`

NODE SIZE REFLECTING A NODE PROPERTY



PICTURES AS NODE ATTRIBUTES

The screenshot displays the CoSBI Lab Graph software interface. The main window shows a network graph with 50 nodes and 100 edges. Each node is represented by a small image, such as a fish, a bird, or a turtle, illustrating the concept of using pictures as node attributes. The interface includes a menu bar (File, Graph, Selection, Windows, Tools, Layout, Actions, Help) and a toolbar with various icons for graph manipulation. On the right side, there is a Properties Inspector panel for 'Graph Graph1' and a Bookmarks panel.

Properties Inspector

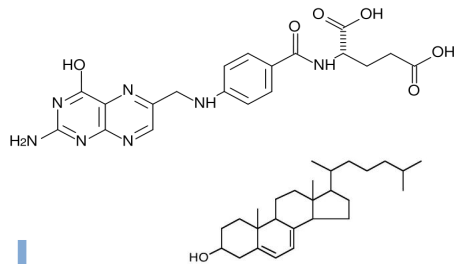
Graph Graph1
Nodes: 50
Edges: 100

Name	Value
Id	334
Name	Graph1
VertexViewBinder	{label,Label}{size,Height}{size
EdgeViewBinder	{label,Label}{color,Color}

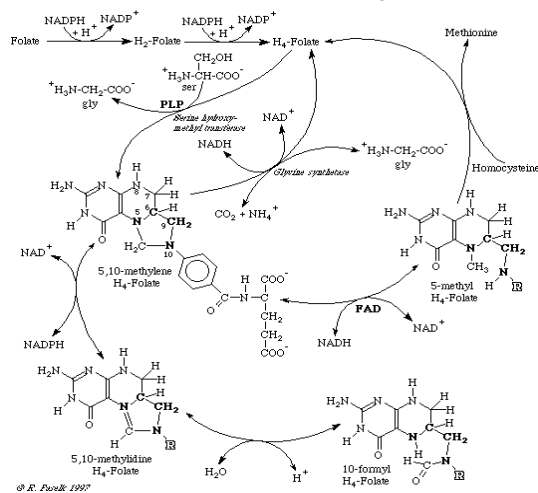
Bookmarks

VITAMINS AND OBESITY

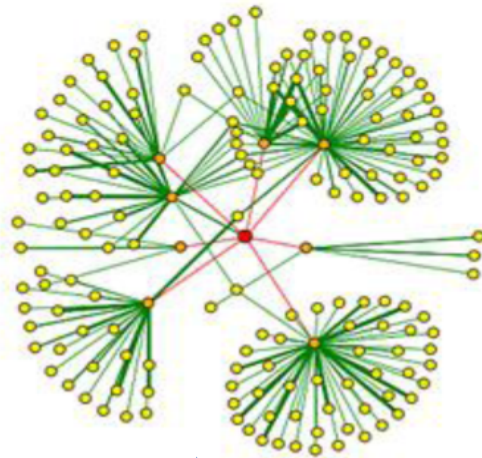
Select the micronutrients



Search Networks & Pathways



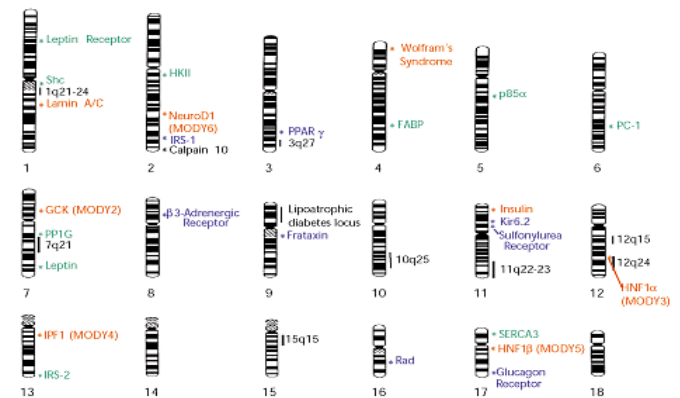
Search & Analyse Protein - Protein Interaction Database



Select protein At Key Nodes

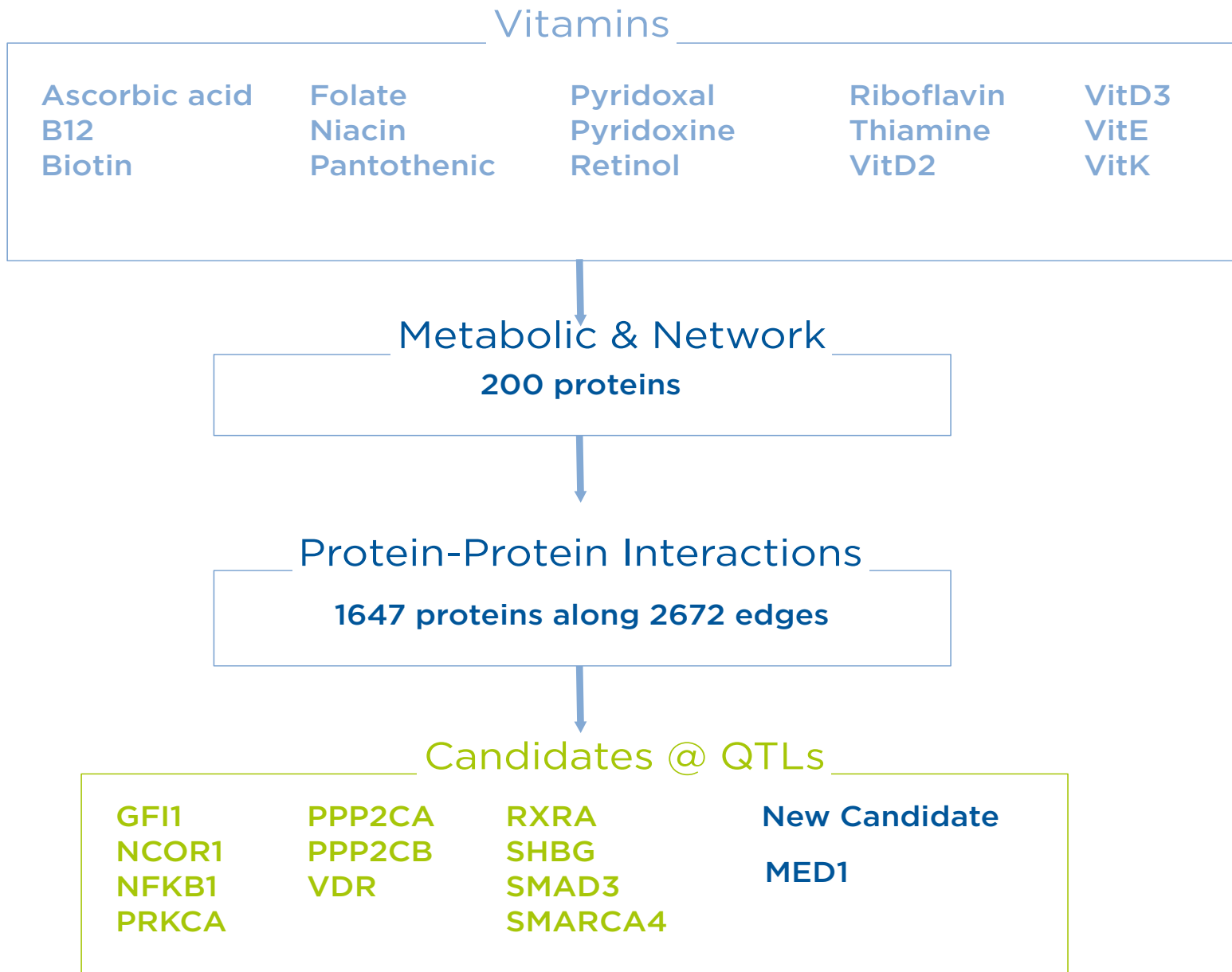
Map to QTL of Obesity/diabetes Phenotype

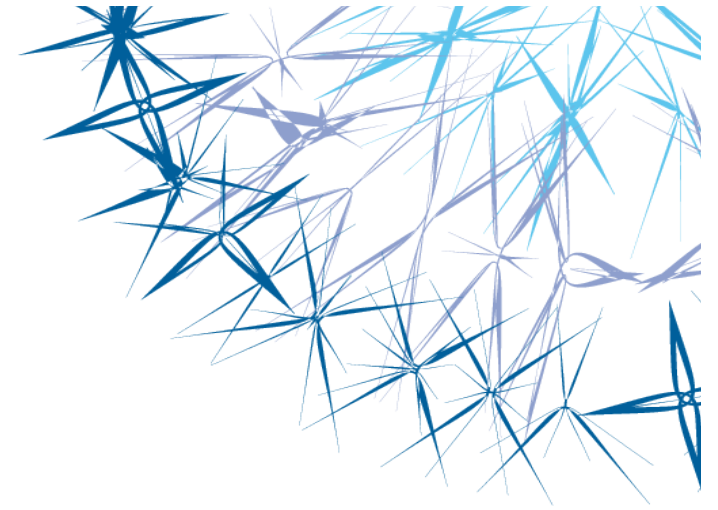
Select genes involved and proteins codified



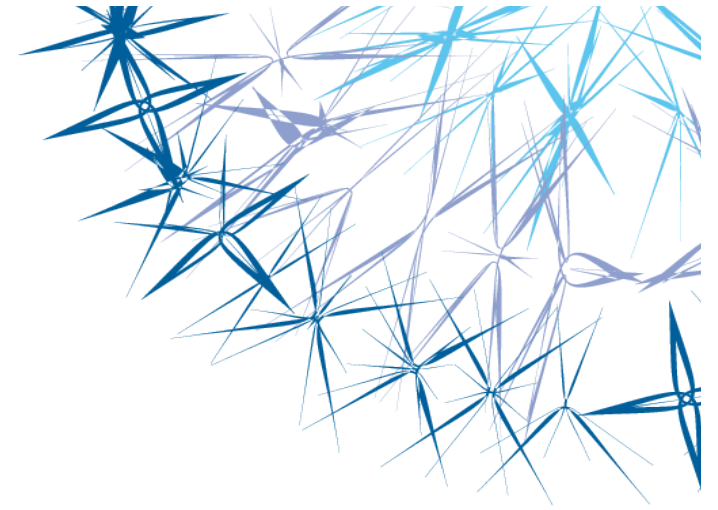
CANDIDATE GENES

VITAMINS AND OBESITY





CONCLUSIONS



www.cosbi.eu



priami@cosbi.eu